# **Annex XV report**

# PROPOSAL FOR IDENTIFICATION OF A SUBSTANCE OF VERY HIGH CONCERN ON THE BASIS OF THE CRITERIA SET OUT IN REACH ARTICLE 57

**Substance Name(s):** 4,4'-isopropylidenediphenol (Bisphenol A)

**EC Number:** 201-245-8 **CAS Number:** 80-05-7

Submitted by: France

**Date:** 02/03/2017

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# List of abbreviations

11-bHSD1: 11β-hydroxysteroid dehydrogenase type 1

3D: Three-dimensional

3T2-L1: cell line derived from mouse cells, having a fibroblast-like morphology and under appropriate conditions, differentiate into an adipocyte-like phenotype

4-OHT: 4-hydroxy tamoxifen, active metabolite of tamoxifen.

5-HIAA: 5-Hydroxyindoleacetic acid (main metabolite of serotonine)

5-HT: 5-hydroxytryptamine or serotonin

5a-R: 5a-reductase AB: Alveolar bud

Abl1: v-abl Abelson murine leukemia viral oncogene homolog 1

ABS: acrylonitrile-butadiene-styrene

ACC: acetyl-CoA carboxylase AChE: acetylcholinesterase ADH: Atypical Ductal Hyperplasia

ADHD: attention deficit hyperactivity disorder

AFSSA: French food safety agency (became ANSES in 2010)

AhR: aromatic hydrocarbon receptor

AIST: Japanese National Institute of Advanced Industrial Science and Technology

Akt: protein kinase B (PKB) mRNA

AMPA receptor: a-amino-3-hydroxy-5-methyl-4-isoxazolepropionic acid receptor

ANIA: french association for food industry

ANSES: Agence Nationale de Sécurité Sanitaire de l'alimentation, de l'environnement et du travail (French Agency for Food, Environmental and Occupational Health & Safety)

AOP: adverse outcome pathway

AR: androgen receptor ARC: arcuate nucleus AREG: amphiregulin

ART: assisted reproductive technology ATGL: adipose triglyceride lipase ATP: Adaptation to Technical Progress

AUC: area under the curve

AVPV: anteroventral periventricular nucleus

BADGE: bisphenol A diglycidyl ether

BAT: brown adipose tissue BBP: Butyl benzyl phthalate

BDNF: brain-derived neurotrophic factor BDP: bisphenol A bis (diphenyl phosphate) Bis-DMA: bisphenol A demethacrylate

BM: basal medium BMI: body mass index

BMP-15: bone morphogenic protein 15

BPA: bisphenol A

BrdU: Bromodeoxyuridine

BSEF: international bromine production organisation

bw: body weight

C/EBPa: CCAAT/enhancer binding proteins *alpha*, a protein shown to bind to the promoter and modulate the expression of the gene encoding leptin

CA: cornu ammonis (area of the hippocampus)

CAF: cancer-associated fibroblasts

CALUX: Chemical Activated LUciferase gene eXpression

CaM: calmodulin

CamKI: calcium/calmodulin-dependent protein kinase type I

CamKIV: calcium/calmodulin-dependent protein kinase type IV

CaMKK: calcium/calmodulin kinase kinase

CARACAL: Competent Authorities for REACH and CLP

CAT: catalase

CBP: CREB-binding proteinCBX: carbenoxolone, inhibitor of 11b-HSD

CD: normal Chow Diet

Ccnd2: cyclin D (cell -cycle activator)

CDK: cyclin dependent kinase; Cdk4 binds Ccnd2 (cell -cycle activator)

CETIM: technical centre for mechanical industries (France)

CHAMACOS: Center for the Health Assessment of Mothers and Children of Salinas study

CHDM: cyclohexanedimethanol

CI: confidence interval CIS: Carcinoma in-situ CKD: chronic kidney disease

CL: corpora lutea

CLH: harmonised classification and labelling CLP: Classification, Labelling and Packaging CMR: Carcinogen, Mutagen, Reprotoxic

CNS: central nervous system

COMIDENT: coordinating committee of dental activites (France)

CREB: cAMP response element-binding protein Crtc1: CREB regulated transcription coactivator 1

CSR: chemical safety report

CYP45017a: 17a-hydroxylase-17,20-desmolase (also named CYP17A1) CYP450arom: cytochrome P450 aromatase (also named CYP19A1)

CYP450scc: P450 cholesterol side chain cleavage enzyme (also named CYP11A1)

DA: dopamine

DAT: dopamine transporter dBPA: deuterated BPA

DCIS: Ductal Carcinoma in situ

DES: diethylstilbestrol DEX: dexamethasone DG: dendate gyrus

DHEA: dehydroepiandrosterone

DiE-1: diestrus-1

DIN: Ductal Intra-Epithelial Neoplasia DMBA: dimethylbenzanthracene DMSO dimethyl sulfoxide DMT: dimethyl terephthalate DNA: deoxyribonucleic acid DNEL: Derived no effect level DNMT: DNA methyl transferase

DOHaD: developmental origin of health and disease

DOPAC: 3,4-Dihydroxyphenylacetic acid (metabolite of dopamine)

DPN: diarylpropionitrile (ERβ antagonist)

DS: dossier submitter E2: 17-β estradiol EB: estradiol benzoate EC: European Commission

ECHA: European Chemical Agency

ECM: Extracellular matrix ECN: embryo cell number ED: endocrine disruptor

EDC: endocrine disruptor chemical

EE2: ethinylestradiol

EFS: embryo fragmentation score EFSA: European Food Safety Agency

EGF: Epidermal Growth Factor

EGPF: enhanced green fluorescent protein

ELF-5: E74-like factor 5

ELoC: equivalent level of concern EMT: epithelia-mesenchymal transition

ENU: N-ethyl-N-nitrosourea EPA: eicosapentaenoic acid ER: estrogen receptor

ErbB4: v-erb-a erythroblastic leukemia viral oncogenehomolog 4

EREs: estrogen response elements

ERK1: extracellular-signal-regulated kinase 1

ERK/MAPK: extracellular signal-regulated kinase/ mitogen-activated protein kinase

ERR: estrogen related receptor

ERRy or ERRgamma: Estrogen-related receptor gamma

Era or ERalpha: estrogen receptor a ERβ or ERbeta: estrogen receptor beta

Ets: v-ets erythroblastosis virus E26 oncogene

EU: European Union

EU28: European Union with its 28 member states (since 2013)

EuPC: European plastic converters

EZH2: Enhancer of Zeste Homolog 2, a histone methyltransferase that has been linked to breast cancer risk and epigenetic regulation of tumorigenesis.

EZH2: Enhancer of Zeste Homolog 2 FABP4: fatty acid binding protein 4

FAO: food and agriculture organization of the United Nations

FAS: Fatty acid synthase

FDA: US Food and Drug Administration

FN: fibronectin

FOPH: Swiss federal health authority FSH: follicle stimulating hormone FVB: Friend leukemia virus B G15: specific GPER inhibitor

GAA1: glycosyl phosphatidyl-inositol anchor attachment protein

GABA: gamma-aminobutyric acid, main inhibitory neurotransmitter in the central nervous system

GAS: q-interferon activation sequence

GATA4: a transcription factor characterised by its ability to bind to the DNA sequence "GATA"

GD: gestation day

GDF-9: growth/differentiation factor 9 GDM: gestational diabetes mellitus

GH: Growth hormone

Glu: glutamate GLUT4: Glucose transporter type 4

GnRH: gonadotrophin-releasing hormone

GPER or GPR30: G protein-coupled estrogen receptor or G protein-coupled receptor 30, the membranous form of estrogen receptor

GPER-1: G protein-coupled estrogen receptor-11.

GR: glucocorticoid receptor (NR3C1) GRF: quanine nucleotide-releasing factor

GSH: Glutathione

<sup>&</sup>lt;sup>1</sup> coordinates fibronectin (FN) matrix assembly and release of heparin-bound epidermal growth factor (HB-EGF). This mechanism of action results in the recruitment of FN-engaged integrin α5β1 to fibrillar adhesions and the formation of integrin  $a5\beta1$ -Shc adaptor protein complexes

GSIS: Glucose-Stimulated Insulin Secretion GSK2beta: glycogen synthase kinase 3 beta

GTT: glucose tolerance test

GW9662: selective PPARy antagonist HBEC: human breast epithelial cell

HB-EGF: heparan binding epidermal growth factor HDL-C: high density lipoprotein – cholesterol HEAL: Health and Environment Alliance (NGO)

hESC: Human embryonic stem cells

HFD: High Fat Diet

HGFL: hepatocyte growth factor-like protein HMEC: Human mammary epithelial cells

HOMA: homeostasic model assessment of insulin resistance

HOTAIR: HOX transcript antisense RNA

HOX genes: a subset of homeotic genes that control the body plan of an embryo

HPG: hypothalamic-pituitary-gonadal axis

HRG1: heregulin 1

HSD3β: 3β-hydroxysteroid dehydrogenase

HSL: hormone-sensitive lipase IAPP: Islet Amyloid PolyPeptide

ICC: islet cells cluster

ICI 182,780: specific inhibitor of ERa/ $\beta^2$  ICR: Institute of Cancer Research

ICSI: intra-cytoplasmic sperm injection (ICSI)

ICV: intracerebroventricular ID2: inhibitor of DNA binding 2

iDMEC: induced differentiated mammary epithelial cell

IGF-1: insulin-like growth factor-1 IGF-2: insulin-like growth factor 2 IGT: impaired glucose tolerance

IKKa: Inhibitor of kappa B kinase alpha

INERIS: French institute for environment and industrial risks

INS-1 Cells: pancreatic beta cells line

 $\label{local_continuity} \begin{tabular}{ll} IOELV: indicative occupational exposure limit values ipGTT: intraperitoneal glucose tolerance tests \\ \end{tabular}$ 

ipITT: intraperitoneal insulin tolerance tests

IQ: intelligence quotientIR: insulin receptor

IRS-1: insulin receptor substrate 1

IS: insulin sensitivity IVF: in vitro fertilisation JAK2: janus 2 kinase

JRC: Joint Research Center, the European Commission's in-house science service

KATP: ATP-sensitive potassium channel (or KATP channel)

KCC2: potassium chloride cotransporter 2

KEWI-WISC: Korean Educational Development Institute's Weschler Intelligence Scales for Children

KGN: human cell line from ovarian granulosa cell tumor

Kiss1R: kisspeptin receptor LBD: ligand binding domain

LDES: Learning Disability Evaluation Scale LDL-C: low density lipoprotein -cholesterol Ldlr: Low density lipoprotein receptor

<sup>&</sup>lt;sup>2</sup> called also faslodex (Wakeling, 1994) acts in an anti-estrogenic manner in all tissues analysed (Dowsett *et al.*, 1995 – Labrie *et al.*, 1999 – Sibonga *et al.*, 1998).

LEF1: transcription factor of the HMG-box family<sup>3</sup>

LH: luteinising hormone

LIN: Lobular Intra-Epithelial Neoplasia

LMO4: LIM domain only 4

LN: lymph node

LOAEL: lowest observed adverse effect level

LPL: lipoprotein lipase

LRH1: liver receptor homolog 1
LTD: long-term depression
MAPK: mitogen-activated protein

MAPK: mitogen-activated protein kinase MAZE test: an appetite-motivated maze test MCF: human breast epithelial cell lines

MDA: Malondialdehyde

MDI: induction medium containing methylisobutylxanthine, dexamethasone, insulin; used in protocols to induce differentiation of 3T3-L1 cells into adipocyte-like cells

MEK: mitogen-activated protein kinase mGluR: metabotropic glutamate receptor MMTV: mouse mammary tumor virus

MoA: mode of action MoE: margin of exposure

mPFC: median prefrontal cortex (area of the hippocampus)

MR: mineralocorticoid receptor (NR3C2) mRNA: messenger ribonucleic acid MSC: member state committee MSCs: Mesenchymal Stromal Cells

MSCA: McCarthy scales of children's abilities

MWM: Morris water maze NAC: N-acetylcystein

NCTR: US National Center for Toxicological Research

ND: no data or not determined

NE: norepinephrine

NEFA: Non-Esterified Fatty Acids

NF-κβ: nuclear factor κβ
NGO: non-profit organisation

NHANES: US National Health and Nutrition Examination Survey

NHS: nurses' health study NHSII: nurses' health study II

Nlgn3: neuroligin 3

NMDA: N-methyl-D-aspartate NMDAR: NMDA receptor NMU: N-nitroso N-methylurea

NOAEL: no observed adverse effect level

NR: mineralocorticoid receptor

Nrxn1: neurexin 1

NTP: US National Toxicology Program

Nur77: nuclear receptor of the Nur family that act as transcription factors in neuron development and maintenance; also known as nerve growth factor IB (NGFIB)

OEL: occupational exposure limit

OP: object placementOR: object recognition

ORa: odds ratio

OVLT: organum vasculosum, lamina terminalis

OVX: ovariectomised

<sup>&</sup>lt;sup>3</sup> it is activated early in all ectodermal rudiments (including hair, whiskers, mammary glands, and teeth). In its absence, those organs fail to develop.

P16: sequesters Cdk4 and prevent its interactions woth D cyclins acting as an effector of cycle arrest and cellular senescence

PCNA: proliferating cell nuclear antigen

PC: polycarbonate

PCOS: polycystic ovary syndrome

PCSK1: gene encoding for proprotein convertase 1, an enzyme acting in the process of proinsulin

and proglucagon in pancreatic islets

PD 98059: inhibitor of ERK1/2.

PE: proestrus

PET: polyethylene terephthalate

PFC: prefrontal cortex

PGF2a: prostaglandin F2-alpha PI3K: phosphoinositide 3-kinase PKA: cAMP-dependent protein kinase

PKB: protein kinase B

PL: prolactin

PND: postnatal day PNW: postnatal week

PPARy: Peroxisome Proliferator Activated Receptor gamma

PPRE: PPARy Response Element

PPT: propyl pyrazoletriol (ERa agonist)

PPT-A: preprotachykinin A<sup>4</sup> PR: progesterone receptor

PRC: polycomb repressive complex

PRL: prolactin

PrlR: prolactin receptor PSD: post synaptic density

PTHrP: Parathyroid hormone-related peptide<sup>5</sup>

PVC: polyvinyl chloride PWD: Post Week Day PXR: pregnane X receptor

RAC: Risk Assessment Committee

RAL: raloxifene<sup>6</sup>

RANKL: receptor activator of NFkB ligand

RCR: risk charaterisation ratio

REACH: Registration, Evaluation, Authorisation and Restriction of Chemicals

RFRP-3: RFamide-related peptide-3 RiME: Risk Management Expert meeting

RISS: Japanse research Institute of Science for Safety and Sustainability RIVM: Netherlands National Institute for Public Health and the Environment

RMM: risk management measures ROS: Reactive Oxygen Species

ROSI: Rosiglitazone

RP3V: rostral periventricular area of the third ventricle

RT-qPCR: reverse transcription quantitative polymerase chain reaction

RU-486: mifepristone, used as an antagonist to glucocortocoid

RXR: retinoid X receptor SC: subcutaneous route

<sup>&</sup>lt;sup>4</sup> PPT-A gene products lead to Substance P and neurokinin A that are members of the mammalian tachykinin peptide family, currently referred to as neurokinins (1456). The physiology of these peptides are linked to neurotransmission.

<sup>&</sup>lt;sup>5</sup> Inactivation of either the PTHrP peptide or the receptor prevents formation communication of androgen receptors and outgrowth of the primary sprout at E16 while it does not affect early bud formation.

<sup>&</sup>lt;sup>6</sup> Raloxifene acts in an agonistic manner on the bone (Turner *et al.*, 1994), but as an antagonist in the mammary gland (Buelke-Sam *et al.*, 1998).

SCD-1: stearoyl-CoA desaturase

SCENIHR: scientific committee on emerging ans newly identified health risks

SCOEL: Scientific Committee on Occupational Exposure Limits

SD: Sprague Dawley

SDN: sexual dimorphic nucleus

SE: standard error

SEAC: committee for socio-economic analysis SERM: Selective ER modulator (Dhingra, 1999). SF-1: steroidogenic factor 1 (transcription factor)

SMA: smooth muscle actin (marker of myoepithelial cells)

SNFBM: national association of manufacturers of packaging boxes and metal capping (France)

SNITEM: national association of medical technology industry (France)

SOCS: suppressor of cytokine signaling proteins

SOD: superoxide dismutase

SRC3 (AIB1): coactivator which plays a key role in transcriptional activation of several estrogenregulated genes<sup>7</sup>

SREBP-1C: sterol regulatory element binding protein-1C

SRp20: splicing factor arginine/serine-rich 3

ss: statistically significant

StAR: steroid acute regulatory protein

STAT: signal transducer and activator of transcription

STOT RE: specific target organ toxicity after repeated exposure (hazard class from CLP)

SVF: Stromal Vascular Fraction

SVHC: substance of very high concern T0070907: a potent PPAR $\gamma$  antagonist

Tamoxifen: anti-estrogen<sup>8</sup>
TAS: total antioxidant status
TBBPA: tetrabromobisphenol A

TBT: PPARγ agonists TD: Terminal Duct

TD2M: Type-2 diabetes mellitus TDI: tolerable daily intake

TDLU: Terminal Ductal Lobular Unit

TEB: Terminal End Bud

TFF1 (or PS2): estrogen-responsive gene trefoil factor

TG: triglyceride

TH: tyrosine hydroxylase

TMCD: 2,2,4,4-tetramethyl-1,3-cyclobutanediol

Tnc: tenascin

TNFa: Tumor Necrosis Factor alpha

TO: T0070907, used as a PPARy antagonist

ToxRTool: Toxicological data Reliability assessment Tool

TUNEL: terminal deoxynucleotidyl transferase-mediated deoxyuridine triphosphate nick end labeling

Ucp1: a marker of energy expenditure through thermogenesis

UDMA: urethane dimethacrylate

US EPA: US Environmental Protection Agency

VO: vaginal opening WAP: whey acidic protein WAT: White Adipose Tissue

<sup>7</sup> including PS2 gene (Shao *et al.*, 2004; Labhart *et al.*, 2005), followed the similar pattern as the ERa

<sup>&</sup>lt;sup>8</sup> Tamoxifen is widely used in the therapy of breast cancer (Gradishar et Jordan, 1997; Wakeling *et al.*, 1991), displays agonistic activity on the endometrium (Jordan et Morrow, 1994), but antagonistic activity on the mammary gland (Jaiyesimi *et al.*, 1995).

WB: Western blott

WHO: World Health Organization

WT: Wild Type

XIAP: x-linked inhibitor of apoptosis protein

# PROPOSAL FOR IDENTIFICATION OF A SUBSTANCE OF VERY HIGH CONCERN ON THE BASIS OF THE CRITERIA SET OUT IN REACH ARTICLE 57

**Substance Name(s):** 4,4'-isopropylidenediphenol (Bisphenol A)

**EC Number:** 201-245-8 **CAS number:** 80-05-7

• It is proposed to identify the substance as a substance of equivalent level of concern to those of other substances listed in points (a) to (e) of Article 57 of Regulation (EC) No 1907/2006 (REACH) according to Article 57(f) of the REACH Regulation.

# Summary of how the substance meets the criteria set out in Article 57 of the REACH Regulation

Bisphenol A is proposed to be identified as a substance of very high concern in accordance with Article 57(f) of Regulation (EC) 1907/2006 (REACH) because it is a substance with *endocrine disrupting properties* for which there is scientific evidence of probable serious effects to human health which gives rise to an equivalent level of concern to those of other substances listed in points (a) to (e) of Article 57 REACH.

BPA has been shown to affect the reproductive function, mammary gland development, cognitive function and metabolism through pathways that commonly involve disruption of estrogenic regulation. The effects on female reproductive function include the induction, after both developmental and adult exposures, of cystic ovaries, changes in the uterus morphology, alteration of fertility parameters as well as estrous cycle disturbance. The estrous cycle is a perfectly synchronised and sequenced event that relies on a permanent endocrine dialogue between the ovary and the hypothalamo-pituitary system. Those pathways differentiate during fetal life and are largely influenced by numerous factors and in particular the steroid environment of the foetus. BPA at the adult stage alters the endocrine steroidogenic function of the ovary and more specifically the production of estrogens by the follicle, leading to disturbance in the estrous cycle. At the neuroendocrine level, BPA can also act during the perinatal/postnatal organisation or adult activation of the hypothalamus-pituitary system through changes in kisspeptin, gonadotrophin-releasing hormone (GnRH) expression, activity or liberation and sex steroid receptor expression that impact estrous cyclicity.

The effects on the mammary gland, depending on the period of exposure, include: modifications in the mammary tissue such as an increased number of terminal end buds (TEBs) relative to the ductal area, fewer apoptotic TEB cells, increased lateral branching and ductal hyperplasia, increased cell proliferation and decreased apoptosis in the glandular epithelium, ductal (and occasionally lobuloalveolar) and intraductal hyperplasia - ultimately increasing its susceptibility to chemical carcinogens. These effects were observed in rodent or in non-human primate following prenatal and/or post-natal exposure to BPA. Available data also support the plausibility that BPA, through interaction with the nuclear estrogen receptors (ERs), or G protein-coupled estrogen receptor (GPER) and indirectly with the progesterone receptor (PR), modulates estrogen and progestin agonist activities. Emerging epigenetic studies have reported changes related

to estrogen-dependent genes (such as EZH2 and HOTAIR), as well as HOX genes (involved in embryogenesis and post-natal development) associated with BPA induced abnormal development and increased cancer susceptibility of the mammary gland.

BPA has been demonstrated to alter memory and learning after developmental, pubertal or adult exposure, based on multiple converging experimental studies reporting this functional effect as well as molecular and cellular changes in the brain (reduced expression of NMDAR, altered synaptogenesis). These effects are mediated through disturbance of estrogenic pathways as evidenced by the reversal of the functional, cellular and molecular effects of BPA by an ER antagonist and interference of BPA with estradiol-induced effects on behavior and spine density/neurogenesis.

The effects of BPA on metabolism in rodent and non-rodent after prenatal and/or perinatal or adult exposures include alteration of insulin secretion and/or release by βpancreatic cell, or of insulin signalisation (signaling mechanisms) within insulin-sensitive organs (i.e., liver, muscle, adipose tissues) leading to variations in the expression levels of hepatic or adipose tissue markers which are indicative of a state of insulin resistance. It is therefore considered that BPA may increase the incidence of type-2 diabetes. Additionally, in vivo and in vitro experimental studies indicate that these effects may involve ERa, ERB or GPR30 pathways. Other hormones such as leptin and adiponectin, which are involved in resistance to insulin and lipogenesis, are also modified following BPA exposure. This shows that BPA could interfere in the balanced interplay between insulin secretion and insulin action that controls glycaemia. Most of the in vitro studies showing adverse effects of BPA on adipocyte differentiation and function point to alteration of endocrine mechanisms (e.g., adiponectin release, insulin signaling cascade effectors). Overall, it is suggested that the pancreas is targeted by BPA, the mechanisms could differ depending on the period of exposure (fetal life or adulthood) and that an ED MoA is involved. Lastly, mainly based on similarities in homeostatic regulation of insulin production and sensitivity between animals and humans, these effects are considered relevant for humans.

The steps of the respective mechanisms of action are specific for each effect. The complexity of the toxic response to BPA suggests multiple MoA that may interact but most importantly, the available evidence shows that disruption of the estrogenic pathway is central and consistently involved in each of the four effects.

In conclusion, on the basis of evidence available in relation to alteration of reproductive function, mammary gland development, cognitive function and metabolism, BPA can be considered an endocrine disruptor for human health.

It is not excluded that BPA may also alter other physiological functions, e.g. the immune function, through a similar ED MoA but the level of evidence is considered insufficient at the moment for this effect to be presented.

The range of effects induced by BPA in relation to its ED MoA is considered serious. All these ED-related effects are characteristically (but not only) observed after developmental exposure to BPA, with consequences that are observed later in life. As they appear a long time after the exposure, they are indeed considered permanent and irreversible. In addition, the effects of BPA are associated with conditions that may lead to a reduced quality of life. In particular breast cancers, neurobehavioural disorders and diabetes are observed with high prevalence and increasing trends during the last decades in Europe and raise indisputable societal concern, also in relation to their potential economic burden on the health systems. Finally, for each of the four effects, the database shows important uncertainties in establishing a quantitative dose-response as well as safe levels, with some studies identifying effects at doses below the point of departure used by RAC for DNEL derivation and on-going discussions on the shape of the

dose-response relationship and the parameters impacting the dose-response (period of exposure and concomitant presence of estrogen in particular). For these reasons, the effects of BPA that are linked to ED MoA therefore give rise to an equivalent level of concern to those of other substances listed in points (a) to (c) of Article 57 of REACH.

Thus, BPA is proposed to be identified as a substance of very high concern in accordance with Article 57(f) of REACH for effects in relation to human health exerted through an ED MoA.

Registration dossiers submitted for the substance? Yes

# **PART I**

# **Justification**

# 1. Identity of the substance and physical and chemical properties

# 1.1 Name and other identifiers of the substance

**Table 1: Substance identity** 

EC number:	201-245-8
EC name:	4,4'-isopropylidenediphenol
CAS number (in the EC inventory):	80-05-7
Deleted CAS numbers:	27360-89-0; 28106-82-3; 37808-08-5; 137885-53- 1; 146479-75-6; 1429425-26-2
CAS name:	phenol, 4,4'-(1-methylethylidene)bis-
IUPAC name:	4,4'-propane-2,2-diyldiphenol
Index number in Annex VI of the CLP Regulation	604-030-00-0
Molecular formula:	C <sub>15</sub> H <sub>16</sub> O <sub>2</sub>
Molecular weight range:	228.29 g/mol
Synonyms:	Bisphenol A BPA 2,2-bis(4-hydroxyphenyl)propane

# Structural formula:

$$HO - CH_3 - OH$$

# 1.2 Composition of the substance

Name: 4,4'-isopropylidenediphenol (Bisphenol A)

**Description:** 80-100%

Substance type: mono-constituent

# Table 2: Constituents other than impurities/additives

Constituents	Typical concentration	Concentration range	Remarks
4,4'- isopropylidenediphenol (bisphenol A) (EC 201- 245-8)	80-100%	No information	

## **Table 3: Impurities**

Impurities	Typical concentration	Concentration range	Remarks	
None relevant for SVHC	identification			

#### **Table 4: Additives**

Additives	Typical concentration	Concentration range	Remarks		
None relevant for SVHC identification					

# 1.3 Identity and composition of degradation products/metabolites relevant for the SVHC assessment

Not relevant for the identification of the substance as SVHC in accordance with Article 57 (f) REACH.

# 1.4 Identity and composition of structurally related substances (used in a grouping or read-across approach)

Not relevant for the identification of the substance as SVHC in accordance with Article 57 (f) REACH.

# 1.5 Physicochemical properties

Not relevant for the identification of the substance as SVHC in accordance with Article 57 (f) REACH.

# 2. Harmonised classification and labelling

Bisphenol A is covered by Index number 640-030-00-0 in part 3 of Annex VI to the CLP Regulation as follows, as amended by Commission Regulation (EU) 2016/1179 (9th ATP)9:

Table 5: Classification according to Annex VI, Table 3.1 (list of harmonised classification and labelling of hazardous substances) of Regulation (EC) No 1272/2008 (as amended by Commission Regulation (EU) 2016/1179)

Index	International	EC No	CAS No	Classification Labelling				Notes		
No	Chemical Identificatio n			Hazard Class and Category code(s)		Pictogram , Signal Word code(s)	Hazard state- ment code(s)	Suppl. Hazard state- ment code(s)	Conc. Limits, M- factors	
604- 030- 00-0	bisphenol A; 4,4'- isopropyliden ediphenol	201- 245-8	80-05-7	Repr. 1B STOT SE 3 Eye Dam. 1 Skin Sens. 1	H360F H335 H318 H317	GHS08 GHS07 GHS05 Dgr	H360F H335 H318 H317			

<sup>&</sup>lt;sup>9</sup> COMMISSION REGULATION (EU) 2016/1179 of 19 July 2016 amending, for the purposes of its adaptation to technical and scientific progress, Regulation (EC) No 1272/2008 of the European Parliament and of the Council on classification, labelling and packaging of substances and mixtures

# 3. Environmental fate properties

Not relevant for the identification of the substance as SVHC in accordance with Article 57 (f) REACH.

# 4. Human health hazard assessment

#### **Context**

As summarised by Kortenkamp *et al.* (2016), the endocrine disrupting properties of Bisphenol A (BPA) have been discovered through screening exercises back in the 1930's. Chemists synthesised the chemical BPA in the laboratory in 1891. In the 1930's, scientists were searching for synthetic chemicals that could replace the expensive natural estrogen in pharmacological applications. They identified BPA as a weak functional estrogen. Its use as a pharmaceutical hormone was precluded by the synthesis of another chemical, diethylstilbestrol (DES), with even more potent estrogenic properties (Dodds and Lawson, 1938). DES was subsequently used as a pharmaceutical but showed severe side effects (Meyers, 1983).

## **General approach**

The WHO/IPCS (2002) definition of an endocrine disruptor (ED) is widely accepted:

"An endocrine disruptor is an exogenous substance or mixture that

- alters function(s) of the endocrine system
- and consequently causes
- adverse health effects in an intact organism, or its progeny, or (sub)populations."<sup>10</sup>

The European Commission's Endocrine Disrupters Expert Advisory group agreed in 2013 "that the elements for identification of an endocrine disrupter were demonstration of an adverse effect for which there was convincing evidence of a biologically plausible causal link to an endocrine disrupting mode of action and for which disruption of the endocrine system was not a secondary consequence of other non-endocrine-mediated systemic toxicity. Relevance of the data to humans should be assumed in the absence of appropriate data demonstrating non-relevance." (JRC 2013)

It is assumed in this report that a substance should fulfill the recommendations from the European Commission's Endocrine Disrupters Expert Advisory group outlined above in order to be identified as an endocrine disruptor, and available information has accordingly been assessed based on:

- Adverse health effects
- Endocrine mode of action (MoA)
- Plausible link between adverse effects and endocrine MoA
- Human relevance

As discussed in Kortenkamp *et al.* (2012) there is a clear distinction between the terms MoA and mechanism of action. The mechanism of action is typically defined as the totality of mechanistic steps, whereas the MoA refers to a less detailed sequence of key events. MoA is not intended to build a comprehensive model of a chemical mechanism of action. This is also in line with definitions proposed in the OECD guidance document on adverse outcome pathways (AOP) (OECD, 2013): "MoA differs from mechanism, in that the MoA requires a less detailed understanding of the molecular basis of the toxic effect".

 $<sup>^{10}</sup>$  Exact text quoted from WHO/IPCS (2002) definition but formatting of the text using bullet points added to emphasise major components of the definition.

For the purpose of the present analysis, the following definition is retained for MoA: "A sequence of key cellular and biochemical events with measurable parameters that result in a toxic effect."

BPA has been shown to trigger various adverse effects on health, as previously discussed at the EU level in the regulatory context of CLP (CLH dossier on fertility, ANSES 2013a) and REACH (restriction proposal of BPA in thermal paper, ANSES 2014). An ED MoA is considered to play a substantial role for several of these effects and it is important to consider and analyse in this report the scope of effects that are at stake when considering the ED properties and potential health impacts of BPA.

The adverse effects of BPA to be reviewed in the present dossier have been selected on the basis of the following criteria:

- The adverse effect is identified with a sufficient level of evidence in previous regulatory EU discussions, i.e. acknowledged by either a harmonised classification or by consideration of the effect in the process of risk assessment by RAC in the context of restriction (ECHA, 2015).
- There is relevant evidence of an ED MoA.

These criteria were considered to be fulfilled for the following adverse effects of BPA in relation to their ED MoA:

- Effect on the reproductive function
- Alteration of mammary gland development
- Alteration of brain development and cognitive function
- Metabolic alterations

It is noted that the three last effects are not acknowledged by an harmonised classification. Indeed, with respect to regulatory priorities in harmonisation as emphasised in article 36(3) of CLP and for the sake of regulatory efficiency, the harmonisation procedure of BPA in the classification dossier (ANSES, 2013a) focused only on effects on fertility. However, these additional effects were included in a previous restriction analysis and already discussed by RAC. Harmonisation of the identification of these additional effects *via* harmonised classification is not expected to result in any major additional regulatory impacts and is therefore not foreseen.

Each of these effects will be presented in a separated section. Depending on the complexity and level of evidence for each effect, a more specific adverse effect may be chosen, described and analysed for its ED-related MoA in more detail for the sake of producing a clear and detailed analysis representative of BPA's ED MoA.

Other effects of BPA on human health e.g. immunotoxicity may involve an ED MoA. However, the adversity of these effects and/or the ED MoA may not yet be clearly demonstrated with a sufficient level of evidence. They are shortly discussed in the general conclusion to ensure a comprehensive picture of the substance effects. Moreover, effects of BPA on the environment in relation to an ED MoA are not addressed in this dossier.

#### Considerations related to the relevance of data

The presentation of the specific adverse effects selected and the analysis of their ED MoA are generally based on literature searches up to May 2016.

Many studies have investigated the effects of BPA by oral route of exposure as well as by

subcutaneous route of administration. As discussed in detail in a previous report (ANSES, 2014), when BPA is administered by subcutaneous route, the daily dose can be controlled with greater accuracy, making it possible to reproduce a linear exposure pattern. In addition, subcutaneous administration bypasses the digestive barrier, intestinal and/or skin metabolism and the enterohepatic first-pass effect. The data collected in rodents show significantly higher proportions of unconjugated BPA after subcutaneous and intraperitoneal administrations, than in the case of an oral administration. It is generally well accepted that unconjugated BPA is the biologically active form. In humans, after oral exposure, BPA is known to undergo a high hepatic first-pass effect resulting in a short half-life (< 6 hours) and a low systemic availability of unconjugated BPA. In rats, BPA is also predominantly glucuronidated, but the BPA-glucuronide formed undergoes enterohepatic recirculation resulting in slower elimination of BPA (half-lives between 20 and 80 hours). The oral administration of an equivalent dose may result in higher plasma levels of unconjugated BPA in rats compared to humans. However, in rodents and to a lesser extent in humans, the hepatic metabolism capacity in newborns is limited, resulting in a reduced hepatic first-pass effect. Therefore, the effects observed after subcutaneous exposure are considered relevant when the exposure has taken place during the developmental period. Studies with BPA subcutaneous administration in which the exposure took place in adults were only considered to be informative during the identification of the MoA of BPA.

The other routes of administration used (intracerebral, intraperitoneal) are anecdotal and the corresponding studies were considered only in relation to the analysis of the MoA.

The selection of the studies have not been restricted to specific levels of doses and both "low doses" as well as "standard doses" for regulatory testing have been considered as relevant for the identification of adverse effects and the understanding of the MoA. It is, however, recognised that the MoA may have a different pattern and modulations across the whole range of doses.

Finally, although not considered as relevant for the identification of an adverse effect, studies performed in non-intact animals (i.e. ovariectomised animals) were included for the understanding of the MoA.

Systematic rating of studies using Klimisch scores was not considered relevant for the present analysis. Klimisch scores are intended to provide a score in comparison to standard regulatory guidelines. However, several of the specific adverse effects examined in the present analysis (e.g. alteration of the mammary gland development, alteration of neural structures and alteration of insulin production and insulin sensitivity) are not addressed in any specific guideline protocol. In addition, the analysis was focused on studies that preferentially investigate mechanistic aspects and not only the adverse effect. Standard regulatory guideline studies are for this purpose neither required nor generally conducted for practical reason as very complex protocols would be needed to additionally dig into specific parameters. Finally, for most adverse effects, a very large database of studies is available and not all studies provide similar results. Therefore it was considered most relevant to follow a weight of evidence (WoE) approach for the present analysis. As defined in ECHA's Practical Guide: How to use alternatives to animal testing (ECHA 2016) "The weight of evidence approach commonly refers to combining evidence from multiple sources to assess a property under consideration". As discussed in the guide, the WoE approach is beneficial when the information from each source individually may be regarded as not sufficient and when several available studies give conflicting results. It also emphasizes that "Expert judgement is vital in the construction and appraisal of the WoE package, namely when considering the reliability, relevance and adequacy, integrating and comparing different pieces of information and assigning a weight to each piece of data."

The present analysis was performed in collaboration with the ANSES' Thematic Working

group on Endocrine Disruptors. Scientific studies considered irrelevant due to major deficiencies in their design and/or reporting were not included in the analysis and are not presented in the report. The studies were considered on the basis of their relevance, reliability and adequacy for the analysis and were qualitatively weighted based on expert judgement to produce a conclusion on the selected adverse effects and their ED MoA. Human data were analysed together. Experimental data were compared to each other with specific consideration given to the periods of exposure in particular. The conclusion of the WoE for each effect was based on the combination of human and experimental *in vivo* and *in vitro* data.

In order to provide further support and transparency to the robustness of the conclusions, ToxRtool was used to assess the reliability of the experimental studies that were considered as the most informative to reach the conclusions. ToxRtool<sup>12</sup> was developed by the European Commission's Joint Research Center in 2009 (Segal *et al.*, 2015) and builds on Klimisch categories by providing additional criteria and guidance for assessing the reliability of toxicological studies. It is applicable to various types of experimental data, endpoints and studies (study reports, peer-reviewed publications).

The rating of the most informative studies to reach the conclusions resulted in a reliability score of 1 (reliable without restriction) or 2 (reliable with restriction) according to ToxRtool. They are identified by a grey shade in the summary tables in each section. Subcutaneous route was considered a relevant route of exposure as discussed above. In some cases, the use of a single dose level in a study was not systematically considered as a substantial flaw but its score was downgraded of one category in the rating in order to take into account this weakness. This final scoring reflects the overall quality of the study itself and its context. For example, studies conducted with one dose level by a same research group having previously performed studies with multiple dose levels, were considered as reliable.

# 4.1 Binding of BPA to hormonal receptors

With two hydroxyphenyl rings, BPA has structural features (Figure 1: Chemical structure of  $17\beta$ -estradiol (E2), diethylstilbestrol (DES) and Bisphenol A (BPA)) that confer the ability to bind to the two nuclear estrogen receptors  $\alpha$  and  $\beta$  (ER $\alpha$  and ER $\beta$ ) (INSERM, 2011).

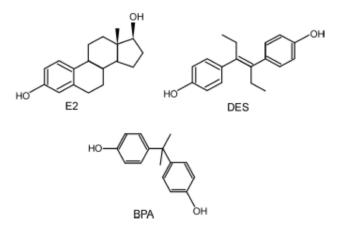


Figure 1: Chemical structure of  $17\beta$ -estradiol (E2), diethylstilbestrol (DES) and Bisphenol A (BPA). From INSERM 2011.

<sup>11</sup> https://www.anses.fr/en/content/endocrine-disruptors

<sup>12</sup> https://eurl-ecvam.jrc.ec.europa.eu/about-ec<u>vam/archive-publications/toxrtool</u>

Kuiper et~al. (1998) have measured the binding affinity of BPA to ER $\beta$ , which is 10,000-fold lower than 17 $\beta$ -estradiol. Similar results were obtained for ER $\alpha$  by Lee et~al. (2012). On this basis, BPA is generally considered as a weak agonist of ER $\alpha$  and ER $\beta$ . Examining the ER conformation, Acconcia et~al. (2015) however reported that BPA may act as an ER $\alpha$  agonist while BPA does not allow the ER $\beta$  ligand-binding domain to assume the right conformation, thus acting as an antagonist.

In addition, several studies indicate that BPA may exert effects at lower concentrations, compared to ER-binding effective doses, and very quickly through the involvement of extranuclear receptors. Studies by Watson  $et\ al.$  (2007 and 2010) have identified quick (< 1 minute) responses with low doses (1 picomolar) of BPA mediated through ER that are localised in the plasma membrane.

BPA may also interact with the transmembrane G protein-coupled estrogen receptor 1 (GPER or GPR30) (Wozniak *et al.*, 2005). GPER is largely recognised as the mediator of 'rapid nongenomic' effects by modulating second messengers and kinase pathways, which may also regulate gene expression (Filardo *et al.*, 2000; Prossnitz and Maggiolini 2009).

BPA was also identified as an estrogen-related receptor  $\gamma$  (ERR $\gamma$ ) ligand (Takanayagi *et al.*, 2006; Abad *et al.*, 2008; Okada *et al.*, 2008b), of which the specific physiological functions are unknown. The ERR $\gamma$  does not bind to estrogens, however ERR $\gamma$  can bind to estrogen response elements. The ERR $\gamma$  is highly expressed in the mammalian brain during development as well as in the brain, lung and other tissues in adults (Acconcia *et al.*, 2015). ERR $\gamma$  is a constitutively active receptor and the affinity of BPA to ERR $\gamma$  is in the order of magnitude of the nanomolar.

As reported in INSERM (2011), several studies (e.g. Sohoni et al., 1998; Li et al., 2010) have shown that BPA binds to the androgen (AR) nuclear receptor. BPA is an antagonist for AR and its affinity is in the order of the micromolar causing a moderate anti-androgenic effect.

Halogenated BPA-derivates, but not BPA, have been shown to bind to thyroid hormones receptors (Kitamura *et al.*, 2002).

Finally, BPA may also activate other cellular receptors although no direct binding has been established. BPA is able to induce the expression of the nuclear receptor involved in the proliferation of PPAR $\gamma$  (Kwintkiewicz *et al.*, 2010). Some studies report a modification of the aromatic hydrocarbon receptor (AhR) expression (Nishizawa *et al.*, 2005; Bonefeld-Jørgensen *et al.*, 2007). BPA is able to activate retinoid X receptors (RXR) in a reporter gene assay with metabolism (Li *et al.*, 2008), to act as a potent agonist for human (but not mouse) pregnane X receptor (PXR) (Sui *et al.*, 2012) and to increase the expression of nuclear receptor Nur77 (Song *et al.*, 2002; Ahn *et al.*, 2008) involved in steroidogenesis.

# 4.2 Effect on the reproductive function

# **4.2.1** Overview of previous evaluation of BPA's effect on reproductive function

### **Overview**

Several recent experimental studies have investigated the effects of BPA on the reproductive system and reported a broad range of effects. Some of the parameters related to the reproductive function were also investigated in epidemiological studies. The main outcomes of these studies are synthetised below to give an overview of the toxicity of BPA on the reproductive function.

In particular in females, the following effects are reported:

- Ovarian toxicity: experimentally, an increased incidence of cystic ovaries has been consistently reported in several studies in mice and rats (Newbold et al., 2009; Signorile et al., 2010; Delclos et al., 2014; Newbold et al., 2007; Adewale et al., 2009; Fernandez et al., 2009). A reduced ovarian weight or small ovaries, depletion of corpora lutea (Nikaido et al., 2004; Takagi et al., 2004; Newbold et al., 2007; Adewale et al., 2009; Nikaido et al., 2005) and of antral follicles (Delclos et al., 2014; Fernandez et al., 2009) as well as decreases in the number of primordial follicles (Wang et al., 2014a; Rodriguez et al., 2010) are often, although not systematically, observed. In some studies investigating the reproductive tract in older animals, more severe lesions were also found, including progressive proliferative lesions of the oviduct (Newbold et al., 2009). Concerning the oocytes development, meiotic abnormalities leading to aneuploidy were shown in several studies using different exposure scenarios. Although rodent ovarian cysts are not the "exact" model for polycystic ovary syndrome (PCOS), it is interesting to note that in humans a crosssectional epidemiological study with limitations showed that women with polycystic ovaries compared to controls have higher serum BPA concentrations (Kandaraki et al., 2011).
- Alterations of uterus morphology: in experimental studies, BPA also induces changes in the uterus morphology in several studies, mainly in the form of hyperplasia of the endometrium. Benign lesions like endometrial hyperplasia or atypical hyperplasia, which is a precursor lesion to adenocarcinoma, were reported in several studies in mice (Newbold et al., 2009 and 2007; Signorile et al., 2010; Hiyama et al., 2011). Malignant invasions (squamous metaplasia or polyps) were also described in 18-month-old mice (Newbold et al., 2009 and 2007). An increased thickness of uterine epithelia and stroma and an increased incidence in cystic, hyperplastic and metaplastic endometrium are also reported in some studies in rats (Mendoza-Rodriguez et al., 2011; Delclos et al., 2014). Epidemiological studies reported contradictory associations between the endometrial hyperplasia and BPA concentration (Cobellis et al., 2009; Itoh et al., 2007; Hiroi et al., 2004). As they presented some methodological limitations, they were not considered to provide conclusive elements.
- Age at puberty: in rodents, the effects of BPA on puberty timing are still not clear. An acceleration of puberty is observed in several studies in mice (Howdeshell et al., 1999, Honma et al., 2002; Nikaido et al., 2004; Naulé et al., 2014; Nah et al., 2011; Tyl et al., 2008). It is also observed in rats in some studies (Patisaul et al., 2014; Adewale et al., 2009; Fernandez et al., 2009), but an absence of effect (Kwon et al., 2000; Rubin et al., 2001, Tagaki et al., 2004; Yoshida et al., 2004; Ryan et al., 2010; Delclos et al., 2014; Laws et al., 2000) or even a delayed pubertal timing was reported in other studies (Tinwell et al., 2002, Tyl et al., 2002). The diverging results may, in part, be explained by differences in sensitivity in terms of exposure period,

strain of rats and/or BPA doses but are not fully understood. In humans, two recent studies performed in a large number of girls do not identify a relationship between the BPA urinary concentration and the onset of their puberty (Wolff *et al.*, 2008 and 2010).

- <u>Estrous cycle disturbance</u>: consistent results of an adverse effect of BPA on the estrous cycle are reported in several different experimental studies, including irregular and prolonged cycles in mice (Honma *et al.*, 2002; Nikaido *et al.*, 2004; Wang *et al.*, 2014a; Nah *et al.*, 2011; Tyl *et al.*, 2008) as well as in most studies in rats (Rubin *et al.*, 2001, Mendoza *et al.*, 2011; Patisaul *et al.*, 2014; Delclos *et al.*, 2014; Adewale *et al.*, 2009; Fernandez *et al.*, 2009; Zaid *et al.*, 2014; Lee *et al.*, 2013;Laws *et al.*, 2000). One epidemiological study investigated the link between BPA and the characteristics of menstrual cycles. An association with shorter luteal phases was observed even if no robust conclusion in humans can be drawn on the basis of this single study (Jukic *et al.*, 2015).
- Fertility parameters: a decline in reproductive capacity, i.e. a decrease in the number of pregnancies and/or a decrease in the number of pups born (decreased litter size), was observed in several studies when exposure of dams occurs in utero or during the first days of life in mice and rats (Wang et al., 2014a; Cabaton et al., 2011; Fernandez et al., 2009; Varayoud et al., 2011). Exposure to BPA during adulthood consistently results in a decrease in the number of pregnancies and implantations in several studies (Moore-Ambritz et al., 2015; Berger et al., 2008 and 2010; Al Hiyasat et al., 2004; NTP, 1985). In humans, high levels of BPA were associated with implantation failures in women undergoing medically-assisted procreation (Ehrlich et al., 2012) or with consecutive spontaneous miscarriages (Sugiura-Ogasawara et al., 2005). Decreased ovarian function as part of an in vitro fertilisation (IVF) was also reported with higher BPA exposure in two studies (Fujimoto et al., 2001; Mok-Lin et al., 2010).

Regarding the male reproductive function and fertility, despite the fact that some studies observe limited or no effects, there is a convergence in several studies in the effects observed after BPA exposure. Sperm production is decreased and several studies report effects on one or several male reproductive organs such as effects on the weight of the testes and the seminal vesicles, the ventral prostate or the epididymides (Tinwell *et al.*, 2002; Ida *et al.*, 2002; Kabuto *et al.*, 2004; Akingbemi *et al.*, 2004; Okada 2008a; Salian *et al.*, 2009b and 2009c; Aikawa *et al.*, 2004; Nakamura *et al.*, 2010; Tan *et al.*, 2003; Takahashi *et al.*, 2003; Chitra *et al.*, 2003; Toyama *et al.*, 2004; Tyl *et al.*, 2008).

In humans, occupational exposure to BPA was associated with a greater risk of sexual dysfunction, a declining sexual function or a decrease in sperm concentration, motility and vitality compared to unexposed workers (Li *et al.*, 2010a, 2010b; Li *et al.*, 2011). No association between BPA exposure and sperm parameters were identified in a population of fertile men (Mendiola *et al.*, 2010), but a link between BPA and impaired sperm quality was established in men consulting an infertility clinic and between BPA and higher embryo fragmentation score (EFS) and a reduced embryo cell number (ECN) in men involved in IVF (Meeker *et al.*, 2010b; Bloom *et al.*, 2011).

Many effects listed above seem to be highly sensitive to many factors across studies, including the level of doses, period of exposure, species and strain differences. Due to variations in the combination of these factors in the different experimental studies available, as well as to variations in the endpoints investigated and sometimes to experimental limitations (limited group size), it is very difficult to capture the specific influence of each parameter on the effect. However, considering the respective limitations and strengths of the different studies available, as well as the reproducibility of effects for many specific endpoints (ovarian toxicity, alteration of uterus morphology, estrous cycle disturbance, impairment of fertility parameters and impairment of sperm

production), the toxicity of BPA on the reproductive function has been established with a sufficient weight of evidence.

# Assessment of RAC under the CLH process

In its opinion of March 2014 (ECHA, 2014), RAC adopted the following conclusions in support of classification of BPA as Repr 1B – H360F as quoted in the text box hereafter:

"Effects on female reproductive capacity:

In one of the supplementary studies (Cabaton et al., 2011), using a forced breeding design enabling the identification of effects that became apparent with time, a reduction in the cumulative number of pups from F1 females exposed in utero to BPA in the highest dose group was observed. This finding was evident in the absence of systemic toxicity in the exposed F0 dams and the F1 generation. The effect on the number of pups was therefore not considered to be a secondary non-specific consequence of other toxic effects. The effect reported in the Cabaton et al. (2011) study supported the effects on fertility reported in the NTP (1985) continuous breeding study and in the multigeneration study by Tyl et al. (2002). There were also other subcutaneous non-guideline supplementary studies with higher doses but with shorter exposure periods than in the study Cabaton et al. (2011), providing some support to the fertility effects in the NTP (1985) and Tyl et al. (2002) studies.

# Effects on female reproductive organs:

RAC concludes that BPA exerts its toxic effects on the ovaries, either due to direct effects on the ovaries or indirectly via effects on the HPO-axis. Regarding the effects on uterus morphology, the majority of the studies reported hyperplasia of the endometrium or no effects. The effect of BPA on the onset of puberty seemed to vary according to experimental design such as exposure period, species, strain and dose. This variation in the onset of puberty was also seen in animals in the positive control group, e.g. orally administered 17a-ethinylestradiol (EE2) markedly delayed the onset of puberty in SD rats in the NCTR (2013) study, whereas it accelerated it in Long Evans rats in the study by Ryan et al. (2010). The guideline studies did not report any significant effects on the oestrous cyclicity. However, in the NCTR (2013) study and in most of the studies using subcutaneous dosing, BPA induced irregularities in the oestrus cycle (Mendoza-Rodríguez et al., 2011, Kato et al., 2003; Fernandez et al., 2009).

# Effects on male reproductive organs:

Several non-guideline supplementary studies included in the CLH report demonstrated effects of BPA on male reproductive function. The original studies had variable shortcomings e.g. small sample sizes, a few or single dose groups, non-oral routes of administration and/or lack of details on the methodologies used, which in some cases limited the usefulness of the findings. RAC notes that differences in strains, doses, routes of exposure or windows of exposure made a direct comparison between the studies sometimes difficult. Despite the limitations in these studies, they were considered acceptable to be used in a weight of evidence approach. More than 2/3 of the supplementary oral route studies included in the CLH report reported effects on male sexual parameters (either on sperm quality, spermatogenesis, sex hormones, or on sexual function). In several studies, exposure to BPA (at various doses or periods of exposure) led to a decrease in the serum testosterone level, to some effects in reproductive organs, and/or to a decrease in sperm production.

RAC concluded that these findings supported the observations reported in the test-guideline studies (Tyl et al., 2002; Tyl et al., 2008; NTP 1985b) and in the recent GLP-compliant NCTR 2013 study.

[...]

Effects on the male reproductive tract, evident as an impaired sperm production following BPA exposure, were observed in several studies. The decrease in sperm production was accompanied by lower testosterone levels. RAC noted that the observed effects on the testosterone levels may have been the cause of the decreased sperm production."

# Identification of the ED MoA of BPA on the alteration of the reproductive function: focus on a specific effect on the reproductive function

Considering that the effects of BPA on the reproductive function have been presented and discussed in an extensive way at EU level under the previous classification and restriction regulatory processes, it was decided not to further present nor discuss the whole database in the main part of the present dossier.

For the purpose of demonstrating that BPA exerts its effects on reproduction through endocrine disruption and, for the sake of clarity considering the extent of the database, the analysis will focus on a more specific well established effect and for which the ED MoA is the most obvious.

With this aim, the following section will focus on the ability of BPA to affect cyclicity of the female reproductive system and how this alteration is exerted through hormonal changes.

The summary of the effects of BPA on toxicity for reproduction presented above is mainly based on the literature collected for the classification and restriction dossiers. However, recent studies investigating alterations of the estrous cyclicity have been collected until May 2016 to produce the detailed analysis presented hereafter.

# 4.2.2 Adverse effect: alteration of estrous cyclicity

## 4.2.2.1 Non-human information

Experimental studies investigating the effects of exposure to BPA on estrous cyclicity are summarised in Table 8 presented in section 4.2.5.

Reliable results were reported in several experimental studies showing an adverse effect of BPA on the estrous cycle, including irregular and prolonged cycles.

In mice, this effect was observed following subcutaneous or oral administration during prenatal (Nikaido *et al.*, 2004; Honma *et al.*, 2002; Wang *et al.*, 2014a) or postnatal exposure (Nah *et al.*, 2011). No effect was observed further to the prepubertal exposure (Nikaido *et al.*, 2005) or adult exposure (Moore-Ambritz *et al.*, 2015). Although an effect was not reported in the two-generation study by NTP (NTP 1985), an increased incidence of females in estrus was identified at the high dose in the two-generation study by Tyl *et al.* (2008) in mice by oral route.

In rats, similar effects were consistently observed in Wistar and Long Evans rats after perinatal exposure by oral route (Mendoza-Rodriguez *et al.*, 2011), postnatal exposure by subcutaneous route (Adewale *et al.*, 2009; Monje *et al.*, 2010) and adult exposure by oral route (Laws *et al.*, 2000). The results were inconsistent with Sprague-Dawley rats, a strain considered sensitive to estrogens. Several studies fail to reveal an effect of BPA on the estrous cycle (Tinwell *et al.*, 2002; Kwon *et al.*, 2000; Tagaki *et al.*, 2004; Yoshida *et al.*, 2004; Ferguson *et al.*, 2014; Tyl *et al.*, 2002) while others report alterations of cycles after perinatal (Rubin *et al.*, 2001; Delclos *et al.*, 2014)), postnatal (Fernandez *et al.*, 2009), peripubertal (Zaid *et al.*, 2014) or adult exposure (Lee *et al.*, 2013). These data

show that the reproducibility of this effect is strain dependant.

In its opinion of March 2014 (ECHA, 2014) in support of classification of BPA as Repr 1B – H360F, RAC produced the following conclusions on estrous cycle disturbances in experimental studies:

"Three of the guideline studies (NTP, 1985, Tyl et al., 2002, 2008, Ema et al., 2001) did not report any significant effects on the oestrous cyclicity. However, in Tyl et al., 2008, a higher percentage of the high-dose females were in oestrus as compared to controls. Furthermore, in the NCTR $^{13}$  (2013) study, in which SD rats were exposed during GD6-PND90, 63% of the animals in the high-dose group had an asynchronous oestrous cyclicity versus 12% in the vehicle control. It was noted by RAC that the control vehicle group was also affected as compared to the naïve controls (0% asynchronous estrous cyclicity). Based on vaginal cytology, disruption of the oestrous cycle at the highest BPA dose was reported on PND69-90 and at the two highest doses on PND150-170 in a similar manner as for the positive control (EE2). The increase of the proportion of animals showing asynchronous estrous cycle on PND150-170 was statistically significant at 100 mg/kg bw/day (n=14), but not at 300 mg/kg bw/day (n=7). Maternal toxicity in this study included a significant reduction in body weight gain (6-13% with an average at 10%) at PND4 and beyond in the two BPA high dose groups. No effect on body weight gain was observed in low dose groups.

Several of the remaining non-guideline studies reported BPA-induced irregularities in the oestrus cycle (Mendoza-Rodríguez et al., 2011, Kato et al., 2003; Fernandez et al., 2009).

In contrast, in the study by Kwon et al. (2000), in which SD rats were exposed via oral gavage to 3.2, 32 or 320 mg BPA/kg bw/day between GD11 and PND20, no effects were reported on the oestrous cycle.

RAC concluded that BPA-treated F0 females were twice more in estrus as compared to controls at 600 mg/kg in Tyl et al. (2008), and that BPA induced irregularities in the oestrus cycle also in the NCTR (2013) study and in most of the studies using subcutaneous dosing (Mendoza-Rodríguez 2011, Kato et al. 2003; Fernandez et al. 2009)."

#### 4.2.2.2 Human information

The link between BPA human exposure and cycle parameters in women has been studied in a single recent study.

In the mid-80's, 221 women with no fertility disorders were enrolled at the time they stopped contraceptive method to become pregnant and followed for up to 6 months as a population-based cohort (Jukic *et al.* 2015). From these women, daily first-void morning urine samples were collected for hormonal measurements, the ovulation day recorded; the plausible conception day and early pregnancy loss were calculated. BPA was measured using one urine sample per week pooled over one cycle.

When looking for associations between the urinary concentration of BPA in a cycle and the length of the follicular phase of the menstrual cycle, the risk of early pregnancy loss, the duration for becoming pregnant and the duration of the luteal phase, BPA concentration of a given cycle was associated with a reduced length of the luteal phase. When looking for associations with BPA concentration in the previous cycle, no association was found with the length of the follicular or luteal phases.

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<sup>&</sup>lt;sup>13</sup> In the present report the NCTR study (2013) is referenced as Delclos *et al.*, 2014

The study and its design are considered as of overall good quality although it should be noted that it is common that women experience instability in the first cycles after cessation of contraception. An imprecision in the determination of the ovulation day through daily hormonal measurement may also influence the outcome of the study.

Based on this single study, no robust conclusion can be drawn on the effect of BPA on the estrous cycle in women.

# 4.2.2.3 Summary and discussion of alteration of estrous cyclicity

An alteration of estrous cycles by BPA is identified in many experimental studies in rats and mice after exposure during different periods of exposure.

In particular, an effect was observed either:

- After exposure of adult females (Tyl et al., 2008; Laws et al., 2000; Lee et al., 2013)
- After exposure during the developmental phase of the reproductive system, i.e. in utero (Honma et al., 2002; Nikaido et al., 2004; Wang et al., 2014a), perinatal (Rubin et al., 2001; Mendoza et al., 2011; Patisaul et al., 2014; Delclos et al., 2014), postnatal (Nah et al., 2011; Adewale et al., 2009; Fernandez et al., 2009) or prepubertal exposure (Zaid et al., 2014)

This effect was recognised by RAC in its opinion in support of classification of BPA as Repr. 1B – H360F as summarised in the RAC opinion on restriction of BPA (ECHA 2015): "RAC's opinion (RAC 2014) was based on adverse effects, such as disturbances in the oestrous cycle, at a dose of 600 mg/kg bw/day (Tyl et al., 2008) and at a dose of 100 mg/kg bw/day (Delclos et al., 2014)."

Proper cyclicity is considered essential to reach successful ovulation. An alteration of cyclicity may therefore directly induce at least subfertility through disturbed (delayed or absent) ovulation. The hormonal regulation of the cycle also influences the maturation process of the ovarian follicles. Different studies have linked estrous cyclicity modification with impaired follicles such as in Zaid et al. (2014) in which BPA treatment induced a persistent diestrous (5/8 animals), an increase of the number of large antral-like follicles that did not reach ovulation and of atretic cystic-like follicles and a decrease in the number of preantral follicles and corpus luteum. These data show that BPA delays the development of preovulatory follicles and their ovulation. Modifications in the length and/or hormonal environment of the different phases may impact the quality of oocytes and the quality of embryos. An association with implantation failure and spontaneous miscarriages may also occur. Therefore, the effect on cyclicity needs to be considered in relation to the alteration of fertility observed (decreased implantation and litter size).

As synthetised by Kortenkamp *et al.* (2012), an association between menstrual cycle characteristics and sub-fecundity and spontaneous abortion has been observed in humans and lifelong menstrual patterns have been associated with chronic diseases, including breast and ovarian cancer, uterine fibroids, diabetes and cardiovascular disease. Chronic anovulation is a well-established cause of female infertility. The few studies of menstrual cycle characteristics and fecundity have found that shorter cycles were less likely to be followed by conception, while both shorter and longer cycles were more likely to be followed by spontaneous abortion. Cycles with up to 4 days menstrual bleeding had lower fecundity, and spontaneous abortion was less likely after cycles with more than 5 days of menstrual bleeding (Small *et al.*, 2006). Alteration of cyclicity is therefore considered as an effect fulfilling fully the criteria of adversity.

The specific pathways at stake during development or in adulthood are expected to be largely mediated through modification of the hormonal regulation of the cycle. They will be discussed in relation to the age at exposure in the next section on endocrine disruptive MoA.

# 4.2.3 Endocrine disruption in relation to the alteration of the estrous cyclicity

# 4.2.3.1 Adult exposure

# 4.2.3.1.1 Background on the regulation of the estrous cycle in rodents

In rat and mouse, the estrous cycle is characterised by the following sequential stages, called estrus, diestrus-1 (also named metestrus), diestrus-2 (also called diestrus) and proestrus; each of which lasting about one day. At the end of the follicle growth, ovulation occurs on estrus stage at 02:00hrs. During the night between proestrus and estrus, females are receptive for mating. After the oocyte expulsion, the follicle becomes the corpus luteum, which predominantly secretes progesterone.

The time-related changes in hormonal secretions by the ovary and pituitary gland during the estrous cycle are well-known processes (Figure 2).

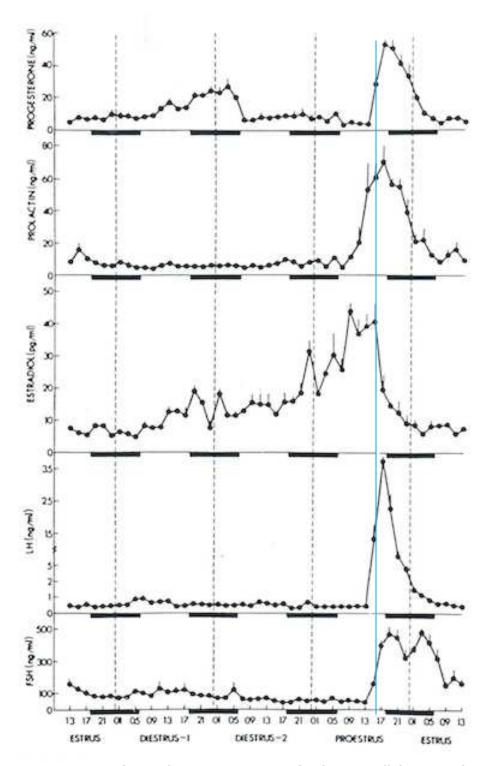


Figure 2: Concentrations of progesterone, prolactin, estradiol, LH and FSH in the peripheral plasma of the 4-days estrous cycle of the rat.

Mean values  $\pm$  SE of 5-6 rats are represented. Horizontal black bars represent the dark period in the animal room (18:00-06:00) of the 24-hr clock. Note that estradiol concentrations are expressed with pg/mL unlike all other hormonal concentrations which are expressed as  $\mu$ g/mL. The blue line indicates 15h00 at proestrus (from Smith  $et\ al.$ , 1975).

The sequence of regulation during the estrous cycle is presented in Figure 3.

The diestrus 1, diestrous 2 and proestrus before 15h00 correspond to the ovarian phase during which the pool of recruited preantral and antral follicles are growing and produce estrogens. Estrogen production results from a collaborative work inside the ovary: the theca-interstitial cells synthetise androgens from cholesterol and the granulosa cells, convert the androgens produced by theca-interstitial cells into estrogens as they specifically express *cypP450arom* (also called *cyp19a1*) encoding the aromatase, catalysing this conversion.

During this period, the follicles grow, the production of estrogens increases and the plasma estradiol concentration shows a peak during a few hours before 15h00 of the proestrus stage. These processes are stimulated by both FSH (stimulating granulosa cells) and LH (mainly stimulating theca-interstitial cells, and granulosa cells incidentally). Importantly, the preovulatory surge of estrogens is also largely due to the self-stimulation as the rise in estrogens further stimulates their own production (Figure 4). The estrogens stimulate follicle growth and protect the follicle from atresia, and consequently more and more cells produce estrogens. Furthermore, in theca-interstitial cells, estrogens act in a paracrine mode to up-regulate the stimulatory effect of LH on androgen production. Lastly, in granulosa cells, estrogens up-regulate in an autocrine fashion, via binding to ER $\beta$ , the stimulatory effect of FSH and of LH on the expression of the cypP450arom.

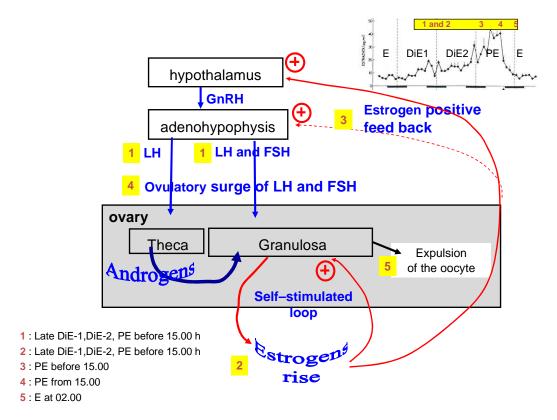


Figure 3: Temporal sequence of the main endocrine controls of the final follicle growth and ovulation in the rat

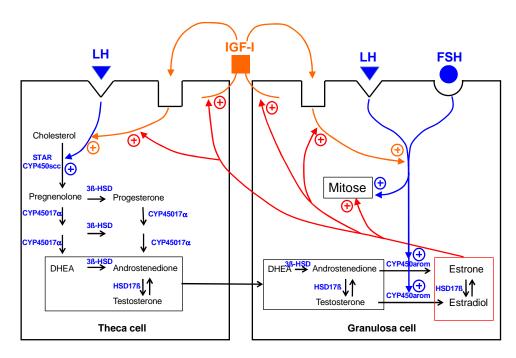
Step 1: From late Diestrus-1 (DiE-1) until proestrus (PE) 15h00 LH stimulates the production of androgens by theca cells and FSH (and incidentally LH) stimulates the conversion of these androgens into estrogens.

Step 2: Concomitantly, estrogens stimulate their own production via a self-stimulated loop detailed in Fig 2. Thus the estrogen production increases.

Step 3: High levels of estrogens act positively on the hypothalamo-hypophysis system and trigger a surge of LH and FSH from PE 15:00hrs (step 4) that induces ovulation at the estrous stage (step 5).

The side insert into fig 3 represents the level of estradiol-17ß in the peripheral plasma during the estrous cycle (from Smith et al., 1975).

The self-amplifying feedback mechanism of estrogens on their own production is mediated by intra-follicular factors. Among them, IGF-1 produced by theca-interstitial cells and granulosa cells plays an important role by enhancing the action of gonadotrophins. IGF-2, SF-1, BMP-15, GDF-9 and GATA4 are also involved. Lastly, in the granulosa cells of the preovulatory follicle, transcription factors such as PPAR- $\gamma$  inhibit CYP450arom transcription, whereas others such as LRH-1, SF-1, CREB and GATA4 activate this transcription. The ovary-specific proximal PII promoter of CYP450arom contains response elements for these transcription factors.



**Figure 4: Main mechanisms of the self-stimulated estrogen synthesis in the preovulatory follicle.** Estrogens (estrone and estradiol-17β) stimulate the proliferation of the granulosa cells, and the secretion and action of IGF-I in the granulosa cells (autocrine action), and in thecal cells (paracrine action). In each cell type, IGF-I potentiates the positive effect of the gonadotrophins (LH in theca cells, and FSH (incidentally LH) in the granulosa cells) on specific steps in the steroid hormone biosynthetic pathway. STAR, steroid acute regulatory protein; CYP450scc: P450 cholesterol side chain cleavage enzyme (also named CYP11A1); CYP45017a: 17a-hydroxylase-17,20-desmolase (also named CYP17A1); CYP450arom: cytochrome P450 aromatase (also named CYP19A1); HSD3β: 3β-hydroxysteroid dehydrogenase; HSD17β: 17β-hydroxysteroid dehydrogenase; IGF-I: insulin-like growth factor I.

Aromatase is an enzyme responsible of the production of estradiol. It is expressed in multiple organs such as gonads, placenta, brain, adipose tissue, blood vessels, skin, bones and uterine mucosa (Simpson *et al.*, 1994). In the non-pregnant female, ovaries are by far the major source of circulating estrogens in vertebrates. Consequently, the increase in plasma estrogen concentration is due to an increase in their production by follicles.

During proestrus, when plasma estrogens reach a threshold, they act positively on the hypothalamo-pituitary system to provoke the ovulatiory peak of LH and FSH. This is known as the positive feedback of estrogens on the hypothalamo-pituitary system. This mechanism is common to all cycling mammals.

More precisely, the ovarian estradiol acts in the hypothalamic preoptic area to trigger GnRH liberation, which in turn stimulates LH increase in the pituitary. This gonadal feedback does not act directly on GnRH neurons but involves a neuronal cell type expressing kisspeptin. This hypothalamic neuropeptide coded by *Kiss1* gene acts upstream of GnRH. Kisspeptin neurons located in the rostral periventricular area of the third ventricle (RP3V; including the anteroventral periventricular, caudal and rostral periventricular nuclei) of the preoptic area send projections to GnRH soma cells, which express the kisspeptin receptor (Kiss1R). Kisspeptin is a crucial regulator of the onset of puberty, sex hormone-mediated secretion of gonadotrophins, and control of fertility. During the proestrous phase, estradiol targets kisspeptin neurons, which express ER $\alpha$  and therefore integrate the positive signal of estradiol necessary to trigger the ovulatory surge of LH, through GnRH liberation.

Furthermore, in rodents, the anteroventral periventricular nucleus (AVPV), where kisspeptin neurones are involved in the positive feedback of estradiol, receives afferent fibers from the suprachiasmatic nucleus. The circadian clock located in the latter nucleus coordinates and provides precise timing for the LH surge which starts at 15h00.

It may be noted that, in rodents, the ovulatory surge of LH triggers progesterone surge (peak following estradiol surge (Figure 2), which is required for female receptivity. During this stage, progesterone acts in hypothalamic areas through progesterone receptors, which are up-regulated by estradiol, thereby leading to the induction of female sexual behaviour.

In conclusion, the estrous cycle appears as a process basically controlled by sequential endocrine/paracrine and autocrine regulations. The key event is the endocrine dialogue between the hypothalamo-pituitary system and the ovarian follicles *via* the levels of estrogens that trigger the ovulatory surge of LH.

## 4.2.3.1.2 *In vitro* information indicative of endocrine activity of BPA

A number of *in vitro* studies show that BPA can alter the activity of ovarian cells. This is very likely to result in the disturbance of the estrous cycle, in particular when the steroidogenic activity of the ovaries is concerned (see above the biological process which controls the estrous cycle). The *in vitro* data are described below and summarised in Table 9 presented in section 4.2.5.

#### Theca-interstitial cells

Using isolated antral follicles from adult cycling female FVB mice, Peretz *et al.* (2011) observed dose-dependent and time-dependent reductions in estradiol-17ß, estrone, testosterone, androstenedione and DHEA-S synthesis after exposure to 100 and 10 mg/L (440 and 44  $\mu$ M) BPA. Using addition of steroid substrates and RT-PCR analyses, they demonstrated that BPA acts by reducing the activity and/or expression of STAR and CYP450scc, but not that of 3ß-HSD and CYP45017- $\alpha$ . Moreover, these effects of BPA are reversible once BPA is removed from the culture media (Peretz and Flaws, 2013).

In contrast, using theca-interstitial cells isolated from immature (30 days old) female Sprague–Dawley rats (previously daily injected with 1mg 17ß-estradiol from 28 to 30 days of age to stimulate ovarian development), Zhou *et al.* (2008) observed that BPA in the culture medium at high concentrations increases the expression of STAR (from 10 to 100  $\mu$ M i.e 2 to 23  $\mu$ g/mL) and CYP450scc (from 0.1 to 100  $\mu$ M i.e from 0.2 to 23  $\mu$ g/mL).

#### **Granulosa cells**

Rodent and porcine granulosa cells

Zhou *et al.* (2008) used a granulosa cells culture isolated from mature Sprague–Dawley rats and observed that an exposure range between 1 to 100  $\mu$ M BPA for 48 hrs reduced the estradiol production and CYP450arom mRNA level in a dose-dependent fashion.

Peretz *et al.* (2011) showed that the mouse antral follicle cultured for 120 hours in the presence of 44  $\mu$ M BPA contained 4 times lower CYP450arom than the controls, but the difference was not statistically significant.

Mlynarcikova *et al.* (2005) reported that 1 to 100  $\mu$ M BPA inhibits FSH-induced estradiol-178 synthesis in cultured granulosa cells isolated from antral porcine follicle.

#### Human granulosa cells

Importantly the negative effect of BPA on aromatase expression and activity was observed using human cells. Watanabe *et al.* (2012) used a KGN cells line (a human ovarian granulosa-like tumor cell line), and exposed them to BPA between 5 to 100  $\mu$ M. They observed a dose-dependent reduction of the mRNA levels and activity of CYP450arom.

Kwintkiewicz et al. (2010) used KGN cell line, and exposed them to BPA in a range between 40-100  $\mu M$ . They observed a dose-dependent reduction of FSH-induced aromatase expression and estradiol secretion, and a reduction of the FSH-induced IGF-1 expression. mRNA levels of transcription factors SF-1 and GATA4 were decreased after BPA treatment. In contrast, both mRNA and protein levels of PPAR $\gamma$  were significantly upregulated by BPA in a dose-dependent manner and the authors suggests that the inhibitory effect of BPA on the expression of aromatase is mediated via PPAR $\gamma$  since overexpression of PPAR $\gamma$  in KGN cells also provokes a decrease in the expression of aromatase and IFG-1

Recently, Mansur *et al.* (2016) assessed the effects of BPA on a human granulosa cells culture obtained from patients undergoing IVF. The cells were exposed for 48 hrs to 8.8 nM, 88 nM, 880 nM, 8.8  $\mu\text{M}$  or at 88  $\mu\text{M}$  of BPA. The progesterone secretion was reduced for 8.8 and 88  $\mu\text{M}$  of BPA but not at lower doses. The highest BPA concentration showed a decrease of the estradiol production. The BPA at 8.8 and 88  $\mu\text{M}$  significantly reduced the mRNA levels of 3 $\beta$ -HSD, CYP450scc and CYP450arom and at lower concentrations (8.8 nM to 0.88  $\mu\text{M}$ ) no change was observed. The BPA exposures concentration did not affect the STAR and CYP17 $\alpha$  mRNA levels. Lastly, 3 $\beta$ -HSD, CYP450scc and CYP450arom protein levels were reduced by 88  $\mu\text{M}$  of BPA.

#### Note

It can be noted that the inhibitory effect of BPA on the aromatase activity and/or expression was also observed *in vitro* in other cell types such as placental cells (Nativelle-Serpentini *et al.*, 2003; Benachour *et al.*, 2007; Huang & Leung, 2009) and Leydig cells (Akingbemi *et al.*, 2004).

### **Summary and conclusion**

In conclusion, the *in vitro* effect of BPA on the theca-interstitial cells steroidogenesis seems to depend on the species and/or the experimental procedure. In contrast, data dealing with the *in vitro* effect of BPA on the granulosa cells steroidogenesis are all converging to show that **BPA reduces the estrogen production by this cell type by reducing the aromatase expression in all species studied including in humans.** 

#### 4.2.3.1.3 In vivo evidence with regard to an endocrine MoA

There are currently few data reporting *in vivo* effects of adult exposure to BPA together with investigation of the MoA. These studies are summarised in Table 10 presented in section 4.2.5.

Lee *et al.* (2013) developed a key study using adult female Sprague-Dawley rats (PND 56) treated by oral gavage with 1 or 100  $\mu$ g/kg/day of BPA for 90 days. Estradiol benzoate (EB,  $1\mu$ g/kg/day) was used as positive control. It is a well-conducted study (n= 18 rats/group, sacrifice of all the animals at the same stage of the estrous cycle, the day of estrous). Both BPA doses and EB lengthened the estrous phase, decreased plasma estradiol and testosterone concentrations, and increased apoptosis in follicle and corpus luteum. They decreased the protein levels of StAR but not those of P450SCC and 3 $\beta$ -HSD in theca-interstitial cells. The magnitude of these effects is important: plasma estradiol-17 $\beta$  concentration was 2 times lower in rats treated by 1  $\mu$ g/kg/day as compared with controls. No change in the levels of FSH in the plasma and the pituitary gland were observed. Both doses of BPA but not EB:

- decreased aromatase levels in the granulosa cells (with a stronger effect with 1 than with 100  $\mu g/kg/d$  BPA),
- decreased estrogen-induced proteins (PCNA calbindin-D9k) and collagen contents of the uterus,
- increased plasma LH concentration and pituitary LH content. The authors interpret this increase in LH levels as the following cascade: BPA primarily acts on the ovaries to reduce their estrogen production; this provokes a partial removal of the inhibitory negative feedback that is exerted by the circulating estrogens on the hypothalamo-pituitary system at this period of the estrous cycle, and, consequently, an increase in LH secretion.

This work is in accordance with *in vitro* data described above. It shows that **one clear-cut primary target of BPA is the reduction of the expression of aromatase.** BPA first decreases estradiol levels by disturbing P450arom protein expression. Then, it is likely that the prolonged status of reduced estradiol subsequently provokes decreased feedback regulation of LH, lengthening of the estrous cycle as well as ovarian cell apoptosis.

Wang et al. (2014b) reported that adult exposure to BPA for 6 hours during proestrous, but not during estrous or diestrous, increases  $\mathit{Kiss1}$  and  $\mathit{GnRH}$  expression as well as levels of LH, FSH and estradiol. The assessment of LH surge showed an increased baseline before the LH surge, but no changes in the timing and level of this surge.

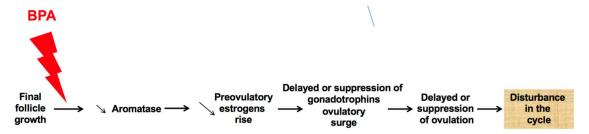
In a recent study, Kurian *et al.* (2015) using a microdialysis method, examined the effects of BPA (0.1, 1, and 10nM) directly infused to the stalk-median eminence on the release of GnRH and kisspeptin in mid to late pubertal ovarian intact female rhesus monkeys. They observed that the highest level of BPA exposure (10 nM i.e about 2 ng/ml which is a relevant concentration as far as human exposure is concerned) suppressed both GnRH and kisspeptin release suggesting that persistent exposures to BPA could impair female reproductive function by directly influencing the hypothalamic neuroendocrine function.

Although the potential effects of such changes were not assessed on the estrous cycle due to the short time of exposure, these studies suggest that BPA can also affect the expression of two key neuropeptides involved in the ovulatory surge of LH process in rodents and non-human primates and that a neuroendocrine mechanism can also contribute to BPA-reprotoxicity (see detailed presentation of neuroendocrine regulation in the next sections).

## 4.2.3.1.4 Summary of the plausible link between adverse effects and endocrine MoA regarding adult exposure

So far, there are indications of direct effects of BPA exposure during adulthood on the neuroendocrine system controlling the estrous cycle in rodents and non-human primates. However, it remains difficult to establish a clear link between the data on BPA-induced changes of this system and alterations of the estrous cycle based on the few data available. Indeed, although BPA-induced changes in the neuroendocrine expression of kisspeptin and GnRH were reported, data are far too limited to propose a succession of key events linking directly neuronal changes to ovarian cycle disruption.

On the contrary, convergent data explain how the effects of BPA on the ovary lead to alteration of the estrous cycle. The negative effect of BPA on ovarian estrogen production is clearly demonstrated: in rodents, domestic animals using both *in vitro* and *in vivo* studies and in human cells *in vitro*. Whereas the effect of BPA on theca-interstial cells depends on the model and the protocol, BPA consistently reduces the conversion of androgens into estrogens in granulosa cells. This reduction is, at least in part, a consequence of a decreased transcription of CYP450arom either *via* a direct effect in the granulosa cells or *via* changes in intrafollicular signaling factors that regulate follicular growth and endocrine activity. Given the regulatory scheme of the estrous cycle, such an alteration in the preovulatory follicle steroidogenic activity is very likely to be associated with a disruption of the estrous cycle (Figure 5) as shown in the Lee *et al.* study. **These results demonstrate a clear endocrine mode of action, namely the alteration of the ovarian steroidogenic activity, underlying estrous cycle disruption in adult rodents.** 



**Figure 5 : Sequential cascade from the endocrine effect of BPA to its adverse effects.**A clearly demonstrated target of BPA is aromatase, in the preovulatory follicle. The BPA-induced reduction in the expression of this steroidogenic enzyme induces a reduction in the synthesis of estrogens. Thus, the preovulatory rise of estrogens is attenuated. Consequently, the estrogen-induced gonadotrophins ovulatory surge, is delayed or suppressed, and this induces disturbances in the cycle.

# 4.2.3.2 Developmental exposure (*in utero*, perinatal, postnatal and/or prepubertal)

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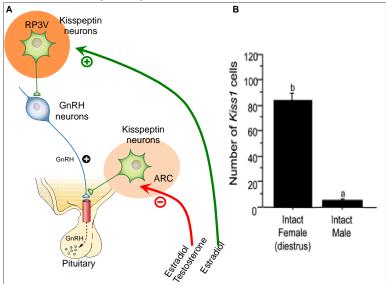


Figure 6: The neuroendocrine system involved in the control of the estrous cycle is regulated in a sexual dimorphic manner.

**A.**Kisspeptin neurons located in the hypothalamic arcuate nucleus (ARC) integrate the negative feedback exerted by estradiol or testosterone and progesterone in both males and females. By contrast, kisspeptin neurons of the RP3V nucleus integrate the positive feedback exerted by estradiol in females. (From Naulé *et al.*, 2016)

**B**. This positive regulation does not exist in males since the RP3V nucleus contains very few kisspeptin neurons in males by comparison to females (From Kauffman *et al.*, 2007). The RP3V nucleus is located in the preoptic area of the hypothalamus and includes the AVPV nuclei as well as periventricular nuclei. Kisspeptin neurons were first localised in AVPV nuclei but were later also detected in other parts of RP3V (periventricular neurons).

The neuroendocrine pathways underlying the gonadotropic function are regulated by sex steroids in a sexually dimorphic manner (Figure 6). Indeed, the positive feedback exerted by estradiol to trigger GnRH/LH preovulatory surge is specific to females. As developed above, ovarian estradiol exerts a positive control during the proestrous phase. It also exerts an inhibitory feedback during the other phases in females. In males, testosterone exerts only a negative feedback. Both positive and negative feedback exerted by sex steroids involve kisspeptin neurons. Two hypothalamic neuronal populations of kisspeptin are differentially involved in the integration of these positive and negative signals. Kisspeptin neurons of the RP3V nucleus are targeted by estradiol during the proestrous phase in females, while kisspeptin cells of the hypothalamic ARC mediate the negative control of sex steroids in both males and females. Kisspeptin neurons of the ARC send also projections to GnRH neurons and co-express two neuropeptides neurokinin B and dynorphin, which are suggested to play also a role in the regulation of the gonadotropic axis. At the neuroanatomical levels, the RP3V contains more kisspeptin neurons in females than in males.

This sexual dimorphism is programmed as early as the perinatal period and is under the control of sex steroids (Figure 7). In males, the perinatal surge of testicular testosterone masculinizes the RP3V region. In females, brain regions are not impacted by sex steroids during the perinatal period since the ovaries are inactive and the neural structures involved in female reproduction are protected from maternal and sibling sex steroids. Exogenous administration of testosterone to female neonates masculinises the preoptic area, in terms of kisspeptin neuronal number, thereby suppressing the ovulatory surge of LH during adulthood. Importantly, the same effect is obtained by administration of estradiol. Indeed, in males, perinatal testosterone is converted in the nervous system

into estradiol, which masculinises brain areas including the preoptic area.

At birth, the expression level of kisspeptin is low in the female RP3V. It increases progressively during the postnatal period under the control of ovarian estradiol. Indeed, the ovarian production of estrogens, which starts around postnatal day 7, promotes *Kiss1* expression in this hypothalamic region. A maximal increase is observed during the prepubertal period and will be necessary for pubertal activation of GnRH/LH axis and initiation of estrous cyclicity and female reproduction.

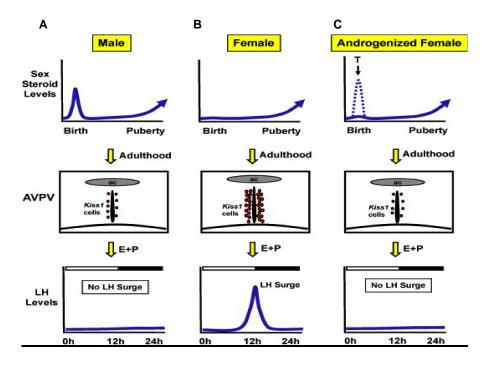


Figure 7: Perinatal organisation of the gonadotropic axis required for normal GnRH/LH surge and ovulation to occur (From Kauffman, 2009)

**A.** In males, gonadal testosterone masculinises the AVPV, thereby leading to a reduced number of kisspeptin neurons in this area and the absence of the ovulatory surge of LH in response to estradiol/progesterone stimulation. **B.** In females, in the absence of ovarian steroids, this area develops normally and the ovulatory surge of LH occurs at adulthood. **C.** Perinatal exposure of females to androgens triggers a male phenotype with low kisspeptin cell number and suppression of the ovulatory surge of LH.

### 4.2.3.2.2 *In vitro* information indicative of endocrine activity at theneuroendocrine level

One *in vitro* study (Klenke *et al.*, 2016) investigated the action of BPA on the neuroendocrine components of regulation of the estrous cycle and is summarised in Table 11 presented in section 4.2.5. In this study, BPA reduces the frequency of oscillations in GnRH neurons from embryonic nasal explants collected after emergence of GnRH cells and other neuronal cell types from the plasma codes.

# 4.2.3.2.3 *In vivo* evidence with regard to an endocrine MoA after developmental exposure

As presented above, the perinatal and postnatal periods of exposure are sensitive to hormonal changes. Changes are required for the permanent programming of the female

neuroendocrine system. Exposure to exogenous factors exhibiting hormone-mimetic activities such as BPA could then interfere with these processes and induce long-term effects on the integrity of the gonadotropic axis and the estrous cyclicity.

#### Studies associating estrous cyclicity alteration as an ED MoA of BPA

Several studies provide evidence of alteration of the estrous cycle as well as indications about the MoA. They are summarised below and in Table 12 presented in section 4.2.5.

In the study from Wang *et al.* (2014a), effects on estrous cyclicity were observed at low doses. However, the only evidence that these alterations might be due to endocrine-mediated mechanisms arises from the observation that part, but not all, of these effects are reproduced in the DES positive controls.

In the study from Delclos *et al.* (2014) issued from the BPA-clarity initiative (Heindel *et al.*, 2015), effects on estrous cyclicity were conclusive in the higher dose group after periconceptional and all life-long exposure. This effect was seen in the positive EE2-treated animals as well.

In addition, the similarity of effects between BPA and positive estrogenic controls or ER alpha antagonist was also observed in Nikaido *et al.* (2004) and Adewale *et al.* (2009), respectively.

This shows that BPA displays an estrogen-like effect.

In another study (Rubin *et al.*, 2001), results in some female offspring exposed perinatally to the highest dose of BPA (1.2 mg/kg bw/d) revealed intermittent extended periods of diestrus, whereas other females exhibited extended periods of proestrus and/or estrus. Beside altered patterns of estrous cycle in approximately 80% of 4-month and 6-month old females, the offsprings of the high-dose BPA females also revealed decreased levels of plasma luteinising hormone (LH) (-18%) in adulthood after ovariectomy. Decreased LH secretion in BPA-treated ovariectomised animals showed an alteration of the endocrine function of the hypothalomo-pituitary gonadotropic axis. However, it should be noted that LH was not evaluated in the ovary of intact animals, thus direct correlation to the estrous cycle disturbances could not be made.

More convincing evidence toward involvement of hormonal disruption was provided by the concomitant observation of estrous cycle disturbance together with modification of LH and/or GnRH release (Monje *et al.*, 2010; Fernandez *et al.*, 2009).

In addition to the studies presented above, two studies were reviewed but could not be considered in our analysis due to major methodological limitations and/or lack of details regarding the methodology, in particular inappropriate schedule of observation to properly monitor estrous cyclicicity (Patisault *et al.*, 2014) and lack of information on the stage at which samples for hormonal monitoring were collected (Zaîd *et al.*, 2014)

Finally, one additional study did not identify effects on cyclicity but provides relevant indications of a disruption of its hormonal regulation. In the study from Veiga-Lopez *et al.* (2014), sheep were exposed to BPA during gestation from GD30 to GD90 at doses of 0.05-0.5 or 5 mg/kg/d. This exposure was not associated with major alterations of the amplitude and/or the timing of the estradiol and LH surges during the preovulatory phase of the estrous cycle. However, the time interval between estradiol and LH peaks (equivalent of a proestrous phase in rodents) appeared to be decreased in BPA-exposed animals as compared to vehicle ones. The impact of such modifications on the overall estrous cycle was not determined in this study. Several other studies in sheep indicate that BPA developmental exposure (either *in utero* or neonatally) can alter the follicle dynamic (Rivera *et al.*, 2011; Veiga-Lopez *et al.*, 2014) or the ovarian response to FSH (follicular growth, FSH-induced estradiol secretion) in prepubertal animals (Rivera *et al.*, 2015).

### Evidence for a neuroendocrine basis to BPA-induced disruption of estrous cyclicity following developmental exposure

Ten studies addressing the effects of developmental exposure to BPA on kisspeptin and GnRH expression or liberation are summarised in Table 13 presented in section 4.2.5. Among these studies, 8/10 report changes in these processes, suggesting a potential long-term effect of BPA exposure at the neuroendocrine level. The limited number of studies and differences in doses and analyses do not allow to conclude whether developmental exposure to BPA inhibits or increases neuropeptide expression. It seems, however, that neonatal and early postnatal exposure diminishes, while a longer exposure time starting from gestation until weaning increases, kisspeptin expression.

Recent neuroanatomical studies described modifications in the expression levels of estrogen receptors in brain areas underlying female reproduction, such as the preoptic area and AVPV subregion or the mediobasal hypothalamus and ARC, following developmental exposure to BPA (Monje et al., 2009; Monje et al., 2010; Rebuli et al., 2014; Cao et al., 2014; Yu et al., 2015). These data clearly indicate that BPA developmental exposure can be associated in animal models with an altered development of the neuroendocrine component of the gonadal axis. From a physiological point of view, it is legitimate to assume that most of these alterations can possibly lead to disruption of estrous cyclicity later in life. However, the majority of these studies did not monitor cyclicity, which precludes any definitive conclusion regarding a potential functional link between these developmental neuroendocrine alterations and perturbation of estrous cyclicity in adults.

# 4.2.3.2.4 Summary of the plausible link between adverse effects and endocrine MoA regarding developmental exposure

Not all studies provide clear indications of a direct link between a disruption of estrous cyclicity due to BPA exposure and endocrine or neuroendocrine mechanisms. In particular, the delay between the expression of the neuroendocrine mode of action evidenced during developmental stages (evaluation of tissue expression of genes/ proteins such as kisspeptin) and the effect on estrous cyclicity that can be evidenced only in fully mature animals render their observation within the same study/animal almost impossible. Nevertheless, many studies show that the basic (neuro) endocrine mechanisms implicated in the finely tuned regulation of the gonadotropic function underlying the estrous cycle can be altered in response to BPA exposure, in particular after developmental exposure. BPA has been shown to affect the hypothalamic expression of kisspeptin, a key neuropeptide in the regulation of the hypothalamicpituitary-gonad (HPG) axis to later achieve the release of hormones at the appropriate time and concentrations during the cycle. In particular, studies by Monje et al. (2010) and Fernandez et al. (2009) provide a link between neuroendocrine changes and alteration of the cycles through concomitant observation of an alteration of hormones of the HPG axis and a cycle disturbance. In addition, the affected targets are similar to a large extent to targets affected by either estrogen agonist or estrogenic positive controls. Thus, animal and in vitro data support the hypothesis of an endocrine-related MoA of BPA to induce perturbation of estrous cyclicity after developmental exposure.

It is noteworthy however, that based on available data it is sometimes difficult to state whether those endocrine alterations are the primary mode of action or just consequences of a non-endocrine related mechanism such as meiotic alteration or epigenetic modifications in the oocytes and/or other follicular cell types. This is typical of the regulatory loop systems that are the basis of endocrinology. As long as an endocrine-related modification can be evidenced for at least one step of these regulatory loops, it can be considered that the substance is acting as an endocrine disruptor.

#### 4.2.3.3 Human relevance

Most of the evidence comes from rodent studies. Peculiarities of the reproductive physiology in those species as potential sources of uncertainties on the relevance of the results for humans are discussed hereafter together with commonalities across species.

#### 4.2.3.3.1 Circadian synchronisation of estrous cycle is specific for rodents

The preovulatory LH surge, which characterises the proestrus depends on neural hypothalamic signals tightly coupled to the 24 hrs light-dark cycle in rodents. The disruption of this signal, through pentobarbital administration during mid-proestrus for example, leads to a delayed ovulation by exactly 24 hours. In rat, the synchronisation of the estrous cycle is related to the expression of an endogenous circadian rhythm. In women, the spontaneous initiation of the preovulatory LH surge generally occurs in the morning in association with high cortisol levels, suggesting a role for the hypothalamus in timing human ovulation. However, the most recent evidence suggests that this neural component of the control system timing the LH surge in women translates diurnal changes in environmental cues rather than an endogenous circadian rhythm.

It appears therefore that in rodents, the modification of the duration of the different phases of the estrous cycle might in some cases reflect disruption of the circadian synchronisation of the GnRH/LH preovulatory surge and that this is likely not the case in humans.

The picture is quite different when the observed parameter is the percentage of females exhibiting regular estrous cycles. This type of modification is more likely to signal a profound alteration of the basic mechanisms underlying the cross talk between the ovaries, the pituitary and the brain, which are well preserved among animal species.

#### **Conclusion**

There is some degree of uncertainty regarding the relevance to humans of rodent data on the estrous cycle disturbances when they relate to the duration of each phase of the cycle. However, when the results are expressed in terms of percentage of females exhibiting irregular estrous cycles as seen in several studies with BPA it is very likely that these effects can be considered as relevant to humans. In addition, alterations of the ovarian steroidogenic activity and/or of the neuroendocrine pathways mediating sexual steroid feedback are evidenced with BPA and provide support for human relevance since these are basic mechanisms underlying the estrous cycle that are common to most mammal species.

### 4.2.3.3.2 Differences in the timing of the ontogeny of the neuroendocrine axis and/or the gonads

The sequential events and regulations during development of the gonads and neuroendocrine reproductive axis is common to all mammals including human. However, the duration of each period is highly variable as shown in Figure 8, Thus, as an example, an exposure to BPA at birth will act on ovaries at different degrees of maturation in rodents (meiosis period) and in humans (meiosis ended).

#### Conclusion

Although the critical periods of exposure may differ in humans, it does not affect the general relevance of the effect/MoA.

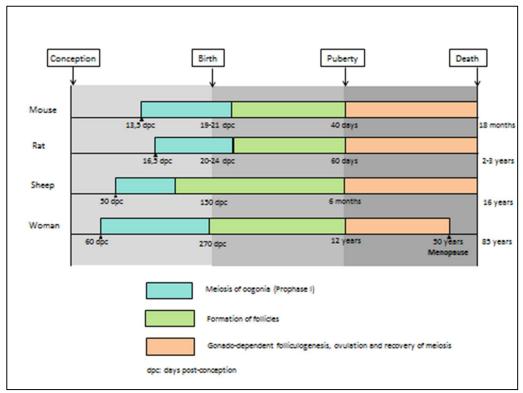


Figure 8: Ontogeny of the ovarian function in different species of mammals.

# 4.2.3.3.3 Differences and commonalities in the endocrine and neuroendocrine control of the ovarian cycle in adults

Ovarian control of the ovarian cycle

The cycle in humans differs from that in rat and mice in its duration (28 days on average), in a clear separation between follicular, and lutea phases, and a uterine cycle characterised by menstruations.

Furthermore, as explained hereabove, the follicular phase of the estral cycle in rodents is characterised by a peak of progesterone induced by the ovulatory surge of LH since the corpus lutea of the previous cycle is still functional at this time in rodents. This peak of progesterone does not impact the running of the cycle and is important to synchronise ovulation and female receptivity to male mounting in rodents. Indeed, liberated progesterone induces female receptivity, which is restricted to this period. This peak of progesterone does not occur in women, since progesterone is secreted during the luteal phase of the estral cycle only.

Furthermore, in humans there is a larger variability in the duration of the cycle from one cycle to another in the same woman and from one woman to another. It results from variability in the duration of the terminal growth of the follicle, leading to variability in the delay to reach the estrogens threshold that will trigger the pituitary gonadotrophins surge. Thus, the duration of the follicular phase is variable. Conversely, the duration of the luteal phase is relatively constant.

Another difference is the control of the *corpus luteum* regression, which is exerted by the *corpus lutea* in primates and by the uterus in rodents, but both mechanisms involving the same hormonal control involving  $PGF2\alpha$ .

In conclusion, there are some differences in the endocrine control of the cycle between rodents and humans. Nervertheless, the key regulatory endocrine mechanisms of the cycle are the same. Importantly, in all cycling mammalian species including primates, it is the progressive increase in estrogens secretion at the end of the follicular phase that triggers the release of LH and FSH ovulatory surges. In all cycling species, the experimental suppression of the production or the action of estrogens during the end of the follicular growing phase suppresses the ovulatory peak of gonadotrophins. Consequently, the BPA-induced reduction of the aromatase expression in the follicle described above is expected to trigger disturbances in the menstrual cycle in humans as well as in rodents.

#### Neuroendocrine control in humans

In humans, the importance of kisspeptin was first demonstrated by the hypogonadotropic hypogonadism of patients carrying a mutation of the KISS1R (de Roux *et al.*, 2003; Seminara *et al.*, 2003). More recent studies show that kisspeptin acts also upstream of GnRH neurones to coordinate GnRH and LH pulsatility (reviewed in Skorupskaite *et al.*, 2014). It stimulates the secretion of both LH and FSH, with a preferential stimulation of the former. Kisspeptin has also been shown to mediate both negative and positive feedback of sex steroids. In women, it seems therefore that sex steroid feedback involves both the hypothalamus and the pituitary gland.

At the neuroanatomical level, kisspeptin neurones extend from the preoptic area through to the infundibular nucleus (homologous to the ARC in rodents), as for GnRH neurones (Figure 9). In the infundibular region, kisspeptin neurones express also neurokinin B and dynorphin. By contrast to rodents where the RP3V and ARC respond to positive and negative sex steroid feedback respectively, the human infundibular nucleus relays signalling of both. It is, however, possible that the two processes are mediated by different neuronal populations.

The kisspeptin system seems also sexually dimorphic, although the critical period and origin of this dimorphism are still unknown. More kisspeptin fibres were detected in the infundibular nucleus and ventral periventricular area in women than in men (Hrabovszky *et al.*, 2010). Sex differences were also reported in the number and expression of kisspeptin cell bodies, which are present in the rostral periventricular zone of the female only.

In addition, the recent study by Kurian *et al.* (2015) on mid to late pubertal ovarian intact female rhesus monkeys suggests that persistent exposures to BPA could impair the female reproductive function by directly influencing the hypothalamic neuroendocrine function as evidenced by an alteration of kisspeptin release and GnRH pulsatility.

The role of kisspeptin in the neuroendocrine control of the HPG axis is relevant to humans. Therefore, it can be considered that BPA-induced alterations of the hypothalamic kisspeptin/GnRH system are also relevant in humans.

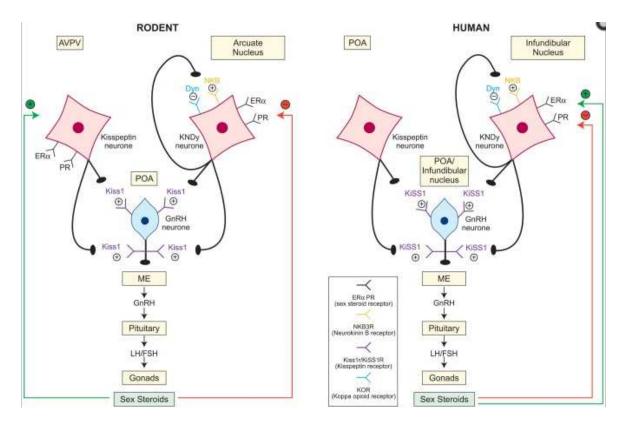


Figure 9: Comparison of kisspeptin neuronal distribution and regulation by sex steroids between rodents (left) and humans (right). Reproduced from Karolina Shorupskaite et al. The kisspeptin-GnRH pathway in human reproductive health and disease. Human Reproduction Update (2014) 20 (4): 485-500. Published by Oxford University Press on behalf of the European Society of Human Reproduction and Embryology (ESHRE) online at: <a href="https://academic.oup.com/humupd/article-lookup/doi/10.1093/humupd/dmu009">https://academic.oup.com/humupd/article-lookup/doi/10.1093/humupd/dmu009</a>

#### Evidence from human data of an ED MoA

The association of BPA with altered hormonal levels in women has been investigated in a few epidemiological studies of good quality.

In a prospective study by Mok-Lin *et al.* (2010), which included women (n=84) following an ovarian stimulation protocol as part of an *in vitro* fertilisation, the authors indicated that there was a negative correlation between urinary levels of BPA (n=203 urine samples; 2 samples during 91 cycles and one sample during 21 cycles of IVF) and ovarian response in terms of number of oocytes collected as well as amplitude of the preovulatory estradiol peak. A mean decrease of 12% in the number of oocytes recovered per cycle and of 213 pg/mL from the estradiol peak was observed for each log unit increase of urinary SG-BPA (BPA specific gravity, i.e., the BPA concentration corrected by the urine specific gravity). The BPA levels found were compared to urinary BPA concentrations observed in the general population in the NHANES 2003-2008 cohort. The concentration of urinary BPA found reflects BPA exposure at the time of collection, but not during the period of follicular maturation several months earlier. In addition, it is noted that it may be difficult to extrapolate the results observed in a sample of infertile women undergoing an *in vitro* fertilisation to the general population.

Nevertheless, the results were consistent with those observed in another recent study.

Ehrlich *et al.* (2012) studied the association between urinary BPA concentrations and early reproductive outcomes among 174 women aged 18-45 years representing a total of 237 IVF cycles at a fertility center in Boston, USA. The study was a follow up of Bloom *et al.* (2011), who previously reported an association between urinary BPA and decreased

ovarian response (peak serum estradiol ( $E_2$ ) and oocyte count at the time of retrieval) in women undergoing IVF. After adjustment for age and other confounding parameters (Day 3 serum FSH, smoking, BMI), there was a linear dose-response association between increased urinary BPA concentrations and decreased number of oocytes (overall and mature), decreased number of normally fertilised oocytes and decreased  $E_2$  levels (mean decreases of 40, 253 and 471 pg/ml for urinary BPA quartiles 2, 3 and 4, when compared with the lowest quartile, respectively; p-value for trend=0.001). Women with urinary BPA above the lowest quartile had decreased blastocyst formation (trend test P-value=0.08). The results from this extended study, using IVF as a model to study early reproductive health outcomes in humans, indicate a negative dose-response association between urinary BPA concentrations and serum peak  $E_2$  and oocyte yield.

Despite the fact that these studies were limited to a specific group of women with fertility disorders and despite the inherent limitations of epidemiological studies investigating a substance with short half-life, these studies provide some indications supporting the ability of BPA to alter hormonal regulation.

#### Overall conclusion on differences and commonalities

Differences between rodents and humans in the regulation of cycles are identified in relation to the role of circadian synchronisation and to differences in the timing of ontogeny of the neuroendocrine axis. In contrast the key principles of endocrine mechanisms of regulation of the cycle are the same between rodents and humans. Overall, these elements therefore bring support to the conclusion that the effects of BPA on disruption of cycles are relevant for humans.

In particular, both components that are shown to be involved in the endocrine MoA of BPA on cycle disturbance, i.e. the role of aromatase in estrogen production as well as the role of kisspeptin neurons in the ontogeny of the HPG axis are known to be relevant for humans.

#### 4.2.4 Summary and discussion

In both primates and non-primate mammals, follicle selection, growth, and maturation, as well as ovulation, oocyte quality, and subsequent *corpus luteum* function, all depend on subtle sequential actions of gonadotropins and intraovarian regulators. Furthermore, the ovary and the hypothalamo-pituitary system are in permanent endocrine dialogue with each other. Consequently, any disturbances in the endo/para/autocrine activities of the ovary and/or the hypothalamus-pituitary system lead to cycle disturbance.

In addition, the estrous cycle is a perfectly synchronised and timely event that relies on specific neuroendocrine circuitries. Those pathways differentiate during fetal life and are largely influenced by numerous factors and in particular the steroid environment of the foetus. Thus fetal exposure to steroidogenic compounds is very likely to result in estrous cycle disturbances after puberty.

This review clearly shows that exposure to BPA at the adult stage alters the endocrine steroidogenic function of the ovary and more specifically the production of estrogens by the follicle, potentially leading to disturbance in the estrous cycle. Although most of the reported evidence relies on rodent studies there are *in vitro* data showing the same negative effect of BPA on the estrogen production in the human follicle cells. Furthermore, an indication of a negative association between the ability of the follicle to produce estrogens and exposure to BPA was observed in women. Lastly, the role of estrogens in the maintenance of the cycle is similar in rodents and humans. Thus, we conclude that it is quite likely that BPA may alter the ovarian cycle in humans through the disruption of the endocrine activity of the ovarian follicle.

At the neuroendocrine level, BPA can also act during the perinatal/postnatal organisation or adult activation of the hypothalamus-pituitary system in rodents or primates. Because of the similarities in sex-steroid-induced regulation of this axis between humans and rodents, it is possible that the changes in kisspeptin, GnRH expression, activity or liberation and sex steroid receptor expression induced by developmental or adult exposure to BPA occur also in humans and therefore impact estrous cyclicity.

Table 6 below summarises the documentation supporting the ED-mediated MoA of BPA proposed for each of the two different periods of exposure. It is likely that both MoA may simultaneously contribute to the effect observed during both periods of exposure but the present analysis has been focused on available lines of evidence.

Table 7 provides an overview of the critical elements in the identification of an endocrine disruptor and how they are fulfilled for alteration of estrous cycle by BPA.

Table 6: Summary table of proposed ED-mediated MoA of BPA on alteration of estrous cyclicity and its documentation

Mode of action documented for the respective periods of exposure	Underlying cellular/mo	lecular events	Alteration of organ/function
Alteration of sex steroid ovarian steroidogenesis during adult exposure	Reduced expression and activity of aromatase  - In vitro: Watanabe 2012, Kwintkiewitcz 2010, Mansur, 2016  - In vivo: Lee et al., 2013	Decreased estradiol levels - In vitro: Zhou 2008, Peretz 2011, Mlynarcikova 2005, Kwintkiewitcz 2010, Mansur 2016	Alteration of estrous cyclicity  - In vivo: Lee 2013, Laws 2000, Tyl 2008
		- <i>In vivo</i> : Lee <i>et al</i> ., 2013	
Alteration of neuroendocrine regulation/ development of the reproductive function during developmental exposure	Alteration of kisspeptin expression  - In vitro: Klenke 2016  - In vivo: Cao 2012, Navarro 2009, Xi 2011, Naulé 2014, Kurian 2015	Alteration in GnRH and/or LH secretion  - In vivo: Monje 2010, Fernandez 2009, Viega-Lopez 2014, Kurian, 2015	Alteration of estrous cyclicity  - In vivo: Honma 2002, Nikaido 2004, Wang 2014a, Rubin 2001, Mendoza 2011, Patisaul 2014, Delclos 2014, Nah 2011, Adewale 2009, Fernandez 2009, Zaid 2014

Table 7: Overview of the elements supporting the identification of an alteration of estrous cyclicity as an ED-mediated effect of BPA

	Adverse effect	Plausible ED MoA	Human relevance
Adult exposure	Key study of Lee 2013*	Direct link established with reduction of estradiol production through reduced	Hormonal regulation of cycles highly conserved in mammals

		aromatase activity	Support from <i>in vitro</i> data in human cells Consistent with indications from human data
Developmental exposure	Identified in several studies*: Honma 2002, Nikaido 2004, Wang 2014a, Rubin 2001, Mendoza 2011, Patisaul 2014, Delclos 2014, Nah 2011, Adewale 2009, Fernandez 2009, Zaid 2014	Physiologically-based high plausibility of alteration of kisspeptin expression involved in neuroendocrine control	Role of kisspeptin identified in humans Support from non-human primate data on BPA-MoA

<sup>\*</sup> Acknowledged by RAC

It is well recognised that the effects of BPA on the reproductive function are more diverse in their expression than alteration of estrous cycles as described in the introduction of this section (section 4.3.1). However, the female cyclicity is highly dependent upon hormonal fine regulation and as so it represents a good indicator for the identification of the nature of BPA endocrine MoA on the reproductive function.

Finally, it is noted that RAC in its opinion of March 2014 on classification (ECHA, 2014) provided an analysis of the MoA in the context of analysis of relevance for humans and reached the following conclusion:

"BPA was shown to influence the female reproductive tract. The associated alterations in pituitary signalling, serum hormone concentrations and reproductive organ morphology were likely causes of the reduced female fertility effects reported in the NTP (1985b) and Tyl et al. (2002) oral multi-generation studies and in several non-guideline research studies. Both oestrogenic and anti-oestrogenic effects of BPA were described and not all expected oestrogenic effects were observed. However, RAC concluded that the observed pattern of effects on the female reproductive tract suggest an overall oestrogen-like response in vivo.

Effects on the male reproductive tract, evident as impaired sperm production following BPA exposure, were observed in several studies. The decrease in sperm production was accompanied by lower testosterone levels. The effects observed on the testosterone levels may be the cause of the decreased sperm production.

RAC concluded that the classification of BPA for adverse effects on sexual function and fertility should be based mainly on the results from rodent studies. Disruption of oestrogenic signalling was considered to be the main MoA for the effects of BPA on fertility, based on current knowledge. The hormonal systems are well conserved between mammalian species, and the effects observed in rodents are therefore also relevant for humans. Detection of the active form of BPA (aglycone/unconjugated BPA) has been reported in humans (serum, cord blood and in placenta), but the credibility of these low-concentration measurements has been questioned due to the analytical techniques applied and potential contamination of the samples. However, after oral administration of low doses of stable isotope-labelled BPA (to exclude confounding sample contamination) and using sensitive and specific methodology, low systemic concentrations of aglycone/unconjugated BPA have been reported in rodents and in non-human primates, suggesting that unconjugated BPA becomes bioavailable in primates and rodents after oral exposure. Additionally, RAC noted that other routes of exposure such as sublingual,

buccal, dermal and inhalation exposure are potential routes of human exposure and they bypass the extensive first-pass hepatic metabolism (first-pass effect).

Taken together, RAC considered the MoA to be relevant to humans."

In conclusion, it is generally considered that the effects of BPA result from complex mechanisms that are most probably interacting together. Not all of them are fully understood. However, the overall database shows that an alteration of the regulation of estrogens is an essential pattern of the MoA.

### 4.2.5 Summary tables of studies

In the Table 8 below, studies are grouped by periods of exposure and sub-grouped by tested species (mouse/rat/other species), then chronological order of publication.

Table 8: Summary table of studies investigating the effects of BPA on the estrous cycle in female animals

Referen-	Species	Routes	Dose	Effect on estrous cycle
ce			Exposure period	
	al exposure		1	
Honma <i>et al.,</i> 2002	ICR Jcl mouse	Subcuta- neous	0, 2, 20 μg/kg bw/d GD11- GD17	$\nearrow$ the length of the estrous cycle of 1 day at both dose (p ≤ 0.05).
Nikaido <i>et</i>	CD-1	Subcuta-	0, 0.5, 10 mg/kg	Increased cycle length of 3 days in both
al., 2004	mouse	neous	bw/d GD15 - GD19	groups with increased time of diestrus $(p<0.01)$
Wang <i>et al</i> ., 2014a	FVB mouse	Oral (gavage)	0, 0.5, 20 or 50 µg/kg bw/d GD11-PND0	7 time in diestrus and metestrus and      □ in proestrus and estrus at 0.5 μg/kg     □ in estrus at 20 μg/kg     No significant effect at 50 μg/kg     (     □ time in metestrus and      □ in proestrus with DES)
Tinwell <i>et al.</i> , 2002	Sprague Dawley and Alderley park (derived from Wistar) rat	Oral (gavage)	0, 0.02, 0.1 or 50 mg/kg/d GD6-GD21	No difference in the stage of the estrous cycles at PND90.
Savabieas	Suffolk	Subcuta-	0 or 5 mg/kg bw/d	No effect on the length of progestogenic
fahani <i>et</i> <i>al.,</i> 2006	sheep	neous	GD30 - GD90	cycles during first breeding season but longer breeding season of 1 month ( $p < 0.05$ ).
Perinatal e	exposure	<u> </u>	1	breeding season or 1 month (p < 0.05).
Naulé <i>et</i>	C57BL/6J	Oral	0, 0.05 or 5	Normal cycle analysed after PND60
al., 2014	mouse	(gavage)	mg/kg/d GD15 to PND21	Normal cycle analysed area 11000
Kwon <i>et</i>	Sprague	Oral	0, 3.2, 32 or 320	No effect in 4-month F1 (irregular estrous
al., 2000	Dawley rat	(gavage)	mg/kg/d GD 11 - PND 20	cycle with DES)
Rubin <i>et al.,</i> 2001	Sprague Dawley	Oral (drinking	0, 0.1, 1.2 mg/kg bw/d	Irregular cycles in 79% (4-week old) and 77% (6-month old) F1 at the high dose
ai., 2001	rat	water)	GD6 – end of	(significant). No significant effect at low dose.
			lactation period	Intermittent extended period of diestrus, or extended period of proestrus and/or estrus.

Takagi et al., 2004	Sprague Dawley rat	Oral	0, 7, 70 or 300 mg/kg bw/d (approx.) GD15 – PND10	No effect on estrous cyclicity (some animals with extended diestrus in the low dose group but not significant)
Yoshida et al., 2004	Donryu rat	Oral gavage	0, 0.006 or 6 mg/kg bw/d GD2 - PND21	No effect on estrous cyclicity
Mendoza- Rodriguez et al., 2011	Wistar rat	Oral (drinking water)	1.2 mg/kg bw/d GD6 – PND21	Observations in adult females (3 months) 79% with irregular cycles vs 6% in controls (determined on 4 consecutive weeks) characterised mainly by several continuous estrus days and in few animals persistent diestrus.
Delclos <i>et al.</i> , 2014	Sprague- Dawley rat	Oral gavage (aqueous solution)	2.5, 8, 25, 80, 260, 840, 2700 µg/kg bw/d and 100 and 300 mg/kg bw/d GD6- PND90	Observation between PND 69 to 90: At 300 mg/kg, 7 incidence of animals with abnormal cycles primarily due to extended estrus and extended estrus/diestrus. Between PND150 and 170, 7 incidence of abnormal cycles from 100 mg/kg. Effect similar to EE2.
Ferguson et al., 2014	Sprague Dawley rat	Oral (gavage)	0, 2.5 or 25 μg/kg GD6 to PND21	No effect on the proportion of days spent in each phase (lower proportion of days in diestrus with EE2, extended estrus transitions)
Postnatal			T = 2 +	
Nah <i>et</i> <i>al.,</i> 2011	ICR mouse	Subcuta- neous	0, 0.1, 1, 10, 100 mg/kg bw PND8	unumber of estrus days at the high dose (examined during 9 days from PND20)
Kato <i>et al.,</i> 2003	Sprague- Dawley rats	Subcutan eous	0, 0.25, 1 or 4 mg/pups PND0 – PND9	Observation from PND 61 to 94: irregular estrous cycles in 4/6 females and persistent estrus in 2/6 at 4 mg/kg; (similar effects with 10 µg/kg E2)
Adewale et al., 2009	Long- Evans rat	Subcuta- neous	0, 50 μg/kg bw/d or 50 mg/kg bw/d PND1- PND3	At 50µg/kg: 14% of females were not cycling anymore by 15 weeks after vaginal opening. At 50mg/kg: 67% of females were not cycling anymore by 15 weeks after vaginal opening.  (100% in controls)
Fernande z et al., 2009	Sprague Dawley rat	Subcuta- neous	0, 6,2/2,5, 62.5/25 mg/kg bw/d PND1- PND10	Irregular estrus cycle in adult F at high dose with high prevalence of estrus after PND90 (p<0.05)
Monje et al., 2010	Wistar rats	Subcutan eous	0, 0.05 or 20 mg/kg bw/d PND1-PND7	Observations of females for 2 weeks from PND85:  7 in proestrous/estrous time (p<0.001) at 0.05 mg/kg (high dose not examined for cyclicity)
Franssen et al., 2016	Wistar rats	Subcutan eous	0, 25 ng/kg or 5 mg/kg bw PND1 – PND5 or PND15	Observations from VO to PND 80: no effect on estrous cyclicity
	al exposure		0 10	No effect on the actions were desired as 5 0 0 10
Nikaido <i>et</i> al., 2005	ICR mouse	Subcuta- neous	0 or 10 mg/kg bw/d PND 15 to 18	No effect on the estrus cycle during 5-8, 9-12 and 21-24 weeks of age.
Zaid <i>et</i> <i>al.</i> , 2014	Sprague- Dawley rats	Oral (gavage)	0 or 10 mg/kg/d For 42 days from PND28	Only 3/8 rats with normal cycles (p<0.05) and 5/8 rats with persistent diestrous (p<0.05)
Adult expo				
Moore- Ambritz et al., 2015	C57BL/6 mouse	Oral (gavage)	0 or 50 μg/kg/d For 12-15 days from day of 1 <sup>st</sup> estrus (approx. PND39)	No effect on duration of each estrous stage during the dosing period (lengthened cycle with DES).

### ANNEX XV - IDENTIFICATION OF 4,4'-ISOPROPYLIDENEDIPHENOL (BISPHENOL A) AS SVHC

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Laws <i>et al.</i> , 2000	Long Evans rat	Oral (gavage)	0 or 100 mg/kg for 25 days in cycling	Reduced number of 4-5 day cycles (p<0.05). Extended diestrus in 6 animals, extended
			animals	estrus in 2 animals, normal cycles in 7
				animals.
				(similar effects but more pronounced with E2)
Lee et al.,	Sprague-	Oral	0, 0.001 or 0.1	Observations of female for 30 days after the
2013	Dawley	(gavage)	mg/kg bw/d	90-day exposure:
	rat		8-week animals	¬ in duration of the estrus phase: p<0.001 at
			exposed for 90 days	0.001 mg/kg, p<0.01 at 0.1 mg/kg and p<0.05 with EB
				No animal in a persistent estrus phase.
Multi-gen	eration stud	dies		
NTP,	CD-1	Oral	0, 300/350, 600/650	No effect on estrous cycle
1985	mouse	(diet)	or 1200/1300	
			mg/kg/d (M/F)	
			1 week before	
			mating until sacrifice	
			(continuous	
			breeding; 2 G)	
Tyl et al.,	CD-1	Oral	0.003, 0.03, 0.3, 5,	F0 treated females were twice more in estrus
2008	mouse	(gavage)	50 or 600 mg/kg/d	as compared to controls at 600 mg/kg
			8 weeks before	
			mating to adulthood	
Ema et	Sprague-	Oral	(2 G) 0, 0.2, 2, 20 or 200	No effect
al., 2001	Dawley	(gavage	μg/kg/d	NO CITECO
2, 2001	rat	until	10 (M) or 2 (F)	
		weaning	weeks premating	
		(2 G)		
Tyl et al.,	Sprague-	Oral	0.001, 0.02, 0.3, 5,	No effect on estrous cycle length.
2002	Dawley	(diet)	50 or 500 mg/kg/d	
	rat			
			10 weeks before	
			mating until PND21	
			(3 G)	

Table 9: Summary of the *in vitro* studies showing an alteration of the ovarian steroidogenesis function likely to result in disturbance of estrous cyclicity if occurring *in vivo*.

Cell type	Reference	Tissue	Concentration	Results	Conclusion	
Theca-	Peretz <i>et al.</i> , 2011	Mouse isolated antral follicles	100 and 10 mg/L (=440 and 44 μM) BPA	↓ Testo, ↓Androstenedione, ↓DHEA-S, ↓ STAR, ↓CYP450scc, → 3 ß-HSD, → CYP450 17apha		
interstitial cells Zhou et al., 2008		Theca-interstitial cells isolated from immature SD rats			No clear-cut conclusion	
	Zhou <i>et al</i> ., 2008	Granulosa cells isolated from mature SD rats	1 to 100 μM BPA	↓ estradiol		
	Peretz <i>et al.</i> , 2011	Mouse isolated antral follicles	100 and 10 mg/L (=440 and 44 µM) BPA	↓ estradiol		
	Mlynarcikova <i>et</i> <i>al.</i> , 2005	Porcine granulosa cells from antral follicles	1 to 100 μM BPA	↓ FSH-induced estradiol	BPA reduces CYP450arom expression and estrogen	
Granulosa cells	Watanabe <i>et al.</i> , 2012	KGN (a human granulosa-like cell line)	5 to 100 μM BPA	↓ CYP450arom	production in rodents, domestic animals and human granulosa cells in all studies.	
		KGN and human granulosa cells	40 to 100 μM BPA	↓ FSH-induced CYP450arom, ↓ estradiol, ↓ IGF-I, ↓ GATA-4, ↓ SF-I, ↑ PPAR-gamma,	cells III dii studies.	
	Mansur <i>et al.</i> , 2016	Human granulosa cells	8.8 to 88 µM BPA	↓ estradiol, ↓ progesterone ↓ Cyp arom(mRNA and protein)		

Table 10: in vivo evidence for endocrine mechanisms potentially underlying alteration of estrous cyclicity as a consequence of adult exposure to BPA

Refe- rence	Species Age	Route (dose) Duration	Effect	Evidence for ED MoA
Lee <i>et al.</i> , 2013	Rat SD PND 56	Gastric: - BPA: 1 and 100 µg/kg bw/d - EB: 1 µg/kg bw/d 90 days	Disruption of the estrous cycle with extended estrous phase of 2-7 days with both BPA doses and EB	Both BPA doses and EB:  ↓plasma estradiol, ↓testosterone,  ↓STAR,  ↑apoptosis in follicles and corpus lutea.  → FSH  Both BPA doses but not EB:  ↓aromatase in granulosa cells  ↓ uterine estrogen-induced proteins (PCNA calbindin-D9k)  ↓ uterine collagens  ↑plasma and pituitary LH  concentrations
Wang <i>et al.</i> , 2014b	Mouse ICR Adult in pro- estrous	Oral (20 µg/kg) or ICV (0, 0.02, 0.2, 2.0, 20.0, and 200.0 nM/3 ml) Single exposure Analysis 6 hours after exposure	↑ GnRH and Kiss 1 expression ↑LH, FSH, E2	Effects blocked by GPR54 and ERβ antagonists Estrous cyclicity not monitored but effects very likely to be related to disruption of estrous cyclicity
Kurian <i>et</i> <i>al</i> ., 2015	Rhesus monkey Pubertal female (approx. 38 months)	Infusion of 0, 0.1, 1 or 10 nM BPA into stalk- median eminence of the hypothalamus for 240 min Simultaneous collection of dialysate	↓ GnRH and kisspeptin release, ↑ pulse amplitude interval at 10 nM	Alteration of GnRH secretion

Table 11: Summary of the *in vitro* study showing an alteration of GnRH activity by BPA

Reference Tissue and treatment period		Type of evaluation	Type of modification	Evidence for PED MoA	
Klenke <i>et</i> <i>al.</i> , 2016	Nasal explants from mice embryos at embryonic day 11.5 (after emergence of GnRH cells and other neuronal cell types from the plasma codes) used after 6 days of culture – 50 µM	Calcium imaging	BPA reduces the frequency of oscillations in GnRH neurons	Direct effect of BPA on GnRH neurons	

Table 12: Evidence for BPA-induced disturbance of estrous cyclicity in animal models and

link with an endocrine disruptive MoA

link with an er	Species	Route	Type of	Type of	Evidence for ED MoA
iterer en ee	Period	Doses	evaluation	modification	27.00.000 101 22 11071
Rubin <i>et al.</i> , 2001	Rat SD GD6- PND21	Oral (drinking water) 0.1-1.2 mg/kg/d	Daily vaginal cytology for 18 consecutive days at 4 and 6 months  8 female offspring/ group were ovariectomised and killed 3 months later to assess LH levels.	↓% of animals with regular estrous cycle ↑ duration of the cycle and ↓ nb of estrous cycles/animal with 1.2 mg/kg/d	↓LH secretion in BPA- treated OVX animals showing a BPA-induced alteration in the endocrine function of the hypothalomo- hypophysis gonadotropic axis. But LH was not evaluated in ovary intact animals
Nikaido <i>et al.</i> , 2004	Mouse CD1 GD15- GD19	Subcutan eous 0.5-10 mg/kg/d	Daily vaginal smears between 9 to 11 weeks	↑mean cycle length ↑time in diestrus	For diestrus: similar increase with DES
Adewale <i>et</i> <i>al</i> ., 2009	Rat Long- Evans PND0- PND3	Subcutan eous 50 - 50000 µg/kg/d sc	Daily 4 days vaginal lavage from 2 weeks after VO and every two weeks for 13 weeks	Time and Dose- dependent decrease of the number of cyclic females	Similar but more rapid and more sustained effect with PPT a selective agonist of ERa
Monje <i>et al.</i> , 2010	Rat Wistar PND1- PND7	Subcutan eous 0.5 or 20mg/kg /d	Daily vaginal smears from PND85 to PND 100 performed in the group exposed to 0.05 mg/kg	↑time spent in proestrus/ estrus at 0.05 mg/kg (not examined at 20 mg/kg)	Females at 20 mg/kg were incapable of producing the LH surge. Alteration of GNRH maturation process and ER $\alpha$ expression in the AVPV and ARC, and PR in the AVPV
Fernandez <i>et al</i> ., 2009	Rat SD PND1- PND10	Subcutan eous ~5-50 mg/kg/d	PND90	Irregular estrus cycles (persistent estrus)	↓pituitary sensitivity to GnRH in estrus in vivo and in vitro ↑GnRH pulse frequency in juvenile and adults
Wang <i>et al</i> ., 2014a	Mouse FVB GD11- PND0	Oral (gavage) 0.5-20- 50 µg/kg/d	From PND21-51 Daily Vaginal smears	0.5 µg/kg/d ↓poestrus and estrus ↑metestrus 20 µg/kg.d: ⋈ estrus Not with high dose 50µg/kg/d	For proestrus and metestrus: idem DES (ref Berger <i>et al.,</i> 2016 for PE mechanisms)

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Delclos <i>et al.</i> , 2014		Oral (gava ge) 2.5, 8, 25, 80, 260, 840, 2700, 100000, 300000 µg/kg/d	Daily vaginal cytology from PND 69 to 90 and from PND 150 to 170	↑% female with extended estrus (highest dose only)	EE2-like effect ↑E2 and PRL- ↓progesterone
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Table 13: Studies addressing the effects of developmental exposure to BPA on the kisspeptin/GnRH system

Reference	Animal model	Doses (route)	Period	Effects on Kisspeptin	Effects on GnRH
Cao <i>et al</i> . (2012)	Long-Evans rats	50µg/kg - 50mg/kg (SC)	PND0-PND2	Diminished Kiss1 expression (mRNAs) in the RP3V at PND10. No modification of Kiss1 in the ARC	
Adewale et al. (2009)	Long Evans rats	50µg/kg – 50mg/kg (SC)	PND0-PND3		No modifcation in GnRH expression in the OVLT
Fernandez et al. (2009)	Sprague- Dawley rats	50µg/50µL - 500µg/50µl (SC)	PND1- PND10		Increased infantile GnRH pulsatility. During adulthood, diminished GnRH-induced secretion of LH and disturbed pulsatility
Navarro <i>et al</i> . (2009)		100 - 500µg/rat (SC)	PND1-PND5	Diminished Kiss-1 mRNAs levels in the hypothalamus at PND30	
Patisaul <i>et al</i> . (2009)	Long Evans rats	50μg/kg – 50mg/kg (SC)	PND1-PND4	No effect of BPA exposure in the AVPV, decrease in the ARC at the higher dose. Diminished kisspeptin cell density in the AVPV and ARC by with ERalpha agonist.	
Monje <i>et al</i> . (2010)	Wistar rats	0.05, 20 mg/kg (SC)	PND1-PND7		mRNAs GnRH increased at BPA-0.05 mRNAs GnRH decreased at BPA-20
Losa-Ward et al. (2012)	Wistar rats	50µg/kg - 50mg/kg (SC)	PND0-PND3	No differences in kisspeptin cell density in the AVPV or kisspeptin-GnRH appositions. Diminished cell density and number of RFRP3 neurones, and RFRP3-GnRH appositions	
Franssen et al. (2016)	Wistar rat	25 ng, 25 mg, 5 mg/kg/d (SC)	PND1- PND15		GnRH IPI increased by 25 ng, reduced by 5 mg
Xi <i>et al.</i> (2011)	CD1-mice	12 – 25 - 50 mg/kg (oral)	G1-PND49 or PND21- PND49	G1-PND49: Increased Kiss1mRNA expression in the hypothalamus at BPA- 25 and 50	G1-PND49: increased GnRH expression levels in the hypothalamus at BPA-25 and 50. No modification in GnRH-R expression levels in the pituitary
Naulé <i>et al</i> . (2014)	C57BL6J mice	0,05-5 mg/kg/d (oral)	GD15- PND21	Increased kisspeptin number in the RP3V of adult females	

### 4.3 Alteration of mammary gland development

# 4.3.1 Overview of previous evaluation of BPA's effect on mammary gland

Effects of BPA on mammary gland have already been assessed in previous European reports (ANSES, 2014; EFSA, 2015; ECHA, 2015). The main outcomes of these evaluations are reported below.

**The EFSA (2015) opinion concluded**: "The proliferative responses and possibly enhanced sensitivity to mammary gland carcinogens seen in animal studies might be of relevance for human health and are therefore included in the risk assessment." and "the CEF Panel concluded that BPA-induced effects on the mammary gland of rats, mice or monkeys exposed pre- or perinatally were "likely" effects".

However, EFSA considered none of the available studies to be sufficiently robust in terms of methodology, or a consistent dose-response for deriving a health-based guidance value based on mammary gland effects.

See sections 3.9 and 4.3 of the EFSA (2015) opinion for more details."

In the restriction BPA dossier (ANSES, 2014), on thermal paper, ANSES considered that the effects of BPA on the mammary gland were "recognised" effects in animals and should be taken into account to assess the risk to human health. ANSES observed that EFSA's draft opinion also considered that the effects of BPA on mammary gland development are "likely" and that these effects are relevant to humans.

ANSES considered that it is important to take into account the possibility of increased cancer risk in the children of women who have a high level of endogenous estrogens or xeno-estrogens during pregnancy and are then exposed to tumour initiating agents. Based on the studies described later in the opinion of RAC (ECHA, 2015), RAC concluded that "the Dossier Submitter considered ductal hyperplasia and effects on the architecture of the mammary gland, including effects on Terminal End Buds (TEB) as critical effects for the human risk assessment. For effects on these undifferentiated epithelial structures (Terminal Ducts (TD and TEB), an oral NOAEL of 25  $\mu$ g/kg bw/day and a LOAEL of 250  $\mu$ g/kg bw/day were proposed by the Dossier Submitter based on Moral et al. (2008).

In its opinion of **June 2015 (ECHA, 2015),** RAC adopted the following conclusions in relation to the analysis of the effect of BPA on mammary gland:

"The overall qualitative conclusion of RAC regarding the mammary gland changes is that BPA caused an acceleration of mammary gland maturation in experimental animals. There are slight indications of relevant intraductal hyperplasia from two studies with subcutaneous exposure (Murray et al., 2007 and Vandenberg et al., 2008).

#### Conclusion

RAC agrees that BPA has been shown to have a proliferative effect on mammary tissue at doses below the doses causing general toxicity (such as kidney weight changes). RAC in principle agrees with EFSA's conclusion on mammary gland effects. The effects on mammary gland development should be taken into account in hazard and risk assessment and in health impact assessment. In line with EFSA (2015), no individual study is however considered robust enough by RAC to serve as critical study for the identification of a starting point for DNEL derivation. Therefore the effects will be accounted for in the setting of Assessment Factors.

The Dossier Submitter derived a LOAEL of 0.26 mg/kg bw/day based on increased body weight and increased cholesterolemia in female mice in Miyawaki et al., 2007."

The evidence for the effects of BPA on mammary gland presented above and considered for the risk assessment included in the restriction dossier were supplemented with recent *in vivo* and epidemiological studies investigating "BPA and breast cancer risk" published until May 2016. The *in vitro* key experimental studies were also collected in order to substantiate the ED MoA and plausible link between adverse effects and the MoA of BPA.

In a first section, background information is provided. In the following sections, the available evidence on adverse effects on the mammary gland are described, together with the data showing that these effects are related to an ED MoA in a causal way.

### 4.3.2 Background information on mammary gland development

As indicated above, this review will address the effects of BPA during the mammary gland development following early life exposure as well as long lasting effects including hyperplasic lesions after fetal / perinatal BPA exposure. Experimental *in vitro* and *in vivo* findings including recent results from epigenetic studies generated since the evaluation quoted above, have been considered in this dossier. All this information is developed in the following sections.

The following information is given in order to facilitate comprehension of the experimental mechanistic studies available on BPA. The mammary gland develops in sequential steps that will be described below. The hormonal implication, mammary epithelial estradiol signaling, and progesterone (PR) and prolactin (PL) receptor involvement depending on the developmental period are presented. There is a summary of key mechanisms that have been described after  $\beta$ -estradiol as well as DES exposure.

Lastly, relevant background information on both epigenetic mechanisms and HOX genes that are key players in embryogenesis and post-natal development, and during the early stages of neoplastic process, are also reported in the section below.

#### 4.3.2.1 Mammary gland structure and its sequential development

Critical events in breast development begin during fetal life with epithelial bud sprouting, whereas extensive branching morphogenesis continues into postnatal life.

Rapid mammary gland development occurs <u>in three distinctive life stages</u>: fetal, peripubertal, and pregnancy (Fenton, 2006). Development of the mammary glands is initiated in the embryo with a major part occurring in adult stage. While development in puberty and pregnancy is hormone-dependent, early postnatal development appears to progress without hormone activation.

#### • Prenatal and early postnatal development:

Epithelial-mesenchymal interactions are critical during development. Formation of the bud, the first appearance of hormone receptors, formation of the primary sprout and ductal elongation have been shown to be regulated by epithelial-mesenchymal signaling. The "primary mammary mesenchyme" is distinguished by the expression of specific genes among them those coding for steroids, namely androgen and estrogen receptors, and for PPT-A (Robinson *et al.*, 1999). Some of the signaling molecules that are required in these processes such as LEF1 or PTHrP have been identified through gene inactivation (Robinson *et al.*, 1999).

#### • Peri-pubertal and adult period:

A rudimentary ductal system present at birth begins to unfold during puberty and gains in complexity during adulthood with recurrent hormone stimulation during menstrual/estrous cycles. Ductal complexity increases further during pregnancy and finally secretory structures of saccular shape, called alveoli, bud all over the ductal system.

As an illustrative example the period around puberty in mice, i.e. between 20 and 30 days of age, is characterised by the reinitiation of ductal growth in the mammary glands with expansion of the epithelial tree into the surrounding stromal tissue (see Figure 10). Estrogens play a major role in this process (Topper and Freeman, 1980). Estrogen-induced cell proliferation during puberty concentrates at the ductal tips. As a result, they enlarge and form club-like structures that measure between 1.5 to 10 times the diameter of the subtending ducts and are called TEBs. These structures invade the stroma and mediate the longitudinal growth of the subtending ducts. When the ductal tree reaches the edge of the fat pad, the TEBs mature into a resting structure, the terminal ends, and ductal growth ceases. In adulthood, the ductal complexity increases through progesterone-induced side branching. Side branching is enhanced during early pregnancy. Alveoli develop later in pregnancy and will be fully distended by milk during lactation.

This morphological differentiation is sustained by high cellular activities. At puberty, as the TEBs become bulbous, they show both high proliferative and high apoptotic activity. A large number of apoptotic cells are observed in TEBs at PND30 in mice. Death of the body cells is essential for the formation of the lumen on the proximal side of the TEBs (Humphreys *et al.*, 1996) and for the growth of the subtending duct (Munoz de Torro *et al.*, 2005).

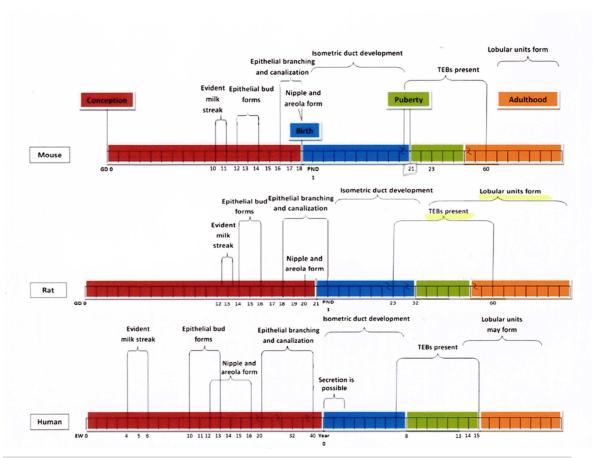
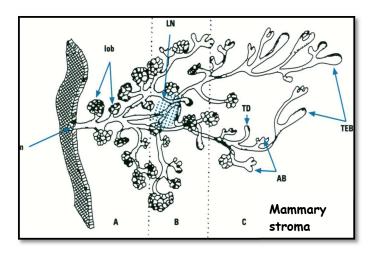


Figure 10: Mammary development in different species (mice, rat, humans) quoted from Davis and Fenton, 2013

**Parenchyma:** constituted of one or two major lactiferous ducts that grow from the nipple into the surrounding fat pad and of terminal epithelial structures such as Terminal End Bud (TEBs).

Stroma: constituted of conjunctive, adipose tissues and of blood vessels



Schematic representation of the fourth (abdominal) mammary gland of a 55 day-old virgin female rat (according to Russo et Russo, 1980 and 1996)

#### Legend:

The gland is divided into three areas:

- Zone A:
  proximal to the
  nipple (n),
  contains more
  numerous lobules
- Zone B: medial, encompasses the lymph node (LN).
- Zone C: distal to the nipple, contains the majority of actively growing terminal end buds

Figure 11: Schematic representation of the fourth (abdominal) mammary gland of a 55-day-old virgin female rat (according to Russo and Russo, 1980 and 1996)

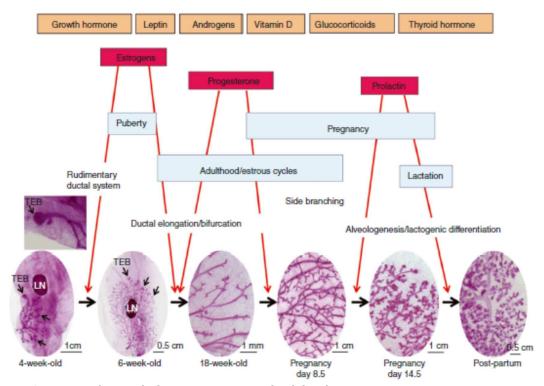


Figure 12: Hormonal control of mouse mammary gland development.

**Legend:** Whole-mount micrographs of inguinal mammary glands of C57Bl6x129Sv mice at different stages of mammary gland development illustrate critical stages in mammary gland development. Corresponding reproductive stages are depicted by light blue boxes. In the red boxes the key female reproductive hormones. Arrows indicate at which stages the respective downstream receptor mediated signaling is limiting. The orange boxes at the top indicate other endocrine factors that have been implicated in mammary gland development but whose precise role remains to be determined. During puberty, the rudimentary ductal tree elongates under the influence of estrogen until the edges of the fat pad are reached and simple ductal system is established through bifurcations. Terminal end buds (TEBs) (arrows) are characteristic of the pubertal stage. In adulthood, the ductal complexity increases through progesterone-induced side branching. Side branching is enhanced during early pregnancy. Alveoli develop later in pregnancy; they will be fully distended by milk during lactation. LN, lymph node.

Cited from Brisken and Ataca, 2015.

### 4.3.2.2 Estrogen, progesterone and prolactin hormones

A large number of mouse mutant strains are available, and tissue recombination experiments allow the generation of epithelial specific mutants. This approach has revealed that mammary epithelial intrinsic ERa signaling is required for pubertal ductal elongation (Mallepell *et al.*, 2006).

The progesterone receptor (PR) is essential in the mammary epithelium for side branching and alveologenesis (Brisken *et al.*, 1998), whereas the epithelial prolactin receptor is required for alveologenesis and milk secretion (Brisken *et al.*, 1999) (see also Figure 12, from Brisken and Ataca, 2015). A small amount of budding of alveolar structures develops during each menstrual/estrus cycle due to the stimulus of ovarian hormones (Russo *et al.*, 2001). During pregnancy and lactation, the number of lobuloalveolar structures increases exponentially under the control of the prolactin pituitary hormone (Brisken *et al.*, 1999; Brisken and Rajaram, 2006).

The mammary epithelium responds differently to a hormonal stimulus depending on its developmental stage. Hormone ablation and replacement experiments have shown that 17- $\beta$ -estradiol induces cell proliferation specifically in pubertal (Daniel *et al.*, 1987) but not in adult mammary glands. In the adult, i.e., more than 8-week-old, female mouse 17- $\beta$ -estradiol pretreatment induces the expression of the progesterone receptor (Haslam and Shyamala, 1979), whereas subsequent stimulation with progesterone triggers cell proliferation (Grimm *et al.*, 2002). Hence, in the adult female, PR signaling is the major stimulus of cell proliferation. Lastly, Fenton (2006) emphasises that dose levels and timing (windows of sensitivities) to EDC exposures may affect the severity, or lack thereof, of an effect on mammary gland growth and consequently breast cancer risk.

# 4.3.2.3 Molecular changes recently suspected to be involved in the process of carcinogenesis

Epigenetics describes a range of DNA and histone modifications that influence levels of gene expression without modifications to the underlying coding sequence (Goldberg *et al.*, 2007). Such modifications include DNA methylation (DNMT), histone modifications (such as the polycomb PRC2) and non-coding RNA. Epigenetic changes have been implicated in the process of carcinogenesis (Esteller, 2007; Jones and Baylin, 2007). Recent studies have shown that abnormal epigenetic silencing of genes can frequently occur during the early stages of the neoplastic process, such as the preneoplastic stages of breast carcinogenesis (Feinberg *et al.*, 2006, Baylin and Ohm, 2006, Fernandez *et al.*, 2012, Faryna *et al.*, 2012).

The EZH2 gene has been shown to be upregulated by estradiol, in an *in vitro* and *in vivo* study conducted by the Mandal team on rat mammary glands (Bhan *et al.*, 2014a). Its expression is modulated through an epigenetic mechanism. EZH2, an enzyme histone methyltransferase, promotes proliferation and neoplastic transformation of breast epithelial cells (Kleer *et al.*, 2003, Collett *et al.*, 2006). EZH2 is linked to two Polycomb repressive complex, PRC2/3 (which are members of the EZH family of histone methyltransferase and initiate gene silencing), and PRC4 which is expressed in cancer and embryonic stem cells (Baylin and Ohm 2006). The polycomb complex PRC2 controls cell fate, development and cancer (Sparman and van Lohuizen 2006, Margueron and Reinberg, 2011). EZH2 has been described as a molecular marker for a precancerous state in morphologically normal breast tissues and preneoplastic progression in the breast (Ding and Kleer, 2006). Lastly, two important genes are targeted by EZH2 namely p57 (CDKN1C), a cyclin dependent kinase inhibitor, and E-cadherin which is important in cell-cell adhesion and migration. Repression of p57 and E-cadherin by EZH2 may lead to the increase of cell proliferation, or increased invasiveness, seen in some breast tumors.

HOTAIR (HOX antisense intergenic RNA) coordinates with chromatin modifying enzymes and regulates gene silencing. It is overexpressed in various carcinomas including breast cancer. HOTAIR is crucial for cell growth and viability and its knockdown induced apoptosis in breast cancer cells. Bhan *et al.*, 2013 showed that HOTAIR is transcriptionally induced by estradiol (E2). Its promoter contains multiple functional estrogen response elements (EREs). Similar to protein-coding gene transcription, E2-induced transcription of antisense transcript HOTAIR is coordinated *via* ERs and ER coregulators, and HOTAIR overexpression is linked to invasiveness and metastasis in several cancers (Bhan *et al.*, 2013).

The HOX genes are an evolutionary conserved family of genes that are key players in embryogenesis and post-natal development (Mallo and Alonso, 2013). Several HOX genes control development of the mouse mammary gland in response to pregnancy (Chen and Capeccchi, 1999). Among them, HOXC6 and HOXB9 are associated with mammary gland development. HOXC6 null female mice show complete absence of mammary epithelium in thoracic and dilated ducts in inguinal glands (Garcia-Gasca and Spyropoulos, 2000). HOXB9 is involved in cell proliferation, cell-cycle progression, and differentiation. The overexpression of

HOXC6 or HOXB9 increases the expression of growth factors (Shrestha *et al.*, 2012). Many HOX genes also appear to play critical roles in tumor cell proliferation and metastasis (Friedman *et al.*, 1994; Chen and Sukumar, 2003). HOXB9 overexpression alters the tumor microenvironment and promotes breast tumor metastasis, associated with clinical outcome in patients (Hayashida *et al.*, 2010; Seki *et al.*, 2012).

#### 4.3.3 Adverse effect

As indicated previously, exposure to an estrogen-like compounds (xenoestrogen) during critical stages of development can interfere with hormonal signaling and may result in persistent morphological abnormalities and altered gene expression (Fenton *et al.*, 2012; Reed and Fenton, 2013; Macon and Fenton, 2013).

Several findings prompted researchers to hypothesise that fetal exposure to xenoestrogens may play a role in the risk of contracting breast cancer observed in the last 50 years. First, a positive association was observed between increased intrauterine levels of estrogens (a phenomenon observed in twin births) and risk of breast cancer in daughters born from such pregnancies (Ekbom et al., 1992) which supports the link between perturbation in the fetal environment and breast cancer. Secondly, epidemiological data also revealed an increased incidence of breast cancer in women exposed in utero to DES (Hoover et al., 2011; Palmer et al., 2002 and 2006). DES has been shown to cause epigenetic changes in HOXA10 expression in the reproductive tract by altered DNA methylation (Bromer et al., 2009 and Bromer et al., 2010). Epigenetic changes as a result of exposure may predispose to malignancies in adulthood.

Fetal exposure to low doses of BPA alters the tissue organisation of fetal mammary gland in rodents and non-human primates, *i.e*, during the period of exposure. These changes include increased ductal area, changes in the periductal stroma and maturation of the fat pad in exposed fetuses (Vandenberg *et al.*, 2007; Tharp *et al.*, 2012), see Table 14.

In addition to an immediate effect on mammary gland development during the period of exposure (fetal or fetal/perinatal exposure) at relevant doses of BPA (25 µg/kg bw/day, by oral or sub-cutaneous routes) in rodent models, BPA leads to effects throughout post-natal life (ANSES, 2013). These mammary gland changes were observed during peri-puberty in mice (PND30) (Markey et al., 2005; Munoz de Toro et al., 2005; Ayyanan et al., 2011), and in different rat strains (PND50) (Murray et al., 2007; Durando et al., 2007; Moral et al., 2008; Jenkins et al., 2009). Mammary gland changes concern ductal elongation, an increased number of TEBs relative to the ductal area and fewer apoptotic TEB cells compared to the controls, increased lateral branching (Munoz de Toro et al., 2005) and ductal hyperplasia. Increased cell proliferation and decreased apoptosis were observed in the glandular epithelium. Ductal (and occasionally lobuloalveolar) hyperplasia have also been described during both the peripubertal period or later on (*Murray et al., 2007; Acevedo et al., 2013*), possibly associated with disorganisation of periglandular stroma (Maffini et al., 2004; Durando et al., 2007; Fischer et al., 2016).

In utero and perinatal exposure of rats and mice to environmentally relevant doses of BPA has also been linked to the development of intraductal hyperplasia that appears during adulthood (over PND100) (Durando et al., 2007; Vandenberg et al., 2008; ANSES 2013), as well as to an increased number of carcinogen-induced tumors (DCIS) in rats (Durando et al., 2007; Jenkins et al., 2009 and 2012; Betancourt et al., 2010) (Table 16). Intraductal hyperplasia ('ductal beading' representing the merging of actively proliferating luminal epithelial cells) is considered to be linked with precursors (pre-neoplastic lesions) of breast carcinoma in rodents and humans (Russo et al., 1990; Russo and Russo, 1996; Vandenberg et al., 2007). The increased tumor response from carcinogen exposure early in life is attributed to the presence of proliferating and undifferentiated structures such as TEB, which are present during the pubertal mammary epithelial expansion.

BPA increases the susceptibility of adult mammary gland to subsequent exposures to chemical carcinogens. Briefly, when BPA exposure is coupled with a genotoxic carcinogen, either DMBA or NMU, several studies reported a significantly decreased tumor latency and increased mammary tumor multiplicity (ANSES 2013; for recent review Seachrist *et al.*, 2016). The most reported effects occurred upon exposure to high doses of BPA (250 µg/kg bw/day, orally or sub-cutaneously administered) either following *in utero* or pre-pubertal exposures (Jenkins *et al.*, 2009 and Betancourt *et al.*, 2011). Yet, an increase in tumor susceptibility was also reported with low dose BPA: 25 µg/kg bw/day (Acevedo *et al.*, 2013).

The effects of chronic exposure to BPA from gestation day 6 (GD6) on rat mammary glands were studied by Delclos et~al.~(2014) in juvenile and adult animals. Ductal hyperplasia, as defined by the authors, was observed in this study as well as one mammary adenocarcinoma at the lowest tested dose level only (2.5  $\mu$ g/kg bw/day, oral route). Some methodological uncertainties (which, in particular, limit the sensitivity of this study) made it difficult to assess the doses from which BPA causes hyperplastic ductal lesions.

In conclusion, there is substantial evidence from rodent studies indicating that early-life BPA exposures lead to increased susceptibility to mammary cancer.

### 4.3.4 MoA (see Figure 13 and Figure 14)

4.3.4.1 Cellular effects: Epithelio-mesenchymal transition (EMT), proliferation and migration

Data presented above on adverse effect describes how fetal exposure to low doses of BPA alters the tissue organisation of fetal mammary gland in rodents and non-human primates, causing effects such as increased ductal area, and changes in the periductal stroma and maturation of the fat pad in exposed fetuses (Vandenberg *et al.*, 2007; Tharp *et al.*, 2012).

• Epithelio-mesenchymal transition (EMT)

Studies using transcriptional analyses on the stromal and epithelial compartments isolated from the fetal female mammary gland (mouse) demonstrate that low dose BPA exposure in dams alters the mesenchymal and epithelial transcriptomes (Wadia *et al.*, 2013; see Table 14). Changes in gene expression in the BPA-fetal exposed mammary gland are related to proteins involved in apoptosis (increased expression of the anti-apoptotic gene, *Birc2*, *Abl1*), myoepithelial differentiation (increase of *Krt8*) and changes in the composition of extracellular matrix (ECM) and in adipogenesis (see Table 14). Several other signaling pathways are affected by exposure to BPA, such as genes involved in EMT (Betancourt *et al.*, 2014), apoptosis and immune function (Moral *et al.*, 2008); see Table 15). While E2 is known to modulate EMT through ERa, the estrogen-receptor involved in BPA-induced EMT is still unknown.

• In vitro effects of BPA: proliferation, migration see Table 16

The effect of BPA on cellular proliferation, senescence and migration was studied in human mammary epithelial cells (HMEC, a model system for studying early events in mammary tumorigenesis). Changes in cellular proliferation and senescence were observed in HMEC treated with BPA (and E2), and associated with dysregulation of proteins such as Bcl2, cyclin D1 and cyclin E (Lee *et al.*, 2012; Qin *et al.*, 2012).

BPA induced also an increase in proliferation using the normal-like human breast epithelial cell line MCF-10F. Activation of proliferation and increased level of estrogen-responsive gene ps2

by BPA (10  $\mu$ M) showed the engagement of ERa and the co-activator SRC3 to a similar level as the positive control group (E2 was tested at 10 nM) (Sengupta *et al.*, 2013). Another study has reported that GPER is required for growth effects and migration in breast cancer cells (SKBR3) and cancer-associated fibroblasts (CAFs) that lack the classical ER (Pupo *et al.*, 2012).

• Disruption of acini by BPA: an estrogenic effect demonstrated in an in vitro 3D model (EMT) (see Table 16)

The effects of BPA on the morphogenesis/disruption of breast glandular structures was also investigated using *in vitro* 3D models for breast glandular structure development, using nontransformed breast epithelial MCF-10F and MCF-12A cells, cultured in a reconstituted basement membrane matrix (Matrigel) (Fernandez and Russo, 2010; Marchese and Silva, 2012). Unlike monolayer cultures, immortalised mammary epithelial cells grown in 3D recapitulate numerous features of the breast epithelium *in vivo*, including the formation of duct-like structures and growth arrested polarised acini with hollow lumen. These well-organised acinar structures reproduce important features of malignant transformation and breast development observed *in vivo*, i.e., the events that trigger disruption of these structures and potentially lead to breast cancer.

MCF-10F cells grown in normal conditions formed duct-like structures in collagen ressembling the ducts of the mammary gland, but did not form colonies in agar model (3D culture); BPA altered the ductular pattern (wider and larger) in the collagen matrix. BPA decreased the formation of tubules and increased the formation of spherical masses in the collagen matrix, similarly to E2, compared to the control DMSO group. Interestingly, the number of solid masses after BPA treatment may even be significantly higher than in cells treated with E2 (Fernandez and Russo, 2010).

MCF-12A cells are estrogen receptors ERa, ER $\beta$  and GPER competent, allowing the investigation of the effects of BPA on mammary gland formation and disruption (Marchese and Silva, 2012). Under normal conditions, MCF-12A cells form organised acini, with deposition of basement membranes and hollow lumen. However, treatment with BPA (10  $\mu$ M) or estradiol, resulted in deformed acini and filling of the acinar lumen, similarly to E2 (1 nM). When BPA treatment was combined with ER and GPER inhibitors, the deformed acini recovered normal features, such as a spheroid shape, proliferative arrest and luminal clearing, suggesting a role for ER and GPER in the estrogenic disruption of acinar formation. Interestingly, a similar outcome was not observed in ERa negative MCF-10A cells treated with E2.

## 4.3.4.2 Changes in estrogen-dependent susceptibility: gene transcription following *in utero* and postnatal exposure (see Table 14 and Table 15)

Transcriptional effects of BPA were also compared with those of EE2, a potent estrogen. Hierarchical clustering analysis suggests that BPA and EE2 similarly affect the transcriptional response of epithelial cells, whereas the response to these agents was different in peri-ductal stromal cells (Wadia et al., 2013). These observations support a model whereby BPA (and EE2) act directly on the stroma of the fetal gland, which expresses ERa, ER $\beta^{14}$  and GPER30 (at GD 18 observation time in Vandenberg et al., 2007, at birth observation time in Wadia et al. 2013). In turn, stroma affects gene expression in the epithelium, even if ERa and ER $\beta$  are below the level of detection at this stage of development. These results indicate that BPA alters gene expression in the BPA-exposed mammary gland during fetal development through

68 (239)

 $<sup>^{14}</sup>$ Note: ERa KO mice display multiple significant defects in reproduction and mammary gland development, ER $\beta$  KO phenotypes are more limited, and GPER KO exhibit no reproductive deficits.

estrogen receptor(s), acting as an estrogenic agent (Wadia et al., 2013).

In addition to modifications observed at fetal stage, modifications on ER are also observed during peripuberty.

- Some studies indicate that mice and rats, after *in utero*/lactational exposure to BPA, have a significantly higher sensitivity to estradiol during peri-puberty, as indicated by histoarchitecture changes and by an increase of E2-dependent gene expression such as amphiregulin (AREG), rather than changes in the ER expression level (see Table 17) and Wadia *et al.* (2007).
  - AREG is a ligand for epidermal growth factor receptor, the unique EGF family member to be transcriptionally induced by estrogen in the mammary glands at a time of exponential expansion of the ductal system.
- Other peripubertal change is associated with an increased number of estrogen-receptor ERa in the exposed animals (Murray *et al.*, 2007).

Among the four *in vivo* studies that have assessed the Progesterone Receptor (PR) expression, three of them reported an increase in the level of PR expression, an estrogeno-dependent gene, in rodents exposed to BPA when they were foetuses or juveniles (Munoz de Toro *et al.*, 2005; Jenkins *et al.*, 2009; Ayyanan *et al.*, 2011). Although Vandenberg *et al.* (2008) did not report an increased expression of PR at 9 months of age in the epithelium, they found increased expression of PR in epithelial cells in intraductal hyperplasia. Lastly, it should be noted that increased PR mRNA expression induced by BPA was similar to the positive control (DES) included in the Ayyanan *et al.* study.

The observed increased expression of PR in mice and rats may explain the increased ductal density and the increased lateral branching as PR is essential for side branching in the mammary epithelium (see Table 18 and Background section in 4.3.2).

Furthermore, exposure to BPA during fetal life provokes at adult stage, an increase of the expression of both *RankL* (a critical connection between progesterone and epithelial cell proliferation) and *Wnt4* (involved in progesterone-induced side branching in early adult life), see respectively Jenkins *et al.* (2009) and Ayyanan *et al.* (2011) and Table 16.

#### 4.3.4.3 Molecular changes involved in the process of carcinogenesis

A series of studies have described a link between molecular and abnormal mammary gland changes in rodents exposed to BPA. Those studies addressed the effects of BPA on EZH2 and HOTAIR involved in the increase of cell proliferation, or increased invasiveness, seen in some breast tumors and which also contribute to breast cancer progression. HOTAIR and EZH2 were recently shown as being estrogeno-regulated genes (Bhan *et al.*, 2013 and Bhan *et al.*, 2014a, respectively).

Doherty *et al.* (2010) showed that *in utero* and lactational exposure of mice to BPA at 5 mg/kg bw/day, or with DES at 10  $\mu$ g/kg bw/day, up-regulates the expression of EZH2 through an epigenetic mechanism. This EZH2 up-regulation by BPA has also been shown *in vitro* in breast cancer cells (Doherty *et al.*, 2010; Weng *et al.*, 2010; Knower *et al.*, 2014 (for review); Bhan *et al.*, 2014a).

Some studies from the group of Mandal (Bhan *et al.*, 2014a – 2014b; Hussain *et al.*, 2015 and Deb *et al.*, 2016) have described the induction of epigenetic marks in the BPA-exposed adult mammary gland.

Bhan *et al.* (2014a) confirmed the previous observation of Doherty *et al.* (2010) showing the induction of EZH2 with BPA. This induction of EZH2 (mRNA and protein) was observed in BPA-treated MCF7 cells as well as *in vivo* in the mammary glands from ovariectomised rats exposed to BPA with 25  $\mu$ g/kg bw for 24h, estradiol (5  $\mu$ g/kg bw) or DES (DES 5  $\mu$ g/kg bw). Induction of HOTAIR expression was also described in both *in vivo* and *in vitro* models. HOTAIR is a key player in survival and maintenance of breast cancer cells (Bhan *et al.*, 2014b) and is transcriptionally regulated by estradiol (see above and in background section). Moreover, Bhan *et al.* (2013) showed that knockdown of ERs down-regulated the BPA- and DES-induced HOTAIR expression.

Studies from this group (Bhan *et al.*, 2013) analysed molecular mechanisms involved in the regulation of EZH2 and HOTAIR by BPA in breast-cultured cells (MCF-7). EZH2 and HOTAIR mi-RNA interacts with gene silencing machinery, through recruitment of PRC2 and LSD1 complexes into the target promoter, which leads to target gene silencing. The complexes bind to the promoters of EZH2 and HOTAIR, and modify chromatin (histone methylation and acetylation).

Altogether, these observations suggest that BPA exposure (at least in the conditions cited above) alters the epigenetic programming of the promoter of these genes. These genes have been shown to be EE2 dependent, their epigenetic modulation by BPA may contribute to their endocrine disruption *in vitro and in vivo* in an EE2-like manner.

An initial study from Markey *et al.* (2005) suggested that the mechanisms by which BPA affects the morphology and secretory function of the mouse mammary gland a long time after the period of exposure, could be mediated through misexpression of HOX genes. Using the same protocol as described above, the group of Mandal showed that the HOXB9 and HOXC6 expressions are transcriptionally regulated by estradiol (E2) and also by BPA in cultured human breast cancer cells (MCF7) as well as *in vivo* in the mammary glands of ovariectomised (OVX) rats (Deb *et al.*, 2016 and Hussain *et al.*, 2015, respectively).

Dhimolea *et al.* (2014) have shown that fetal exposure to BPA at 250 µg/kg bw/day (*via* subcutaneous administration) triggers changes in the post-natal mammary gland epigenome (rats). In particular, methylation of lactalbumin gene (promoter)<sup>15</sup> was increased (*via* proactivation histone H3K4 trimethylation) at PND4 concomitantly with enhancing mRNA expression of the gene. The majority of differentially methylated genomic DNA segments between BPA- and vehicle control animals were observed at PND21. However, no link between methylation status and gene expression was observed at PND21 and PND50 at these two periods.

Altered methylation of genes involved in DNA repair (BRCA1), cell cycle regulation and apoptosis (CCNA1 and CDNK2A/p16), has been observed with low concentration of BPA (100 nM). This altered gene methylation was observed in cells such as human breast normal-like cells, human normal mammary epithelial cells (HMEC) and MCF-10F cells (Fernandez et al., 2010b and 2012; Qin et al., 2012), or in other ER positive human breast epithelial cell lines. Rounded ductal colonies (similar to spheres) treated with BPA showed increased methylation levels of key genes associated with tumor development such as BRCA1, CCNA1, CDKN2A, indicating that BPA alters the epigenome in a manner that promotes proliferation, senescence and tumour development at  $10^{-8}$  M  $- 10^{-7}$  M (Qin et al., 2012). Additional cell line studies have demonstrated aberrant methylation of the genome resulting in downregulated pro-apoptotic genes (Fernandez et al., 2012).

Hypermethylation of promoter CpG islands is often associated with tumor suppressor genes (BRCA1, E-cad, CDNK2A/p16) and is recognised as one of the key events leading to tumor

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<sup>&</sup>lt;sup>15</sup> Hormone-dependent marker of differentiation.

initiation and progression (Faryna *et al.*, 2012; Knower *et al.*, 2014). Silencing of CDKN2A/p16 could allow BPA-exposed epithelial cells from the mammary gland to escape senescence.

# 4.3.5 Summary of the plausible link between adverse effects and endocrine MoA: Effects of BPA on mammary gland

Because of temporality issues, it is rather difficult, if not impossible to demonstrate all the mechanisms or MoA together with the delayed effects they are leading to (increased susceptibility to carcinogens or modification of morphology) in a standalone study. What one can achieve at best is to demonstrate key events known to be involved in specific MoA which lead ultimately to adverse effect, with each point when animals need to be culled being demonstrated in independent studies.

There is substantial evidence from experimental studies indicating that BPA fetal exposure alters the tissue organisation of fetal mammary gland in rodents and non-human primates, causing effects such as increased ductal area, and changes in the periductal stroma and maturation of the fat pad in exposed fetuses. The experimental data have also shown that early-life BPA exposure may lead to increased susceptibility to mammary cancer.

Some studies using transcriptional analyses on the stromal and epithelial compartments isolated from the fetal mammary gland (mouse) demonstrate that BPA exposure in dams alters the mesenchymal and epithelial transcriptomes (Wadia *et al.*, 2013). Changes in gene expression in the BPA fetal exposed mammary gland were related to proteins involved in apoptosis (increased expression of the anti-apoptotic gene, *Birc2*, *Abl1*), myoepithelial differentiation, changes in the composition of ECM and in adipogenesis.

BPA MoA involves estrogen receptor and/or ER co-regulators. In human breast cells, activation of ER and GPER were shown after BPA exposure leading to disruption of acini formation. Involvement of ER dependent mechanisms was highlighted using positive controls, 17βestradiol or DES. When BPA or 17β-estradiol were combined with ER and GPER inhibitors (ICI 182 780 and G15, respectively), these effects were reversed (Marchese and Silva, 2012). Alterations of the ductular pattern, described after BPA exposure, were similar to those formed by the cells treated with positive control (E2) as shown by Fernandez and Russo (2010). BPA induced increased cellular proliferation in human mammary epithelial cells (Qin et al., 2012; Sengupta et al., 2013) similar to the positive control: 17β-estradiol in Lee et al. (2012). Additionally, activation of proliferation and increased level of estrogen-responsive gene ps2 by BPA showed the engagement of ERg and the co-activator SRC3 to a similar level as 178estradiol (Sengupta et al., 2013). Lastly, the BPA-induced increase in the expression of HOTAIR, a procarcinogenic gene, was suppressed after ER invalidation and the same observation was made for DES. Besides ERs, BPA may act via co-factors. GPER is known to be required for growth effects and migration in cancer (SKBR3) cells and CAFs that lack the classical ER. As these proliferative effects were cancelled when GPER expression was silenced by shGPER, it can be concluded that BPA induces stimulatory effects as a GPER agonist in these breast cancer cells and CAFs (Pupo et al., 2012).

It can be noted that a series of studies described how BPA alters the epigenetic programming of the promoter of HOTAIR and EZH2. These genes have been shown to be EE2 dependent and have been shown to be modulated by BPA. They are involved in the increase of cell proliferation, or increased invasiveness, seen in some breast tumors and also contributing to breast cancer progression.

It is shown that rodents and non-human primate mammary gland and human breast cells are targeted by BPA exposure and that mechanisms could involve different key events as indicated above. The schemes presented below aim to link those different key events in a dynamic way up to the adverse effect.

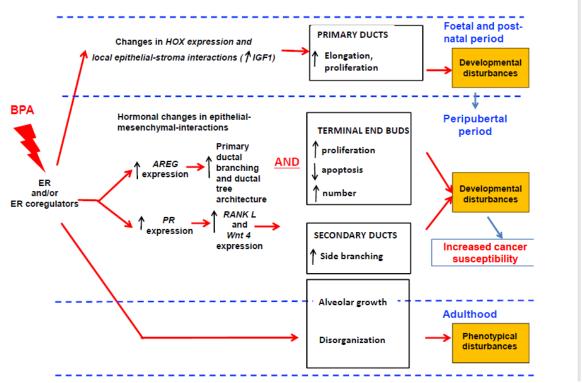
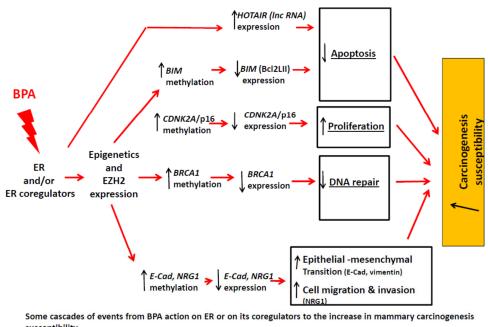


Figure 13: Some cascades of events from BPA action on ER or on its coregulators to the developmental and phenotypical disturbances of the mammary gland.



susceptibility

Legend: Lnc RNA stands for long non coding RNA. When gene silencing was involved in the MoA, the corresponding effect was

Figure 14: Some cascades of events from BPA action on ER or on its coregulators to the increase in mammary carcinogenesis susceptibility.

### 4.3.6 Human information

Very few epidemiological studies are available.

Aschengrau *et al.* (1998), in a case-control study found no association between occupational exposure to BPA and breast cancer. Another study determined blood BPA level in women with and without breast cancer (Yang *et al.*, 2009). There were no significant differences in blood BPA level between the cases and controls. In those studies, a link between BPA exposure and human breast cancer could not be determined.

More recently, Sprague *et al.* (2013) conducted a cross-sectional study in 264 postmenopausal women for whom a mammography was performed. This mammography was used to measure breast density (an important breast cancer risk factor) and was reported as a ratio of breast density (percentage of high density areas to the total breast area). BPA was measured on a single spot blood sample at the time of mammography. The authors observed that compared with women with non-detectable BPA values (n = 193), those who had BPA values, below (n = 35) or above (n = 34) the median of detectable values had a higher mammary density ratio (P trend = 0.01). However, this cross-sectional study coupled with a single measure of exposure, with a very low detection rate of the molecule and focusing on a risk factor (breast density) and not to the occurrence of the disease (breast cancer), does not provide evidence of a causal link.

Trabert *et al.* (2014) conducted a population-based case-control study in Poland. 575 incident cases of post-menopausal breast cancer (which represents the majority of cases in the Caucasian population) were compared to 575 women without breast cancer and matched on age (+/- 5 years) and region of residence (Warsaw, Lodz). All control women were also menopausal. There are some aspects that deserve attention such as: large population size, data analysis that takes into account many confounding factors and fairly compelling sensitivity analyses. Exposure to BPA was estimated on the basis of a urine assay on samples collected for 12 hours at night. This is a more reliable measure than a simple spot, but unfortunately it cannot account for the exposure in any previous critical window or for a long period before the occurrence of a disease. Again, measurement of exposure is the major limitation. Considering this limitation, the authors found no significant association between BPA exposure and the risk of developing breast cancer.

A more biologically relevant study design would be longitudinal epidemiological studies measuring BPA *in utero*, as breast cancer most likely takes years to develop and even be established in the womb (Soto *et al.*, 2008). Indeed, it should be noted that the existing studies evaluate BPA exposure on the date of the study when the effects looked at are long-term effects appearing more than 40 years after exposure.

## 4.3.7 Transposition to humans: interpretation issues

4.3.7.1 Comparison of the pre- and postnatal development of the human breast to the mouse mammary gland

The anatomy of the human breast is more complex than the mouse mammary gland which contains 15–25 ducts where each gives rise to a lobe containing multiple terminal ductal lobular units and 2 distinct stromal compartments, the intralobular and interlobular stroma, which has a single stem ductal tree embedded in a homogeneous fatty stroma. Nevertheless, terminal ductal lobular units (TDLU) present in humans are structurally similar to TEBs in rats

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<sup>&</sup>lt;sup>16</sup> \* below the analytical detection limit

and mice during the same life stage. These structures are undifferentiated and highly proliferative, and as such they are sensitive to the effects of carcinogens and other chemicals. In terms of hormonal regulation, there seem to be substantial similarities across species. In most mammals, the ovaries first secrete estrogens in response to increased secretion of gonadotropins, and sexual maturity coincides with the establishment of cyclic peaks of ovarian progesterone secretion. Progesterone levels increase after ovulation when the body anticipates pregnancy, and continue to rise when pregnancy is established.

Pathologists observe proliferative activity in the breast epithelium during the luteal phase, when progesterone levels peak (Masters *et al.*, 1977; Longacre and Bartow, 1986), suggesting that mouse and human mammary epithelia may indeed be similarly regulated, at least with regards to hormonal control of cell proliferation. Recently developed *ex vivo* models of the human breast have shown that progesterone elicits cell proliferation (Tanos *et al.*, 2013; Wang *et al.*, 2013). Notably, the dog, a species with a particularly long luteal phase, is especially prone to mammary carcinoma (Sleeckx *et al.*, 2011).

The flowchart hereabove in section 4.3.2.1 (see Figure 10) summarises the different developmental steps in species such as mice, rat and also human (quoted from Davis and Fenton, 2013).

## 4.3.7.2 Comparison of precancerous/ cancerous phenomenon between species

Precancerous lesions of the breast are atypical epithelial proliferations which develop within the lactiferous duct tree and are of two types: ductal and lobular. These two types differ not only in their location but also in the type of their constituent cells. Histological diagnosis of precancerous lesions is difficult and inter-pathologist reproducibility is poor as shown by a number of studies. The classification of precancerous lesions in humans is divided into the terms ductal (DIN) or lobular (LIN) intraepithelial neoplasia. Ductal carcinoma *in situ* (DCIS) is a preinvasive cancerous lesion. In the United States, DCIS accounts for almost 20% of the cancers picked up in screening (1 case of DCIS per 1300 screening mammographies) (Ernster *et al.*, 2002).

When left in place, a preneoplastic or precancerous lesion can turn into a preinvasive carcinoma or an *in situ* carcinoma which can itself turn into an invasive carcinoma. The theory about the existence of a continuum between the normal mammary gland and invasive breast cancer, even if it may appear too simplistic, is based on direct and indirect arguments (Antoine *et al.*, 2010). Recent epidemiological studies have shown that women with a history of benign breast lesions had an increased risk of breast cancer.

Similarly, after 10 years' follow-up in women who had undergone a diagnostic biopsy of low-grade DCIS without any other treatment than biopsy excision, 32% of these women had a diagnosis of invasive cancer in the same breast (Page et al., 1995). The natural development of high-grade DCIS or of clinically palpable DCIS, on the other hand, is not well characterised since, in most cases, the tumour is removed in its entirety by surgery which is also the case with atypical ductal hyperplastic (ADH) lesions.

The substantial increase in the number of biopsies performed on the basis of infra-clinical images and recent data provided by molecular study of the lesions have shed new light on the risk of hyperplastic lesions becoming cancerous. Molecular markers of tumoral transformation in the breast such as the estrogen receptor, expressed by normal epithelial breast cells, are expressed by more than 70% of DCIS and the proto-oncogene HER2/neu is overexpressed in half the cases of DCIS but not in atypical hyperplasias (Allred *et al.*, 1992).

Rodents, i.e. rats and mice, have been widely used to study mammary carcinogenesis, in models of either spontaneous or induced tumours. The main advantage of the rat model is that the carcinoma most resembles human breast cancer; breast cancer in mice is often of viral and hormone-dependent origin (Cardiff *et al.*, 2000; Gould, 1995). In CD-1 mice, spontaneous

non-neoplastic and neoplastic lesions are not very common (less than 5%: (Gad, 2007)).

The different strains of rats used have shown different sensitivities in developing neoplasms induced by chemicals or by radiation, Sprague-Dawley or Wistar being more susceptible than the Fisher rat. In Sprague-Dawley rats, the incidence of spontaneous tumours is close to 50% in chronic studies (example, historical data (NTP, 2010)). Certain strains, such as Wistar-Furth, show increased susceptibility to mammary carcinogenesis *via* chemical carcinogens (Gould, 1995).

The factors of mammary gland susceptibility include, in addition to genetic factors, the degree of differentiation of the breast tissue at the time of exposure, physiological and hormonal status, and diet. Susceptibility is increased in prepubertal females during the mammary development period: the ducts end in TEBs which will progressively differentiate into alveolar buds (AB) and alveolar lobules. The greatest number of tumours was induced in female SD rats at between 40 and 46 days, the period of most active differentiation of the TEBs regarded as the target of chemical carcinogens (Russo and Russo, 1996). Breast carcinomas were induced in rats by chemical agents or ionising radiation. The most commonly used chemical carcinogens include the polycyclic aromatic hydrocarbon dimethyl-benzanthracene (DMBA) or the alkylating agents N-ethyl-N-nitrosourea (ENU) and N-methyl-N-nitrosourea (NMU). Carcinogen-induced tumors in rodents are characterised using the same criteria as used for human breast DCIS. Studies in human and rodent models demonstrate that hormonal factors that affect mammary gland development also influence susceptibility to carcinogens.

These similarities include the development in a multistage process. Most of the cancers induced by DMBA (or NMU) are hormone-dependent with a similar morphological pattern i.e. hyperplasia, intraductal hyperplasia regarded as preneoplastic, adenomas/ adenocarcinomas. DCIS are regarded as a morphological progression towards breast carcinoma from intraductal proliferative lesions.

## 4.3.8 Summary and conclusion

There is evidence from rodents and non-human primate studies that prenatal and post-natal exposure to BPA causes endocrine modifications in the mammary tissue, ultimately increasing its susceptibility to chemical carcinogens, as previously reported (ANSES, 2013b and review by Soto *et al.*, 2013). All data presented here support the possibility that BPA, through interaction with the nuclear ERs, or GPER (which may also play a role, see Filardo *et al.*, 2006), and indirectly with PR, modulates estrogenic- and progestin agonist activities. Emerging epigenetic studies have reported changes related to estrogen-dependent genes (such as EZH2 and HOTAIR), as well as HOX genes (involved in embryogenesis and post-natal development) associated with the BPA induced abnormal development and cancer increased susceptibility of mammary gland.

### 4.3.9 Summary tables of studies

The *in vivo* and *in vitro* experimental data are reported in the following tables from Table 14 to Table 18.

Table 14: Summary table of *in vivo* experimental studies investigating the effects of BPA on mammary gland following foetal exposure with a mammary gland evaluation around birth

Literature reference	Species	Route of exposure	Pre-natal exposure	Endpoint : fetal mammary gland development	BPA and ER expression	BPA effects
Vandenberg et al., 2007	CD1 Mice	Alzet osmotic pump	-	- Advanced maturation of the fat pad in the stroma and changes in the extracellular matrix, - altered growth, decrease in cell size and delayed lumen formation in the epithelium.	-No statistically significant differences in ERa or ERβ mRNA expression between BPA-exposed animals versus controls or by immunohistochemistry quantification.  ERa et Erβ transcripts present at GD 18.  ERa-positive cells present in the loose connective tissue and fat pad compartments of the developing stroma and blood vessels.	
Wadia <i>et al.</i> , 2013	C57BI6 (ER+/-) Mice	Alzet osmotic pump	GD8 to GD19 - BPA: 250 ng/kg bw/day (in DMSO) EE2: 10 ng/kg bw/day Epithelial and stromal dissection in female fetuses at GD 19.		Estrogen receptors: ERa (only stroma), and GPER (stroma periductal), ERRY (not present).  Other nuclear receptors: GR, TRa (epithelium and stroma), AR (stroma).	Transcriptome analysis results: BPA acts on ductal epithelium and periductal stroma  1-BPA directly affects the periductal stoma which correlates with morphological changes: altered expression of genes involved in the focal adhesion pathway (down- regulation of Tenascin (Tnc)), and in adipogenesis pathways (upregulation of PPARY, a master gene of adipogenesis, as well as other adipogenic genes such as low density lipoprotein receptor (Ldlr), G protein- coupled receptor 81(GPR81), and Fabp4)).  2-Significant similarities in the transcriptional changes induced by BPA and EE2 in the exposed epithelium compartment: - changed expression of genes regulating the apoptosis pathway,

						with up-regulation of anti-apoptotic genes baculoviral IAP repeat-containing protein 2 (Birc2) and v-abl Abelson murine leukemia viral oncogene homolog 1 (Abl1).  - up-regulation of the hepatocyte growth factor receptor/Met proto-oncogene (Met), implicated in branching of the mammary gland.
Tharp <i>et al.</i> , 2012	Rhesus macaques (M. mulatta)	Oral	From GD 100 to GD 165 400 µg/kg bw/d.  Period corresponding roughly to the third trimester of human gestation.  Unconjugated seric BPA concentration of 0.68 ±- 0.31 ng/ml	Significantly increased density of mammary buds (increased number of buds per ductal area).  More advanced overall mammary gland development: -increased number of buds (i.e. incipient branches), terminal ends, branching points, bifurcating ends, total mammary gland area, ductal area, and number of ductal units (namely, the number of lactiferous ducts that define the number of lobes), as compared with controlsprenatal differentiation of myoepithelial cell layer (compared to mice, postnatally).	Both ER α and ER β are expressed in epithelial cells at birth. No differences in the expression of ERs are observed between BPA-treated animals and controls.	

Table 15: Summary table of mice in vivo experimental studies investigating the effects of BPA on mammary gland following a pre and/or postnatal exposure with a post-natal or adult mammary gland evaluation

Literature reference	Species	Route of exposure	Perinatal exposure Gestation or gestation and lactation	Endpoints : Developmental alterations (peripubertal and at adult age)	BPA and hormonal expression	BPA effects
	Mice			Epithelial density alteration Canalaire hyperplasia, stroma changes		
Markey et al., 2001	CD-1 Mice	Alzet osmotic pump	GD9 to GD20 BPA: 25 and 250 µg/kg bw/day (in DMSO) Observation: PND 10, PND 30 (puberty) and PND 180 (mice killed on the afternoon of proestrus).	Dramatic expansion of the ductal network at 1 month (25 µg/kg bw/day).  Most apparent changes in the histoarchitecture of the mammary gland at 6 month of age (25 and 250 µg/kg bw/day): - significant increase in all epithelial (ductal and alveolar) structures relative to the control group, including a significant increase in the relative area of AB (300%), of terminal ducts (TD, resting structures) and terminal end buds (TEB); - increased DNA synthesis in the stroma at 6 months of age; - increased presence of secretory product in alveoli resembling early gestation period.	ND	These histoarchitecture changes coupled with an increased presence of secretory product within alveoli, resemble those of early pregnancy, and may suggest a disruption of the hypothalamic-pituitary-ovarian axis and/or misexpression of developmental genes (such as homeobox genes: HOXA9, HOXB9, and HOXD9 see Chen et Capecchi, 1999).
Munoz-de- Toro et al., 2005 (A. Soto's team)	CD-1 Mice	Alzet osmotic pump	GD9 to PND4 25 and 250 ng/kg bw/day (in DMSO) Observation at PND20, PND30 and PND120 (mice at PND120 were killed on the afternoon of proestrus at PND120).	-Increased number and area of TEBs relative to the ductal area at PND30 in the BPA-exposed animals indicating that ductal growth might be impairedIncreased epithelial densityDecreased apoptotic activity from 25 ng/kg bw/day,  -Increased number of lateral branches at PND 120 in BPA-exposed mammary glands (25 ng/kg bw/day).	Expression of ERa in both the epithelial and stromal compartments of mammary glands at PND30. No effect of BPA on the expression of ERa (either in stroma or in epithelial ducts) at PND 30.  Expression of PR only observed in the epithelium at PND30. Increased expression of PR in the epithelium with	The morphological changes found in 30-d-old animals exposed perinatally to BPA could be attributed at least in part to an increased sensitivity to estrogens. Of note, BPA enhances mammary glands sensitivity to estradiol in ovariectomised CD-1 mice.  Progesterone is the main mediator of lateral branching and alveolar growth Increased

					BPA from 25 ng/kg bw/day. Clusters of PR positive cells are believed to be indications of future branching points (see Seagroves <i>et al.</i> , 1998).	expression of Wnt4 (mediator of lateral branching, downstream mediator of progesterone action) at PND30 in BPA-exposed animals at 250 ng/kg bw versus control.
Vandenberg et al., 2008 (A. Soto's team)	Mice CD-1	Alzet osmotic pump	GD8 to PND16 BPA: 0.25-2.5- 25 µg/kg bw/day (in DMSO) Observation at 3, 9, 12, 15 months.	- Significant increase of the volume fraction of AB, but not terminal ducts, using Whole Mount analysis, both at 3 months of age in the 0.25 µg/kg bw/day BPA group, and at 6 months of age in the 0.25 and 2.5 µg/kg bw/day BPA group.	No effect of BPA on the expression of ERa, PR or Ki67 (marker of proliferating cells) at 9 months of age.	Intraductal hyperplasia/ increased proliferative index (cf also Murray et al., 2007 in rat and mice Bern, 1983) which suggest an increased estrogen-sensitivity.
				- At 9 and 12-15 months of age, presence of ducts with a beaded appearance classified as intraductal hyperplasia (see also Murray et al., 2007). Of note, however, a trend to the lack of beaded ducts in the 12–15-month-old in the 25 μg/kg bw/day BPA group (due to either a too small sample size or to regression of beaded ducts in this group during later life).	Increased expression of PR and proliferation index (Ki67) in epithelial cells in intraductal hyperplasias.  Because PR expression in mammary epithelium is thought to be dependent on estrogen exposure, the expression of PR in these beaded ducts suggests that they are estrogen-sensitive.	
Ayyanan et. al., 2011	Mice C57Bl6	Oral route (Drinking water)	in utero until PND24 0.6-1200 µg /kg/j (n=18-animals per dose levels) Observation: PND30, 3 and 6 months  DES (0.12 or 1.2 µg/kg-bw/d) another estrogen receptor (ERa) agonist as a positive control.	<ul> <li>At PND30: significant increase in adjusted number of TEBs at a dose of 3 μg/kg-bw (non-monotonic dose-response)</li> <li>At 3 months of age: increased cell number with BPA and DES. Increased proliferation/ apoptosis ratio (at 6 μg/kg/d)</li> <li>Up to more than 1 year, no palpable mammary carcinoma is detected. Of note C57Bl/6 mouse strain has no predisposition to mammary</li> </ul>	- At PND30,  *ERa mRNA expression is not affected by BPA nor by DES treatment.  *Increased PR mRNA expression similar with DES and BPA treatment.  Decreased SLP1 (secretory leuko-protease inhibitor) similar with DES and BPA treatment at PND30.  - At 6 and 12 month of age, pronounced increase of PR-positive cells within the	At PND30: non-monotonic dose response trend in AREG mRNA expression with BPA.  At peri-puberty, BPA increases mRNA expression of well-characterised estrogen – regulated genes ( <i>PR</i> , <i>AREG</i> ) which are linked with the increased TEB number and proliferation.  In adult females, BPA increases Wnt-4 and RANKL mRNA levels

				luminal epithelial cells in the 6 μg/kg bw/day BPA group.	RANKL are important downstream mediators of progesterone function, both of which are implicated in the control of stem cell proliferation; their downstream signaling pathways are deregulated in mammary carcinogenesis  Epigenetic changes as a mechanism underlying the effects observed in adult females is suggested
Fischer et al., 2016	Mice CD-1	Osmotic infusion mini-pump	GD9-GD21 Dose: 5 mg/kg bw/day (sesame oil) Female pups ovariectomised six weeks after birth, then treated at 8 weeks with estradiol (E2 300 ng) or with vehicle. Observation: 6 hours after treatment	-Decreased expression of ERa (:2.5) -Increased expression of ERβ (x3).	Gene expression analysis: BPA exposure caused high gene expression of ER β, likely leading to subsequent decrease in the expression of several chemokines (CXCL12, CXCL4, CXCL14, CCL20), some interleukins (IL1β ), interferons (Irg1 and Irf9) and of leukocyte marker (inflammation and immunity mediators).

Table 16: Summary table of rat in vivo experimental studies investigating the effects of BPA on mammary gland following a pre and/or postnatal exposure with a post-natal or adult mammary gland evaluation

Literature reference	Species	Route of exposure	Gestation / gestation and lactation exposure	Endpoints : Evaluation of mammary gland developmental alterations at peripubertal and at adult age	BPA and ER (and PR) expression	BPA effects
Murray et al., 2007 (A. Soto's team)	Rats Wistar-Furtl Rat	Alzet osmotic pump	GD9-PND1 (in utero) 2.5-25-250 and 1000 µg/kg bw/day in DMSO (n=4) Observation: PND50 (peripubertal period) and at PND95.	At PND50 and PND95: -increased (intra)ductal hyperplasia (3-4 fold ?) and increased epithelial and stromal proliferation (see Durando et al., 2007)intraepithelial neoplasia from 250 and 1000 µg/kg bw/day, described as cribiform like structures and classified as CIS 'carcinoma in situ'	Increased ER a and Ki67 expression at PND50 and PND95 in hyperplasic lesions versus normal ductal structures.	Increased estrogenic activity.
Durando et al., 2007 (A. Soto's team)	Wistar rat	Miniature osmotic pump	GD 8 to GD23 (in utero) BPA: 25 µg/kg bw/day in DMSO with or without administration of Nnitroso Nnmethylurea (NMU) at a subcarcinogenic dose of 25 mg/kg at PND50 with observation at PND110 and PND180.  Observation of female offspring at PND30 (prepuberty)-PND50 (puberty) and PND110-PND180 (adulthood).	-At PND50: increased proliferative/apoptotic ratio (stroma and parenchyma) -At PND110: increased ductal hyperplasia -At PND 50, 110, 180: stromal desmoplastic reaction (including mastocytes, fibroblasts, inflammation) - At PND110 and PND180: increased sensitivity to NMU (ductal hyperplasia); -At PND180: ductal carcinoma with BPA and NMU versus negative control.	Note: ER a and β are only expressed in the stroma and the epithelium at the end of gestation.  Note: NMU is a direct acting carcinogen which generates tumors that closely mimic the human disease in terms of tumor histology and ovarian hormone dependence (Russo 1996, Thompson 2000, Murray and Rosso 2009)	- In peripubertal period: proliferative/apoptotic ratio modified increased sensitivity to E2 (cf Munoz de Toro et al., 2005) - At adulthood, ductal hyperplasia (pre-neoplasic lesion) associated with increased stromal density (modification of the interactions between the epithelium- stroma, mastocytes).
Delclos et al., 2014	Sprague- Dawley rat	Oral (gavage)	2.5, 8, 25, 80, 260, 840, 2700, 100	At <b>PND 21</b> : significant elevated incidences of mammary gland duct	Not determined	Significantly higher plasma levels of estradiol and prolactin in the female

			000, 300 000 μg BPA/kg bw/day Negative controls: naive and vehicle Positive control: EE2 0.5 and 5 μg/kg bw/day F0: females exposed from GD6 up to labour onset Pups from PND 1 until tissue harvesting, up to PND 90 GLP study. (Mod. OECD TG 408)	hyperplasia of minimal severity in the female groups at 2 700 and 100 000 µg/kg bw/day, but not at 300 000 µg/kg bw/day.  At PND 90: minimal severity of mammary gland duct hyperplasia also reported in the high dose female BPA groups, increase was statistically significant at 300 000 µg/kg bw/day group (Poly-k test) and 2700, 100 000 and 300 000 µg/kg bw/day ( JT/SW or RTE statistical tests).  BPA did not cause duct hyperplasia in the mammary glands of male rats, while conversely the reference estrogen EE2 induced hyperplasia in the male but not the female mammary gland.  One mammary adenocarcinoma observed at the lowest tested dose level only (2.5 µg/kg bw/day)		BPA groups at 100 000 and 300 000 µg/kg bw/day whereas the EE2 values (positive controls) were only mildly elevated in comparison to negative controls.
Acevedo et al., 2013 (A. Soto's team)	Sprague- Dawley rat	Alzet osmotic pump	GD 9 to GD21 and GD9 to PND21; BPA: 0.25-2.5-25 and 250 µg /kg bw/day (without co-carcinogen administration); Observation at: PND50-PND90-PND140 and PND180 BPA quantification (detectable at 250µg/kg bw/day	Neoplasic lesions (including DCIS) observed postnatally and at adulthood (see Murray et al., 2007).	ND	How BPA contributes to the initiation and progression of neoplasia, mechanisms are discussed (change in the hypothalamic-pituitary-ovarian axis, increased sensitivity to ovarian hormone: BPA acting as an estrogen).
Moral et al., 2008 (Russo's team)	Sprague- Dawley rat	Oral Gavage	GD9-PND1 (in utero) BPA: 25 and 250 µg/kg bw/day (in sesame oil) Observation at PND21-PND35- PND50 and PND100	At PND21: increased number of TEB (Terminal End Buds) and lobular structures (at 250 µg/kg bw/day) without major influence on proliferative index.  At PND100: increased number of undifferentiated structures by this age	ND	Very large transcriptomic analysis at PND21, PND35, PND50 and PND100: -Few changes at PND21 and PND35At PND50: increased number of up-modulated genes related to: apoptosis, immune function and response to stress (Cd3d, cathepsin

				as TDs (Tubular Ducts at 250 μg/kg bw/day).		E, Ctse, Cd5, and Sod2) at low dose and related to differentiation at high dose.  -At PND100, important cluster of upregulated genes related to immune response (Cd3d, Ctse, Cd5, Ltb, Cxcl10, Ccl5, Mefv, Cd2, A2m, and Il1b) at high dose.
Jenkins et al., 2009 (Russo- Lamartinière- Jenkins' team)	Sprague- Dawley rat	Oral Gavage	PND2 - PND20 BPA: 25 and 250 µg/kg bw/day (in sesame oil), administrated to lactating mothers 5 days/week (~ to 15 adm./mother). Observation at PND 50  DMBA administrated in F1 females at PND50 (dose: 30 mg/kg).	At PND50: Without DMBA: increased cell proliferation (Ki67) and decreased apoptosis in TEBs on PND50 at 250 µg/kg/ bw/day  With DMBA: dose-dependent increase in number of mammary tumors, with reduced tumour latency	No change for ER a or PR-B expression. Increased expression for PR-A (PR-A and coactivators). Note: PR-A is the predominant isoform at puberty.	At PND50: Increased expression of PR-A, coregulator SRC proteins (SRC-1, SRC-2, and SRC-3), up-regulation of Akt expression/activation plausibly linked to a greater incidence of side branching and alveologenesis, increased susceptibility to carcinogenesis.
Betancourt et al., 2010 (Russo- Lamartinière- Jenkins' team)	Sprague- Dawley rat	Oral Gavage	GD10-GD21 BPA: 25 and 250 µg/kg bw/day (sesame oil) from GD10-GD21 followed with DMBA administration in prenatally exposed females on PND50 (dose: 30 mg/kg bw) with observation at PND 50.  BPA: 250 µg/kg bw/day (sesame oil) from GD10- GD21 followed with DMBA	At PND50: no significant increased tumour incidence.  At PND100: increased tumour incidence indicating an increased susceptibility to carcinogenesis.  Increased cell proliferation (Ki67) at PND100 in epithelial cells but not in the stroma.  Note: In studies of cancer causation or chemo-prevention, the standard protocol for administering DMBA is at day 50, because this is within the period (days 40–60) of high mitotic index in the terminal ductal structures of rats (Russo et al., 1983; Welsch 1985).	- Down regulation of ERa at PND50 and up-regulation at PND100 Upregulation of co-regulator SRC proteins (SRC-1, SRC-2, and SRC-3) at PND100 and of SRC-3 at PND50.	Proteome analysis (Maldi-Tof et LC-MS/MS): At PND50: proteins and major pathways altered by prenatal BPA exposure: up-regulation of vimentin (epithelial to mesenchymal transition), down-regulation of SPARC and TGFβ (cell proliferation and differentiation, regulation with the extracellular matrix), up-regulation of 14-3-3 (signaling pathway and proliferation), members of the Raf and ERK families (key signal transduction proteins known to be involved in tumorigenesis)

			administration in prenatally exposed females on PND100 (dose: 30 mg/kg bw) with observation until ~ PND300			
Ibrahim et al., 2016	Albino rat	Oral Gavage	Adult female rat BPA: 5 mg/kg bw/day (in corn oil) for 8 weeks.	At adult age, increase in the number of the ducts and acini of the mammary gland with hyperplasia of their lining epithelium, increase in both amount and distribution of collagen fibers.	No significant effect on ERa expression.	NB. Adult exposure

Table 17: Summary table of in vitro experimental studies investigating the effects of BPA on mammary gland

Literature reference	Human cells	Cell characteristics	Condition of exposure	Evaluated endpoints :	Results:
Diel <i>et al.</i> , 2002	MCF-7	MCF-7: Human breast cancer cell line (immortalised cells, ERa positive)	BPA: 10 <sup>-8</sup> M-10 <sup>-7</sup> M - 10 <sup>-6</sup> M- 10 <sup>-5</sup> M, ICI 182 780 (anti- estrogen) RAL (Raloxifene) E2 Selective ER modulator (SERM).	Cell proliferation. Apoptosis Expression of: progesterone receptor (PR); AR; ERa protein. Gene expression such as estrogen dependent.	BPA: strongly inhibited apoptosis, slightly increased cell proliferation at 10 <sup>-5</sup> M, BPA did not effect the expression of PR mRNA, AR mRNA, ERa protein versus positive controls.
Qin et al., 2012	НМЕС	HMEC: Human normal mammary epithelial cells	BPA: 10 <sup>-8</sup> M - 10 <sup>-7</sup> M Exposure to BPA at passage 8 for 1 week. E2	Cell proliferation. Senescence.	BPA at 10-7 M increased significantly the proliferation, sphere size and senescence (increased number of human heterochromatin protein-1y positive cells, protein levels of both p16 and cyclin E (transition G1/S)) of HMEC, in a quite similar manner as E2.  BPA increased DNA methylation of genes related to development of most or all tumor types, such as BRCA1, CCNA1, CDKN2A.
Fernandez and Russo, 2010 (review)	MCF-10F	MCF-10F: spontaneously immortalised breast epithelial cell line, considered as "normal stage" (ERα negative, and ERβ positive cells).	BPA: 10 <sup>-6</sup> M-10 <sup>-5</sup> M -10 <sup>-4</sup> M-10 <sup>-3</sup> M BBP (Butyl benzyl phthalate) Treatment during two weeks in DMSO or media. E2: 0.007 nM - 70 nM -3.6 μM	Ductules formation in collagen (3D model). In-vitro neoplasic transformation.	BPA increased cell proliferation, but significantly decreases ductules formation in collagen vs control (3D cultures) at $10^{-6}\text{M} \cdot 10^{-5}\text{M}$ BPA increased the percentage of spheric solid masses vs control in a statistically significant manner at $10^{-6}\text{M}$ . BPA was toxic at $10^{-4}\text{M}$ and $10^{-3}\text{M}$ . No statistically significant increase in the invasive capacity with BPA treatment.
Fernandez et al., 2010 (Russo's team)	Transformed, invasive and fully malignant-derived MCF-10F cancer cell types	In vitro model of transformed and tumorigenic cells generated from MCF-10F: transformed by E2 (trMCF), invasive (bs MCF), and obtained from tumors generated on SCID mice (CaMCF),		Analysis of epigenetic and transcription changes	Analysis of epigenetic changes induced by estradiol during the neoplastic process. Data showed that the methylation pattern of different genes (such as neuroregulin NRG1)related toductulogenesis, branching pattern, and during the invasion and tumor stages are involved in 'early and late stages of breast cancer'. Other processes such as proliferation, apoptosis (BIM), and estrogen metabolism were also altered.

		1	Τ	T	T
		compared to MCF10F		Human breast tissue samples	Methylation of NRG1, and associated down-regulation of NRG1, (gene and protein), were observed in bsMCF10 as compared to original MCF10F).  NRG1 was also partially methylated in invasive breast carcinoma as compared to normal breast tissue
Fernandez et al., 2012 (Russo's team)	MCF-10F	MCF-10F (see Fernandez et al., 2010 and Fernandez & Russo, 2010)	BPA: 10 <sup>-6</sup> M-10 <sup>-5</sup> M during two weeks in DMSO or media.	Gene expression and DNA methylation analyses.	Increased expression of genes at 10 <sup>-6</sup> M involved in DNA repair, (such as BRCA1, BRCA2), and down-regulation at 10 <sup>-6</sup> M-10 <sup>-5</sup> M of several genes involved in apoptosis (such as BCL2L11, also known as BIM). Hypermethylation of several genes was shown (in particular BCL2L11 or BIM, IGF1R and Twist).  Results also indicate that breast tissue of women with BRCA1 or BRCA2 mutations could be more susceptible to the effects of BPA.
Marchese and Silva, 2012	MCF-12A MCF-10A	MCF-12A: non-transformed breast epithelial cells) (Era, ERβ and GPER competent); they form spheroid acini and hollow lumen in matrigel matrix.  MCF-10A: Immortalised non- malignant breast cells (GPER competent; ERa negative/ and ERβ very low); they do not form acini.	BPA 10 µM; E2 1 pM-1 nM ICI 182 780 (antiestrogen) G15: GPER inhibitor  Two weeks of treatment.	Acini development in Matrigel (in-vitro3D model) including evaluation of: -cell proliferation, -ECM -differentiation, -central lumen formation in MCF-12A.	BPA and 17β-estradiol treatment resulted in misshaped acini and filling of the acinar lumen in MCF-12A. When these chemicals were combined with ER and GPER inhibitors (ICI 182 780 and G15, respectively), these effects were reversed.  No effect of BPA on MCF-10A vs control.  Note: The correct functioning and organisation of 3D acini depends on a fine balance between cell proliferation and cell death (apoptosis). An overstimulation or perturbation of these processes will lead to aberrations in glandular structure, such as formation of large misshapen acini with filled lumen and no apico-basal polarity (see Haenssen et al., 2010; Debnath et al., 2003; Reginato et al., 2005; Yanochko et al., 2006

					and Dimri <i>et al.</i> , 2005)
Lee et al., 2012	MCF7	MCF-7: Human breast cancer cell line (ER positive)	BPA 10 <sup>-7</sup> M - 10 <sup>-6</sup> M E2 and DES (10nM) ERa agonist (propyl pyrazoletriol, PPT) ERβ antagonist (DPN, diarylpropio- nitrile) ER antagonist (ICI 182,780)	Cell proliferation. Gene expression Senescence	BPA induced proliferation of MCF-7 cells in a dose-dependent manner, similarly to E2. BPA induced cyclin D1 expression and reduced p21 expression associated with the G1/S transition, resulting in the proliferation of MCF-7 cells. These effects were inhibited by the ER antagonist suggesting that the effects of BPA on cancer cells occur through ER signaling and mainly through ERa based on the results with ERa and ERβ agonist).
Filardo et al., 2000	SkBr3	SKBR3: breast cancer cells (ERα and ERβ negative, but GPER/GPR30 positive). GPR30 is an orphan receptor unrelated to nuclear estrogen receptors	<u>E2</u> : 1nM	Proliferation. Mechanism of action : E2 via GPER	The group of Filardo described for the first time the presence of the completed unrelated transmembrane receptor GPR30 which can mediate estrogen responsiveness in ER negative breast cancer cells (SKBr3).  17β-estradiol activates the mitogen-activated protein kinases, Erk-1 and Erk-2, via the expression of GPR30 and not via known estrogen receptors.  17β-estradiol also induces the release of HB-EGF (heparan binding epidermal growth factor) and activation of the EGF receptor, suggesting a novel signaling pathway with potential significance for breast cancer (see also Filardo, 2002).
Pupo et al., 2012	SKBr3	Both <u>SKBR3</u> and <u>CAF</u> cells are ERα and ERβ negative, GPER/GPR30 positive	BPA from100 nM to 1μM (in DMSO)	Proliferation and migration of cancerous cells.	Note: Pandey et al. (2009) have reported that, in SKBr3, the activation of GPER/GPR30 signaling by E2 and tamoxifen (an ER antagonist but GPER agonist) induces a transcription factor network, which resembles that induced by serum in fibroblasts. The most strongly induced gene, CTGF, promotes proliferation and cell migration.  In both SKBR3 cells and CAFs after 5-day treatment at 1µM, BPA induced cell proliferation and migration through GPER. These proliferative effects being cancelled when GPER expression was silenced by shGPER. These findings indicate that BPA induces stimulatory effects as a GPER agonist in both ERnegative SKBR3 breast cancer cells and CAFs.

					The use of specific pharmacological inhibitors and gene-silencing procedures indicates that BPA induces the expression of the GPER target genes CTGF, c-FOS and EGR-1, and the corresponding proteins through the GPER/EGFR/ERK transduction pathway in SKBR3 breast cancer and CAFs cells. Thus, GPER is required for growth effects and migration stimulated by BPA in both cancer (SKBr3) cell and CAF.
Magruder et al., 2014	SKBr3	SKBR3 - breast cancer cells (ERα and ERβ negative, GPER-1+)  Murine 4 T1 cells	BPA: 10 nM 17a estradiol 10nM 17β estradiol 10nM ICI 182 780 : 1μM	Adhesion, Cell growth. Molecular mechanisms	GPER-1 stimulation of murine 4 T1 or human SKBR3 breast cancer cells with $17\beta$ -estradiol (E2 $\beta$ ) promotes the formation of focal adhesions and actin stress fibers, resulting in increased cellular adhesion and haptotaxis on fibronectin (FN), but not collagen. These actions are also induced by BPA (data not shown), ICI 182 780 (ER antagonist), but not the inactive stereoisomer, $17\alpha$ -estradiol (E2 $\alpha$ ).

Sengupta et al., 2013	MCF-7 and MCF-7:5	MCF-7 (ER positive) MCF-7: 5C (estrogen deprived) MDA: MB-231 cells transfected with wild- type ERa (MC2 cells) or mutant ERa (JM6 cells, D351G).	BPA: 10 <sup>-13</sup> to 10 <sup>-5</sup> M, or at 10 <sup>-6</sup> and 10 <sup>-5</sup> M for gene expression analysis E2: 10 <sup>-9</sup> M RAL: Raloxifen 4-OHT: 4-hydroxy tamoxifen; dose dependence	Cell proliferation. Apoptosis. Analysis of E2- dependent gene ( <i>TFF1</i> (or <i>PS2</i> ).	-Increased cell proliferation (max. at 10 <sup>-6</sup> M) in MCF-7 cells with BPA, -Induced apoptosis with BPA and E2 in MCF-7: 5C cellsUp-regulation of <i>TFF1</i> ( <i>PS2</i> ), an estrogen-regulated gene with BPA (at 1-10 μM).  4-OHT, an estrogen antagonist via ERafailed, as expected, to up-regulated PS2 expression.In vivo, transcriptional activation of PS2 by BPA in comparison to E2 and 4-OHT treatment: -Recruitment of ERα and SRC3 at the promoter region of PS2 gene which has a well-characterised functional estrogen-responsive element (ERE) in its promoter (Metivier <i>et al.</i> , 2002) with BPA at 10 <sup>-6</sup> M and 10 <sup>-5</sup> M in a concentration-dependent manner, equivalent to results obtained with E2 treatment4OHT did not recruit, as expected, SRC3 and was comparable to vehicle treatment.  Activation of TGFα gene in MDA -MB 231 cells stably transfected with wt ERα (MC2 cells) or mutant ERα (JM6 cells, D351G) with BPA, but at higher concentrations as compared to E2.  Molecular docking of BPA to the ligand-binding domain (LBD) has shown that the binding mode predicted for the agonist conformation of ERa is more likely.  BPA decreases the apoptosis via ERα. Note: The coactivator, SRC3 (AIB1) plays a key role in transcriptional activation of several estrogen-regulated genes, including PS2 gene (Shao <i>et al.</i> , 2004; Labhart <i>et al.</i> , 2005), similarly to ERα.
Yang et al., 2013	hESC	hECS: derived from human Chinese blastocysts.	BPA: 10 <sup>-6</sup> - 10 <sup>-7</sup> - 10 <sup>-8</sup> -10 <sup>-9</sup> M in DMSO E2: 10 <sup>-9</sup> M (positive control)	Three-dimensional (3D) culture system used for analysis of differentiation of	Low dose BPA and E2 could influence the mammosphere area of induced differentiated mammary epithelial cells (iDMECs) and upregulate the expression level of Oct4 and Nanog proteins,

			Treatment during the differentiation process.	progenitor cells into epithelium mammary cells. Expression of CK18, E-cadherin (usually considered as a marker of mammary epithelial cells), epithelial cell adhesion molecule (EpCAM). Expression of pluripotency molecular markers: Oct4, Sox2 and Nanog.	while only BPA could downregulate the expression of E-cadherin protein.
Song et al., 2015	MCF-7 SkBr3	MCF-7: (ERα positive, GPER+) SkBr3: (ERα and ERβ negative, GPER+)	BPA: from 10 <sup>-10</sup> to 10 <sup>-5</sup> M at 6 dose levels in DMSO versus control group.	Mechanisms of the BPA- increased proliferation in MCF-7 and SkBr3.	BPA upregulates the protein levels of cell nuclear antigen PCNA and Bcl2 in MCF-7 and SkBr3 cells.  Since neither ICI 182,780 (a specific inhibitor of ERα/β) nor G15 (the specific inhibitor of GPER) abolished the BPA-induced proliferation of MCF-7 and SkBr3 cells, it suggests that the stimulation effects of BPA on the proliferation of breast cancer cells was independent of Erα and GPER.  Silencing of ERRγ (estrogen related receptor gamma) by its specific siRNA significantly abolished the BPA-induced proliferation of breast cancer cells (MCF-7 and SkBr3), while si-ERRα had no effect. BPA up regulated the mRNA and protein levels of ERRγ and triggered its nuclear translocation via a time dependent manner.  These results indicate that BPA can trigger the proliferation of breast cancer cells via ERK1/2/ERRγ signals.  Note that the expression of ERRα and ERRγ are associated with an unfavorable and favorable prognosis of breast cancer, respectively (Ariazi et al., 2002). The nuclear immunoreactivity of ERRγ was detected in 79% of breast cancer patients and tended to correlate with the lymph node status (Ijichi et al., 2011) while exogenously transfected ERRγ increased MCF-7 cell proliferation (Ijichi et al.,

		2011). These results suggested that ERRY plays an important role as a modulator of estrogen signaling in breast cancer cells.
		Considering that over expression of ERRy can increase the risks of breast cancer (Dairkee <i>et al.</i> , 2008) and BPA has great binding affinity with ERRy (Okada <i>et al.</i> , 2008), therefore we hypothesised that BPA can stimulate the proliferation of breast cancer cells. In the present study, we revealed that nanomolar BPA can significantly increase the proliferation of both ER positive and negative breast cancer cells, and ERRy mediated this stimulation effect of BPA.

Table 18: Summary table of recent experimental studies (on homeogens and epigenetics) investigating the effects of BPA on mammary gland following pre and/or postnatal exposure

Literature reference	Species	Cell characteristics for the in vitro study	Period of exposure: Fetal, post-natal or adult	Endpoints	MoA: d'action: epigenetic mechanisms
Dhimolea et al., 2014 (A. Soto's team)	Rat Wistar Furth		GD9-PND1(fetal) BPA 250 μ/kg/day (subcutaneous - osmotic pump) Obs: PND 4, 21, first estrus after PND 50	Not tested cf previous studies -Post-natal mammary gland development: significant increase in proliferation/apoptosis ratio -Ductal carcinoma in situ	Fetal BPA exposure triggers changes in the post-natal and adult mammary gland epigenome and alters gene expression pattern  PND4: Higher levels of pro-activation histones H3K4 trimethylation of α-lactalbumin promotor in treated mammary gland compared to control, with concomitant increase (x2) in RNA expression of this gene.  PND21: majority of methylated gDNA segments (Nimblegen ChIP array)  PND50: majority of gene expression differences between BPA and vehicle-treated rats (transcriptional analysis)
Doherty <i>et al.</i> , 2010	Mice CD1 In vivo and in vitro	MCF-7: Human breast cancer cell line (immortalised cells, ERa and Progesterone (PR) positive)	GD9-GD26 (in utero) BPA 5 mg/kg (i.p.); DES 10µg/kg (i.p.) Female offspring with in utero exposure were euthanized 6 weeks after birth.	Female offspring mammary tissue analysed for mRNA analysis andprotein analysis.  Epigenetic mechanisms	BPA exposure <i>in utero</i> significantly alters <b>EZH2</b> expression (protein, >2 fold) and activity (histone H3, trimethyl K27) in adult mammary tissue when compared to control mice.  DES increased also EZH2 expression (gene and protein) which is elevated in mammary gland of mice exposed <i>in utero</i> .  EZH2 is a key epigenic regulator. Histone methylation by EZH2 is a known epigenic modifier in breast cancers.

		in MCF-Z  DES: 5 dose levels from $5.10^{-6}$ ; to $10^{-10}$ M  BPA: 5 dose levels from 2.5. $10^{-4}$ ; to 2.5. $10^{-8}$ M for 48 h.		BPA exposure of (MCF-7) breast cancer cells increases functional activity of EZH2, i.e., histone H3 trimethylation. Similar results with DES.  Discussion. Two important target genes of EZH2 are p57 (CDKN1C), a cyclin dependent kinase inhibitor, and E-cadherin which is important in cell-cell adhesion and migration. Repression of p57 and E-cadherin by EZH2 may lead to the increase of cell proliferation, or increased invasiveness, seen in some breast tumors.
Bhan et al., 2014a (J Steroid Biochem Mol Biol)	Rat Sprague Dawley	Same protocol in vivo and in vitro as Bhan, 2014b: Ovariectomised adult rats. BPA sc: 25 µg/kg, 24h. Positive control: E2 and DES 5 µg/kg  MCF-7 cells: BPA 100 nM Positive control: : E2 0.1 nM, and DES 10nM)	Epigenetic mechanisms	<ul> <li>-in vivo, BPA increases HOTAIR expression (mRNA, 4-fold increase) in the mammary gland. This effect is also observed with DES (x 4.3) and E2 (x3.3).</li> <li>HOTAIR is a long non-coding RNA (which is not translated into protein), and is a key player in gene silencing and breast cancer.</li> <li>-in vitro, BPA (100 nM) increases HOTAIR expression (x6) in MCF-7 cells</li> <li>-molecular analysis of the HOTAIR promoter in BPA-treated cells (same protocol as for EZH2): the HOTAIR promoter contains functional estrogen-response elements (ERE2 and ERE3); ER (ERα and ERβ are involved for BPA-induced HOTAIR expression). Along with ER, co-activators such as MLL (2 methyl-transferases H3K4, MLL1 and MLL2) and CBP/p300 (histone acetylase) bind to the HOTAIR promoter and modify chromatin in the presence of BPA. These effects are inhibited when BPA-treated cells are knocked-out for ERα or ERβ.</li> </ul>
Bhan et al., 2014b (J Mol Biol)	Rat Sprague Dawley In vitro	Same protocol in vivo and in vitro as Bhan 2014a Ovariectomised adult rats BPA sc: 25 µg/kg, 24h (Positive control: E2 et DES 5 µg/kg)  In vitro: BPA 100 nM Positive control: E2 and DES (10 nM) MCF-7, T47D, MDA-	Epigenetic mechanisms	<ul> <li>-in vivo, BPA increases EZH2 expression (mRNA, 6-fold increase, and protein) in the mammary gland. This effect is also observed with DES (x6) and E2 (x4.8)</li> <li>-in vitro, BPA (100 nM) increases the expression of EZH2 (mRNA et protein) in MCF-7 and T47D cells, but not in MDA-MB-231 (devoid of nuclear ER receptors). Results indicate an ER-mediated estrogenic effect of BPA.</li> <li>-molecular analysis of the EZH2 promoter in BPA-treated cells         <ul> <li>EZH2 promoter contains functional estrogen-response elements (ERE1 et ERE2) (luciferase-based reporter</li> </ul> </li> </ul>

			MB-231		construct) - Both ERα and ERβ are involved in the BPA-induced EZH2 expression (ChiP assay, chromatin immunoprecipitation). BPA increases the fixation of ER co-régulators such as MLLs (MLL2 et MML3, key enzymes involved in methylation and acetylation of histones H3K4) and CPB/P300 (histone acetylase HAT). These effects are also observed with DES and E2 - BPA increases the functional activity of EZH2, i.e., level of histone acetylation and H3K4 tri-methylation, at the EZH2 promoter, as well as polymerase II.
Hussain et al., 2015 (BBA)	Rat Sprague Dawley In vitro		Same protocol in vivo and in vitro as Bhan, 2014a and b Ovariectomised adult rats. BPA sc: 25μg/kg, 24h. Positive control: E2 and DES 5 μg/kg Cells: MCF-7, T47D, MDA-MB-231 BPA (several doses) 4h	Developmental genes	BPA induced HOXC6 expression both in vivo and in vitro. HOXC6 is a homeobox-containing gene associated with mammary gland development and is overexpressed in a variety of cancers including breast cancer.  - In vivo BPA (25 μg/kg) increases HOXC6 gene expression (mRNA and protein in the mammary gland. These effects are also observed with E2 and E2+BPA  -In vitro BPA (10 nM to 100nM) increases HOXC6 expression (x6) in ER-positive breast cancers cells (MCF-7 and T47D), but not in ER- cell line, indicating a potential regulation by ER. This effect is also observed with E2 (1 nM).  -molecular analysis of the HOXC6 promoter in BPA-treated cells (same protocol as for EZH2 and HOTAIR). Along with ERα, co-activators such as MLL2 and MLL3 and CBP/p300 (histone acetylase) bind to an HOXC6 promoter ERE region upon treatment with BPA affecting its gene expression. This effect is also observed in E2-treated cells; reversed in ERα-knock-out BPA (or E2) treated cells
<b>Deb</b> <i>et al.</i> , <b>2016</b> (Gene)	Rat Sprague Dawley In vitro	Same protocol in vivo and in vitro as Bhan, 2014a and b Ovariectomised adult rats.  BPA sc: 25µg/kg, 24h. Positive control: E2 5			BPA induced HOXB9 expression both in vivo and in vitro. HOXB9 is a homeobox-containing gene that play a key role in mammary gland development and is associated with breast and other types of cancer. It is involved in several processes including cell proliferation, cell-cycle progression.

CF-7 oses 0-BPA n (Ctrl: E2)	-In vivo BPA (25 μg/kg) increases HOXB9 expression (x4) in the mammary gland of ovariectomised rats. In vitro BPA (100nM) induces HOXB9 expression (x9.4) in MCF-7 cells. While E2 also increases HOXB9 expression, BPA counteracts the level of E2-induced HOXB9 in comparison to E2 alone.
	-HOXB9 promoter ERE is responsive to BPA. Molecular analysis of the HOXB9 promoter in BPA- treated cells (same protocol as for EZH2 and HOTAIR). Along with ER, ER-co-activators such as MLL3 and CBP/p300 (histone acetylase) bind to the HOXB9 promoter in the presence of BPA, modify chromatin and lead to its gene activation even in the absence of estradiol

## 4.4 Alteration of brain development and cognitive function

## 4.4.1 Overview of previous evaluation and focus on a specific effect

# Overview of the effects of BPA on the brain and behaviour based on previous works

A summary of the toxic effects of BPA on the brain and behaviour is presented hereafter on the basis of the information collected in the restriction proposal of BPA in thermal paper (ANSES, 2014).

**In animals,** several studies investigated brain functionality and behaviour following pre-, post- and/or perinatal exposure to BPA.

The studies examining the effects of pre- or perinatal exposure to BPA **on anxiety** were conducted at a wide range of exposure levels, thereby making it difficult to establish a comparison between the results achieved. Indeed, BPA has been shown to have no effects (Stump, 2010), to increase the level of anxiety (Cox, 2010; Poimenova, 2010) or to reduce it (Tian, 2010). It is not known whether an 80-fold difference in the administered doses can explain the different effects induced by BPA. Anxiety-related behaviour can be modulated by various factors including the chemical properties of the compound, the behavioural task used and the experimental protocol. Therefore, no clear conclusion can be drawn. It is however noted that many studies have been recently published on BPA and anxiety but they were not included in the present overview.

Impairment of **memory** including spatial and avoidance abilities was observed in mice exposed to BPA (0.5-50 mg/kg/d) during gestation and lactation (Xu *et al.*, 2010b). In addition, spatial memory performance was also reduced in both male and female rats exposed to a lower dose of BPA (40  $\mu$ g/kg/d) during the same periods (Poimenova *et al.*, 2010).

Suppression of **sexual dimorphism in the neural structures involved in sexual behaviour** was reported in offspring mice orally exposed *in utero* to 8 mg/kg/d (Cox *et al.*, 2010). However, it was not observed in two rat studies using respectively 2 to 200  $\mu$ g/kg/d BPA by oral route during gestation and lactation and 50  $\mu$ g or 50 mg/kg/d BPA by subcutaneous injections during PND0-3 (Ryan *et al.*, 2010 and Adewale *et al.*, 2009).

Suppression of **sex differences** was identified in rat offspring with regard to their locomotor activity and exploratory behaviour in the open-field (Kubo *et al.*, 2003; Rubin *et al.*, 2006; Palanza *et al.*, 2008), activity and avoidance memory (Kubo *et al.*, 2001), and social novelty behaviour (Cox *et al.*, 2010; Palanza *et al.*, 2008). In addition, decreased exploratory behaviour was observed only in females in Poimenova *et al.* (2010). These effects have been observed in experimental studies using a dose-range of 25 to 250 ng/kg/d BPA by subcutaneous route (Rubin *et al.*, 2006) and 10 µg to 8 mg/kg/d BPA by oral route (all other studies).

**Changes in maternal behaviour** related to pre- or postnatal exposure to BPA (10  $\mu$ g/kg/d to dams by oral route) have also been reported (Palanza *et al.*, 2002).

Effects on **cerebral development** linked to pre- or perinatal exposure to BPA have been demonstrated in several studies including changes in neural differentiation (Funabashi *et al.*, 2004; Patisaul *et al.*, 2007; Rubin *et al.*, 2006), alterations of the glutamatergic NMDA receptors and aminergic systems (Tian *et al.*, 2010; Matsuda *et al.*, 2010; Xu *et al.*, 2010a and 2010b), changes in the expression of estrogen receptors (ER)  $\alpha$  and ERB (Xu *et al.*, 2010b; Mahoney *et al.*, 2010), and changes in the number of neurons

responsive to oxytocin and serotonin (Adewale *et al.*, 2009). These changes occurred in particular regions like the hypothalamus (more precisely in sexually dimorphic areas) and the hippocampus, a region involved in cognitive processes, namely those linked to NMDA receptors.

These neural effects were identified in experimental studies following prenatal or neonatal exposure to 25 ng/kg/d to 50 mg/kg/d BPA by subcutaneous route and to 0.05 to 50 mg/kg/d BPA by oral route. Thus, they could explain and support, at least in part, the behavioural effects observed concomittantly.

## Assessment by RAC under the restriction process

In its opinion of June 2015 (ECHA, 2015), RAC adopted conclusions on the analysis of BPA-induced effects on brain and behaviour as quoted in the text box hereafter:

"The Dossier Submitter considered the oral study by Xu et al. (2010) in mice as the key study for neuro-developmental toxicity. The critical effects in this study were the alteration of memory and learning functions paralleled by a decrease in the expression of glutamate NMDA receptors.

The EFSA (2015) opinion concluded on neurological, neurodevelopmental and neuroendocrine effects as follows: "[...] In summary, there are indications from prospective studies in humans that prenatal BPA exposure (BPA exposure during pregnancy) may be associated with altered child behaviour in a sex-dependent manner. However, the associations were not consistent across the studies and it cannot be ruled out that the results are confounded by diet or concurrent exposure factors. The associations reported do not provide sufficient evidence to infer a causal link between BPA exposure during pregnancy or childhood and neurodevelopmental effects in humans.

A number of new studies report changes that may indicate effects of BPA on brain development (effect on neurogenesis and on gene expression, neuroendocrine effects, effects on the morphology of certain brain regions, etc.). Whether such changes are mechanistically related to the neurobehavioral responses reported following exposure is attempted addressed by some studies but with inconsistent results.

Several new animal studies investigated anxiety-like behaviour, learning and memory, social behaviour and sensory-motor function. Some studies report changes in anxiety-like behaviour after BPA exposure. Some, but not all, studies reported significant impairment of either learning and/or memory capacities. A few studies also report effects on social behaviour and sensory-motor function. However, the studies present methodological shortcomings, such as small sample size, lack of consideration of the litter effect, not properly controlled variability of exposure through diet and inadequate statistics. Using a WoE approach, the CEF Panel assigned a likelihood level of "as likely as not" to neurological, neurodevelopmental and neuroendocrine effects of BPA. [Since the likelihood]<sup>17</sup> level for this endpoint is less than "likely" (see Appendix A), this endpoint was not taken forward for assessing the toxicological reference point, but was taken into account in the evaluation of uncertainty for hazard characterisation and risk characterisation".

See sections 3.4 and 4.3 of the EFSA (2015) opinion for more details.

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<sup>&</sup>lt;sup>17</sup> Included by RAC for clarification.

RAC considers that the results from the Xu *et al.* (2010) study suggest that developmental exposure to BPA can interfere with learning and memory capacities in different learning tasks in rodents, including spatial learning and passive avoidance learning together with down-regulation of the NMDA receptors. However, the effects of BPA on learning and memory abilities of laboratory rodents are not fully consistent, as both positive and negative effects are reported in different studies.

Two studies that were not included in the restriction report or in EFSA (2015) were submitted during public consultation (Elsworth *et al.*, 2013 and Ferguson *et al.*, 2014). Elsworth *et al.* (2013) showed effects on brain development (loss of midbrain TH-immunoreactive neurons and loss of hippocampal spine synapses) in non-human primates at low BPA doses. No alterations in sexually dimorphic behaviors in male and female Sprague-Dawley rats were observed by Ferguson et al. (2014).

#### Conclusion

RAC in principle agrees with EFSA's conclusion on effects on brain and behaviour. Since effects on brain and behaviour have been observed at and below the range where kidney effects occur, RAC considers it prudent to take them into account in hazard and risk assessment and in health impact assessment. RAC however acknowledges that the available information does not allow a quantification of the dose-response relationship, therefore this endpoint will be accounted for in the setting of Assessment Factors."

# Focus on a specific effect of BPA on the brain based on previous assessments and recent publications

More recent studies have investigated the effects of exposure to BPA on behaviour. In particular, alteration of learning and memory was confirmed in several new studies and it appears as the endpoint with the most convincing evidence among the other central effects induced by exposure to BPA as well as the endpoint with the most specific indications of its link with endocrine disruption.

Therefore, also for the sake of clarity, considering the extent of the available toxicological data, the present section addresses in detail only alteration of learning and memory ability. This adverse effect which is now well established is the focus of the analysis below. The following section analyses i) the ability of BPA to affect learning and memory in detail and ii) the endocrine basis of these changes.

For this purpose, studies addressing BPA effects on learning/memory and its MoA were collected up to May 2016.

#### 4.4.2 Adverse effects

# Background on learning and memory and underlying cellular and molecular mechanisms

Learning and memory is the function by which individuals acquire knowledge about their environment and refers to the process by which the learned information is encoded, stored and later retrieved. Several brain areas are involved in learning and memory (cortex, hippocampus, amygdala...) and damage to a given region may cause the loss (partial or complete) of a specific function. The medial temporal system of high vertebrates includes the hippocampus, which is one of the most important areas involved in the processing of memory. Neurological studies in human patients and in rodents established the central role of this brain region for contextual memory. The hippocampal formation was associated with a wide variety of cognitive functions such as spatial navigation and planning, memory encoding and retrieval, relational processing, and

novelty detection. The rodent hippocampus was also shown to play a key role in both spatial and non-spatial memory (visual object recognition, temporal processing of information...).

At the cellular level, learning and memory involve various neuronal plasticity mechanisms, which include long-term potentiation, synaptogenesis, modulation of intrinsic excitability, adult neurogenesis in the dentate gyrus and modulation of the glutamatergic neurotransmitting system. At the molecular level, memory involves changes, which have been extensively studied in rodents, especially in glutamatergic Nmethyl-D-aspartate (NMDA) and q-amino-3-hydroxy-5-methyl-4-isoxazolepropionic acid (AMPA) receptors. The NMDAR is an ion channel protein composed of four subunits (including GluN1 and GluN2 subunits) and stimulated by the glutamate neurotransmitter. These receptors contribute to the induction of synaptic plasticity, are also involved certain types of brain neuronal plasticity and are necessary for the encoding of many types of memory. It has been shown that the hippocampus is essential for spatial learning and memory and is subjected to the induction of long-term potentiation involved in synaptic plasticity. The detailed steps of these processes are presented in Figure 15. Among the required kinases, phosphorylated ERK and CamKIV translocate into the nucleus to phosphorylate the transcription factor CREB. Phosphorylated CREB induces gene transcription and protein synthesis necessary for structural changes at existing synapses as well as synaptogenesis.

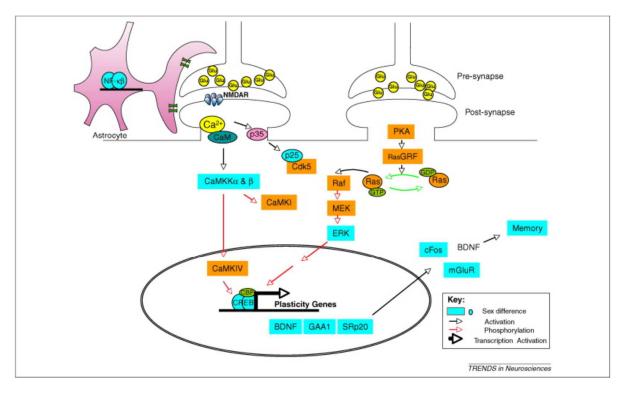


Figure 15: Steps involved in induction of long-term synaptic plasticity at various hippocampal synapses in rodents. From Mizuno & Giese, 2010

At glutamatergic synapses, activation of NMDA receptors induces Ca<sup>2+</sup> influx into the postsynaptic cell. This triggers activation of various synaptic kinases and induction of gene transcription (C/EBPB, EPA, BDNF) by phosphorylated CREB followed by protein synthesis. Sex differences are highlighted in blue. Abbreviations- BDNF: brain-derived neurotrophic factor; CaM: calmodulin; CaMKI: Ca<sup>2+</sup>/CaM kinase I; CaMKIV: Ca<sup>2+</sup>/CaM kinase IV; CaMKK: Ca<sup>2+</sup>/CaM kinase kinase; CBP: CREB-binding protein; Cdk5: cyclin-dependent kinase 5; CREB: cAMP-responsive element binding protein; ERK/MAPK: extracellular signal-regulated kinase/ mitogen-activated protein kinase; GAA1: glycosyl phosphatidyl-inositol anchor attachment protein; Glu: glutamate; GRF: guanine nucleotide-releasing factor; MEK: mitogen-activated protein kinase kinase; NMDAR: NMDA

receptor; NF- $\kappa\beta$ : nuclear factor  $\kappa\beta$ ; mGluR: metabotropic glutamate receptor; PKA: cAMP-dependent protein kinase; SRp20: splicing factor arginine/serine-rich 3.

Learning and memory processes are very sensitive to both endogenous (neurotransmitters, hormonal systems) and exogenous factors (e.g. stress, drug abuse). Gender-specific differences are observed in the execution of behavioural tasks. Sex modulations in the molecular pathways involved in memory processes are highlighted in blue in Figure 15.

### 4.4.2.1 Non-human information

Studies investigating the effects of exposure to BPA on learning and memory as well as potential related alterations in brain regions underlying these processes are summarised in Table 21 presented in section 4.4.5. They are discussed by type of exposure (developmental, prepubertal and pubertal, adult) and species studied (mice, rats, then other species).

## **Developmental exposure**

Twenty-one studies addressed the developmental effects of exposure to BPA on learning and memory in rodents (8 in mice, 13 in rats).

- **Spatial memory** was assessed in 16 studies by using dedicated paradigms (Morris water maze, Barnes maze, Y maze, or radial maze). About 75% of these studies(12/16): 5 in mice (Xu et al., 2010b; Jasarevich et al., 2011, 2013; Jang et al., 2012; Kumar et al., 2014) and 7 in rats (Poimenova et al., 2010; Goncalves et al., 2010; Kuwahara et al., 2013; Xu et al., 2014b; Wang et al., 2014b; Johnson et al., 2016; Hass et al., 2016) reported learning and memory disturbances in adult animals developmentally exposed to BPA, whereas no effects were observed in 4 studies: 2 in mice (Nakamura et al., 2012; Viberg et al., 2011) and 2 in rats (Stump et al., 2010; Jones et al., 2012). Among these studies, 8 analysed spatial memory in both males and females. Four studies (2 in mice, 2 in rats) reported sex differences in BPA-induced effects (Jasarevic et al. 2011; Jasarevic et al., 2013; Goncalves et al., 2011; Jonhson et al., 2016), while a similar memory deficit in both sexes was observed in 1 rat study (Poimenova et al., 2010). No effects in either sex were reported in 3 analyses (1 in mice and 2 in rats) (Nakamura et al., 2012; Stump et al., 2010; Jones et al., 2012).

At the molecular level, the behavioural alterations induced by developmental exposure to BPA were associated with reduced hippocampal expression of NMDAR subunits (Xu et al., 2010a and 2010b) and changes in hippocampal ER levels (Xu et al., 2010b; Xu et al., 2014b). Modifications in levels of corticosterone and their receptors (Poimenova et al., 2010), as well as changes in hippocampal expression of two cell adhesion proteins (Nrxn1 and Nlgn3) involved in synaptic plasticity (Kumar et al., 2014) were also reported.

- **Short-term or long-term memory** was analysed in 5 studies: 1 in mice (Xu *et al.*, 2010b) and 4 in rats (Kubo *et al.*, 2001; Goncalves *et al.*, 2010; Xu *et al.*, 2011a; Kuwahara *et al.*, 2013) by using the passive avoidance test. They all described memory impairment in males and females developmentally exposed to BPA, with one study reporting sex differences in BPA-induced effects (Kubo *et al.*, 2001). Reduced hippocampal expression of NMDAR subunits was found in the same study assessing long-term effect of early exposure to BPA (GD7-PND21) on both spatial memory and short-term/long-term memory (Xu *et al.*, 2010b). In another study investigating the effects of a single exposure to BPA (1 single injection at PND18 and males analysed 1h or 24h later), the levels of NMDAR subunits were not affected, but their phosphorylated levels

were increased (Xu et al., 2011a). These differences in response in NMDAR expression are very likely to be related to the duration of exposure to BPA (single vs repeated exposure). Repeated exposure to BPA (GD6 to GD17) also triggered a decrease in adult neurogenesis in the dentate gyrus of developmentally exposed females, together with a decrease in hippocampal levels of phosphorylated ERK and CREB, and brain-derived neurotrophic factor (BDNF) (Jang et al., 2012), three key actors of synaptic plasticity and memory processing.

- **Visual memory** recognition was analysed in two rat studies, which reported behavioural impairements in males and females in one study (Goncalves *et al.*, 2010), and only in males in the other (Wang *et al.*, 2016) following exposure to BPA. In Wang *et al.* (2016), the levels of Akt, MAPK and their phosphorylated forms were reduced, and phosphorylated levels of BDNF and CREB inhibited, indicating inhibition of processes involved in synaptic plasticity.
- **Fear memory** was enhanced in females following exposure to BPA from GD10 to PND20 (Matsuda *et al.*, 2013). Serotonin metabolite concentrations were increased in hippocampus, striatum, midbrain, pons and medulla oblongata of juvenile females but not in adult females or juvenile/adult males. The expression levels of Tph2, Slc6a4, monoamine oxydases (enzymes involved in serotonin synthesis, transport and metabolism) were also enhanced.

One study was performed on a non-human primate model (Rhesus macaque) with the objective to compare BPA effects during prenatal exposure or juvenile stage (Elsworth *et al.*, 2013). BPA during fetal development of Rhesus macaques reduced the number of dopaminergic neurons or of spine synapses in the hippocampal CA1 region, but not in the prefrontal cortex. No behavioural evaluation was conducted in the part of the study where animals were exposed developmentally.

### Prepubertal and pubertal exposure

**Spatial memory** was tested in 6 rodent studies (4 in rats, 2 in mice) using the Morris water maze and object recognition placement test. Impaired memory was found in 4/6 studies (Kim *et al.*, 2011., Xu *et al.*, 2011b; Jain *et al.*, 2011; Diaz Weinstein *et al.*, 2013), with reported sex differences in the study analysing both sexes (Xu *et al.*, 2011b). Memory assessed by the passive avoidance and object recognition tests was found affected by prepubertal/pubertal exposure to BPA (Xu *et al.*, 2011b, Jain *et al.*; 2011; Bowman *et al.*, 2015; Diaz Weinstein *et al.*, 2013). One study described no effect on **fear memory** (Kuwahara *et al.*, 2014).

Reduced adult neurogenesis in the dentate gyrus (Kim *et al.*, 2011) and spine density in the hippocampal CA1 area as well as an oxidative stress (Jain *et al.*, 2011) were reported in these studies.

The Elsworth study performed in juveniles of African green monkeys found no effect by using a cognitive task that tests working memory capacity (Elsworth *et al.*, 2013). The number of dopaminergic neurons and of spine synapses remained unaffected in the hippocampus, contrary to the decrease noted when BPA was administered during fetal development. It is, however, important to indicate that this study presents a limitation due to the analysis of animals without taking account of their sex and that sex-specific effects may hence be missed.

#### Adult exposure

**Spatial memory** was analysed in 5 rodent studies (2 in mice, 3 in rats) by using the Morris water maze and the object recognition placement test. Out of 5 studies, 4 showed impaired memory following adult exposure to BPA (Xu *et al.*, 2013b; Eilam-Stock *et al.*,

2011; Inagaki et al., 2012; Fan et al., 2013).

**Short-term and long-term memory**, as assessed by the passive avoidance test, was also impaired in mice (Xu *et al.*, 2013b). One study in African vervet male monkeys using the two-choice spatial delayed response task showed impaired memory (Elsworth *et al.*, 2015). The number of spine synapses was diminished in the prefrontal cortex and CA1 region.

At the neuroanatomical level, these behavioural alterations were associated with disrupted synaptic density and histological alterations in the hippocampus and/or prefrontal cortex in rodents (Inagaki *et al.*, 2012; Xu *et al.*, 2013b; Eilam-Stock *et al.*, 2011) and primates (Elsworth *et al.*, 2015). At the molecular level, the expression level of NMDAR and AMPAR subunits and synaptic proteins were affected (Xu *et al.*, 2013b; Eilam-Stock *et al.*, 2011). Reduced acethylcholine esterase was also observed in the hippocampus (Fan *et al.*, 2013).

In one study analysing spatial (Morris water maze) and non-spatial memory (passive avoidance test) in ovariectomised female mice, BPA treatment suppresses the reduction in abilities observed in ovariectomised animals (Xu et al., 2015b). An enhancement of capacities was observed in male mice in a fear-conditioning task (Zhang et al., 2014). Changes in expression levels of NMDAR and synaptic proteins were observed in these studies.

#### 4.4.2.2 Human information

## **Epidemiological studies**

While several epidemiological studies investigated the association between perinatal exposure to BPA and child behaviour, only two studies investigated a link between exposure to BPA and the cognitive performance of children more specifically (Casas *et al.*, 2015; Hong *et al.*, 2013).

Casas et al. (2015) assessed prenatal exposure to BPA by urinary measurements in two spot samples collected in the first and third trimester of pregnancy in 438 pregnant women from the Spanish mother-child cohort INMA-Sabadell (median, 25th-75th percentiles; 1st-trimester sample: 2.1, 1.2-3.7 µg/L; 3rd-trimester sample: 1.8, 1.0-3.1 μq/L). Cognitive and psychomotor abilities were assessed by psychologists at both 1 and 4 years of age with standardised neuropsychological tests (Bayley Scales of Infant Development BSID at age 1, McCarthy Scales of Children's Abilities (MSCA) at age 4; expected mean (standard deviation)=100(15)). Numerous potential confounders, including socioeconomic status or other well-established neurotoxicants (PCBs and methylmercury during pregnancy), as well as strong determinants of children's neuropsychological abilities such as maternal intelligence score, were taken into account, some of them having been assessed prospectively. Multivariable models were conducted and non-linearity was considered. A statistically significant decrease of the psychomotor score at age 1 was observed among the highest-tertile urinary BPA concentrations, compared to the lowest-tertile concentrations (beta=-4.3, 95% CI:-8.1,-0.4), while no association was observed with the MSCA motor score at age 4. No association was observed between prenatal urinary BPA concentrations and the cognitive scores of children aged 1 or 4 years. No sex interaction was observed.

Hong et al. (2013) assessed childhood BPA exposure from urinary measurements in first-morning spot sample of 1008 children aged 8-11 years recruited from school lists of 5 different regions in Korea (median BPA level was 1.2 ug/L). The learning performance of the children was assessed using the parent-reported questionnaire (Learning Disability Evaluation Scale; LDES) and an abbreviated form of the Korean Educational

Development Institute's Wechsler Intelligence Scales for Children (KEDI-WISC) was administered. Several potential confounders, including socioeconomic status or other well-established or suspected neurotoxicants (lead during childhood, phthalates) were taken into account. Multivariable models were conducted and non-linearity was considered. Statistically significant adverse associations were observed between the urinary BPA concentrations and the parent-reported learning score (LDES). No association was observed between the urinary BPA concentrations and the intelligence scores of the children.

BPA has a short biological half-life and intra-day and inter-day variabilities of urinary BPA concentration are well-known. The findings of the existing literature are thus limited by the use of single spot urinary BPA measurement that might mostly reflect BPA exposure during the preceding hours or days. Note that some authors expect that large sample size, as observed in these two studies, might help to limit the noise and the potential bias induced by exposure measurement.

## In vitro human study

An *in vitro* study showed that exposure to BPA (100 nM) of primary human cortical neurons, from 16-21 week-old fetal brain, decreased mRNA levels of potassium chloride co-transporter 2 (KCC2) (Yeo *et al.*, 2013). This potassium chloride co-transporter is responsible for chloride extrusion from mature neurons. During CNS development, chloride concentration is of fundamental relevance for migration of cortical inhibitory precursor neurons to their proper final locations in the brain's cortex. KCC2 expression increases during the period preceding the chloride shift, which characterises mature neurons. Similar effects were observed for rat and mouse in the same study. The delayed perinatal chloride shift induced by BPA was mediated through an epigenetic modification of the Kcc2 gene. Effects were more accentuated in females than in males.

## 4.4.2.3 Summary and discussion of effects on brain and behaviour

Epidemiological evidence on the potential role of exposure to BPA early in life on learning and memory performance is still insufficient. However, about 74% of rodent studies (total of 26/35) reported impaired spatial and non-spatial memory following exposure to BPA, regardless of the period of exposure. In the remaining 26% of studies, there was either no effect (6/35) or enhanced performance (3/35). The causes of these differences are difficult to address given the differences in the experimental conditions between studies (sex, period of exposure, BPA doses), but the overall evidence has recently substantially increased and altogether strongly points toward the conclusion that BPA alters memory in rodents. In addition, as the 74% of studies reporting impaired cognitive behaviours were also performed under various experimental conditions (various doses, routes and period of exposure and test species), this means that this impairment is a robust BPA-induced effect.

Another interesting finding which can be drawn from this analysis, is the sex-dependent effect observed in a number of studies. Furthermore, although not systematically assessed in all these studies, the neural mechanisms associated with the behavioural alterations consist of a reduction in the level of expression of NMDAR subunits, kinases, enzymes involved in neurotransmitter regulation, and synaptic proteins as well as decreased spine density or neurogenesis. Such molecular, cellular and structural changes are fully relevant and could underlie the impaired learning and memory performance observed in the same animals.

Finally, although there is a limited number of studies conducted in non-human primates, these studies have shown that BPA during the prenatal stage of development has detrimental effects on the midbrain DA system and on spine synapses in the

hippocampus, while it has no effect when applied at a juvenile stage (Elsworth *et al.*, 2013). They also indicated in adult BPA-exposed monkeys a potential for significant cognitive impairment (Elsworth *et al.*, 2015).

It is also interesting to note that down-regulation of estradiol-induced increase in spine synapses in the hippocampus and prefrontal cortex in adult ovariectomised monkeys was also reported in another study of the same laboratory that is discussed in the next section 4.4.3.2 (Leranth *et al.*, 2008).

Overall, BPA has been demonstrated to alter memory and learning after developmental, pubertal or adult exposure, in multiple converging experimental studies reporting this functional effect as well as molecular and cellular changes in the brain in line with the functional changes observed.

## 4.4.3 Endocrine disruption

## 4.4.3.1 In vitro information indicative of an endocrine MoA

Few studies have investigated the potential MoA of BPA in neural development *in vitro* or *ex vivo*. The data are summarised in Table 22 (presented in section 4.4.5).

In cultured hippocampal neurons from 24-hr old rats, BPA (10 and 100 nM) promoted dendritic development (length of dendrites and mobility and density of dendritic filopodia) (Xu et al., 2014a). The promoting action of BPA on dendritic development was completely blocked by the ER antagonist ICI 182,780. The effect of BPA could be exerted through enhancement of F-actin cytoskeleton (critical in morphological changes occurring in dendritic filopodia and spines during development and synaptic plasticity), and modifications in expression levels of Rho proteins (involved in intracellular actin regulation). These changes were shown to be mediated through ER- via ERK1/2 signaling pathways since they were suppressed by the ER antagonist.

In another study with rat cultured hippocampal neurons, BPA (10-1000 nM) rapidly increased the mobility and density of dendritic filopodia and phosphorylation of the NMDAR subunit N2B known to be primarily expressed in immature synapses of the hippocampus during postnatal development (Xu *et al.*, 2010c). The ER antagonist ICI 182,780 completely reversed BPA-induced effects on filopodia and GluN2B subunit activation.

An  $ex\ vivo$  study confirmed the action of BPA on ER observed  $in\ vitro$ . In cultured adult rat hippocampal slices, BPA (10 nM) enhanced long-term depression of synaptic transmission (Hasegawa  $et\ al.$ , 2013). This effect was suppressed by 4-OH-tamoxifen, an estrogen-related receptor gamma (ERR $\gamma$ ) antagonist, but not by ICI 182,780 (ER antagonist).

In conclusion, despite their limited number, *in vitro* studies provide evidence that BPA affects dendritic development and synaptogenesis in hippocampal rat neurons (Xu *et al.*, 2010c; Xu *et al.*, 2014a) through changes in ER-dependent pathways. Neural development and cerebral plasticity is based on a subtle balance between cell proliferation and apoptosis. The effects observed *in vitro* with BPA, if they occur *in vivo* may induce neural dysfunction. The *ex vivo* study performed by Hasegawa *et al.* (2013) extends the effects of BPA to other estrogen-receptor signaling pathways such as ERR- $\gamma$ . This might suggest that estrogens act through multiple pathways, which can be impacted by BPA.

## 4.4.3.2 *In vivo* effects with regard to an endocrine MoA

Several of the studies that observed an effect of BPA in the alteration of learning and memory (presented also in Table 23 in section 4.4.5 as discussed before) were also considered to provide relevant information on a potential endocrine MoA and are summarised in more detail below.

Developmental exposure: In rats, Xu et al. (2014b) described a direct link between the behavioural effects induced by BPA and an estrogenic endocrine disruption. Male rats were exposed to BPA from the last gestation day to PND21. BPA impaired spatial memory and increased exploratory activity. These effects were associated with decreased expression of ER $\alpha$  (mRNA and protein levels) at PND7 and 21 and increased expression at PND11 in the hippocampus of both hemispheres. All these effects were completely reversed by the ER antagonist ICI 182,780. Detailed analysis of the molecular mechanisms showed that BPA impaired ER $\alpha$  translocation to the nucleus and decreased the phosphorylation of nuclear and total ER $\alpha$ , which were also reversed by ICI 182,780.

In the hippocampus of mice, exposure to BPA decreased the expression of GluN1, GluN2A and GluN2B subunits of NMDAR and ER $\beta$ . These effects were associated with impaired spatial and avoidance memory (Xu *et al.*, 2010b).

<u>Postnatal exposure</u>: In another study, Xu *et al.* (2011a) reported that single subcutaneous injection of 18-day-old male rats with BPA alone or EB alone increased the latency in retention test in the passive avoidance task at 1h but not 24h after exposure. These behavioural changes were associated with increased levels of phosphorylated NMDAR subunits. Interestingly, when males were exposed to both substances, no effect was observed as if there was a blockade of estradiol-induced responses by BPA. Pretreatment with ER antagonist ICI 182,780 inhibited BPA- or EB-induced effect on phosphorylated NMDAR subunits.

Adult exposure: The behavioural disturbances induced by EB in ovariectomised mice were partially blocked by BPA (Xu et al., 2015b). Co-treatment with BPA also blocked the estradiol-induced modifications of synaptic interface and expression of synaptic proteins in the hippocampus. In agreement with these data, BPA blocked estradiol-enhanced object placement memory in ovariectomised female rat, and object recognition memory performance during proestrus (high levels of estradiol) in intact females (Inagaki et al., 2012). In the same study, BPA altered estradiol-induced increase in spine density of pyramidal neurons in the hippocampus. These rodent studies are consistent with the data published by Leranth et al. (2008) showing that BPA blocked the estradiol-induced increase of spine synapses in the prefrontal cortex and hippocampus of ovariectomised female monkeys.

Altogether, these data point to a MoA for BPA targeting the estrogenic pathway with alterations of learning and memory as a consequence.

- Other studies, that did not address the behavioural effects of exposure to BPA on memory, provided additional ED-related evidence of cerebral changes in brain areas underlying learning and memory. They are summarised below and in Table 23 (presented in section 4.4.5).

In the hippocampus of male rats, BPA reduced the expression of GluN1, GluN2A and GluN2B NMDAR subunits (Xu *et al.*, 2010a). These changes were associated with a decreased expression of ER $\beta$  and a dose-dependent increase of aromatase cytochrome P450 (P450arom).

In the brain of mice orally exposed twice at GD6 and GD15 to BPA, expression of genes

involved in neurodevelopment was decreased (*Xist*, *Gdi1*, *Nlgn3* and *Park3*, *Fmr1*) (Kumamoto & Oshio., 2013). These effects were associated with reduced expression of the AR gene and the mRNA levels of ER $\beta$ , and the increased expression of ER $\alpha$ .

In several studies, exposure to BPA has been shown to modulate the expression of key steroidogenic enzymes in different cerebral structures. In the prefrontal cortex of adult rats orally exposed, BPA decreased the expression levels (mRNAs and protein) of  $5\alpha$ -reductase 2 ( $5\alpha$ -R2) and  $5\alpha$ -R3 (Castro *et al.*, 2015), two enzymes involved in testosterone metabolism. When exposed subcutaneously for 4 days during adulthood, the expression (mRNAs and protein levels) of  $5\alpha$ -reductase 1 ( $5\alpha$ -R1) was reduced in females but not in males (Castro *et al.*, 2013). BPA also increased the expression of the aromatase gene, involved in testosterone metabolisation into neural estradiol, in males and females and the aromatase protein levels in males.

In adult mice (10-week-old) orally exposed during 12 weeks, BPA decreased the expression of ER $\beta$  protein in males but not in females. These effects were associated with increased anxiety state level and decreased brain levels of testosterone in males. No effect on brain and serum levels of estradiol was observed in females (Xu *et al.*, 2015a).

An *in vivo* study was conducted on adult female African green monkeys, which were ovariectomised and implanted with the vehicle, EB alone, BPA ( $50 \mu g/kg/d$ ) alone or EB plus BPA for 20 days (Leranth *et al.*, 2008). Estradiol induced a synaptogenic effect, in hippocampal regions (CA1, CA3 and dendate gyrus (DG)) and prefrontal cortex, which was abolished by the continuous exposure to BPA.

# 4.4.3.3 Summary of the plausible link between adverse neural effects and endocrine MoA

BPA disrupts learning and memory processes. The cognitive effects induced by BPA are associated with the disruption of two important pathways in cerebral regions: i) NMDAR, their down-stream targets leading to gene transcription (ERK, CREB, BDNF...) and synaptic proteins involved in synaptic plasticity, and ii) estrogen-dependent pathways involving  $ER\alpha$  or  $ER\beta$ .

The possibility that BPA alters the cellular and molecular pathways involved in learning and memory processes through disruption of estrogen-dependent pathways was first suggested in the study of Xu *et al.* (2010a and 2010b). In this study, decreased expression of ER $\beta$  was also reported, but no demonstration was given of a potential link between BPA-induced effects on memory and estrogenic pathway alteration. In the study of Xu *et al.* (2014b), the link between BPA-induced effects on learning and memory processes and estrogenic pathway disruption was clearly established from the demonstration that the ER antagonist ICI 182,780 reversed both BPA-induced effects on ER $\alpha$  (modulation and regulation) and memory. Three other studies performed in adults indicated that BPA is also able to interfere with estradiol-induced effects on behaviour and spine density in rodents (Xu *et al.*, 2015b; Inagaki *et al.*, 2012) and on synaptogenesis in non-human primates (Leranth *et al.*, 2008). Additional evidence was provided by *in vitro* studies showing that BPA-induced effects on NMDAR signalling and synaptic proteins were reversed by the ER antagonist. In one *in vitro* study, BPA-induced disruption extended to other non classical estrogen receptors (ERR $\gamma$ ).

The modulatory effects on cognitive behaviours and processes by estrogens are now well established, although they were studied more extensively in females than in males (for reviews in females, see Galea *et al.*, 2013; Pawluski *et al.*, 2009). The importance of the estrogenic pathway in the regulation of cognitive behaviour and synaptic plasticity has

also been reported in male rodents (Picot et al., 2016). In the male nervous system, may act directly or through its non-aromatisable metabolite testosterone dihydrotestosterone to activate the AR. Testosterone can also be aromatised locally into neural estradiol, which stimulates ERs. In this context, it has been shown that performance in spatial learning and memory abilities, such as object recognition, fear conditioning and spatial memory tasks are decreased by castration and restored by testosterone replacement. The estrogenic component of this regulation in males is demonstrated in the following studies. Intra hippocampal injection of estradiol enhanced memory in a spatial water maze task, possibly through an interaction with muscarinic cholinergic systems (Packard et al., 1996). Acute treatment using estrogens, estrogen receptor agonists or selective estrogen receptor modulators were shown to facilitate long-term potentiation in adult hippocampal slices, affecting the number and shape of dendritic spines in CA1 pyramidal neurons and decreasing thorn density of hippocampal CA3 neurons (Kramar et al., 2009). Estradiol through both ERa and ERB was also reported to regulate NMDAR-mediated transmission and thus synaptic plasticity in the dentate gyrus of juvenile males (Tanaka et al., 2013). In mice, activation of hippocampal  $ER\alpha$  after learning impaired memory formation in contextual fear conditioning tasks (Cho et al., 2015). Detailed effects of adult estrogens on learning and memory and the mechanisms underlying these effects in both males and females are described in recent reviews (Frick et al., 2015; Hamson et al., 2016). Figure 16 illustrates potential molecular mechanisms underlying estradiol-induced regulation of NMDAR signaling.

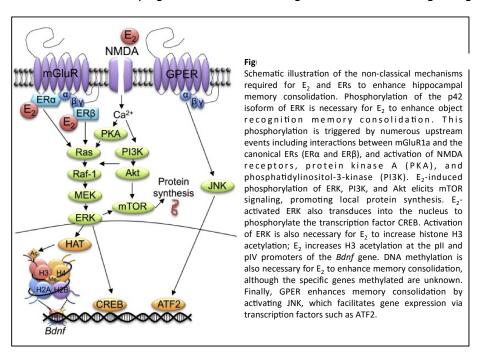


Figure 16 : Schematic illustration of the non-classical mechanisms required for E2 and ERs to enhance hippocampal memory consolidation (From Frick, Hormones & Behavior 2015)

In comparison with the numerous data reporting the role of sex steroid hormones in adults, fewer studies have addressed their developmental effects on learning and memory processes. Roof and Havens (1992) showed that perinatal testosterone might act by increasing the size of the granule cell layer of the hippocampus, thereby improving rat performance on a spatial navigation task during adulthood. Receptors of sex steroid hormones, and in particular ERs, are highly expressed during the perinatal period in the male brain, which is sensitive to the perinatal surges of testosterone. ERß knockout males exhibited memory impairment in a hippocampus-mediated fear-conditioning paradigm. ERß activation improved performance in hippocampus-dependent

memory tasks, enhanced long-term potentiation in hippocampal slices of wild-type but not ER $\beta$  knockout mice and increased dendritic branching and density of mushroom-type spines. In females, the perinatal brain is protected from the potential masculinising effects of sex steroid hormones by the alpha-fetoprotein. It is, however, important to note that ovarian estradiol increases from postnatal day 7 and could hence act on the female brain to regulate cerebral functions.

These sex differences in hormonal impregnation during the critical periods of development, and also during adulthood, may explain the sex differences observed in the expression of cognitive behaviours and their alteration by BPA.

Altogether, data therefore provide evidence that disturbance of, in particular, estrogenic pathways is involved in alteration of learning and memory by BPA.

## 4.4.3.4 Human relevance

Cognitive function in humans involves signaling pathways, which seem similar to those described above in rodents. The involvement of NMDAR signaling pathway in memory processes in healthy and diseased brain has been largely reviewed (e.g. in Gilmour *et al.*, 2012; Campos *et al.*, 2016; Arnsten *et al.*, 2016). The link between human cognitive functions and sex steroids, and the similarities and differences between humans and rodents are reviewed below.

#### Periods of sex steroid liberation in humans:

Sex steroids are synthesised from the developmental period in both humans and rodents. However, the periods of sensitivity to sex steroids seem to differ between mammalian species. Testosterone release by fetal testes peaks between gestational weeks 12 and 18 in humans whereas in rodents it occurs during the last days of gestation. Indeed, testosterone level rises again during the three first postnatal months in boys and in the first four hours after birth in the rat. In girls, unlike female rodents, the fetal ovaries are active and synthesise estrogens. It is, however, suggested that the female brain may also be protected from the masculinising effects of steroids by the alpha-fetoprotein. The following rise in sex steroids occurs during the pubertal period and secretion is maintained during adulthood in both sexes in humans and rodents.

Overall, there are some differences between humans and rodents in the timing of sex steroid secretion during brain development but in both species sex steroids are present during these critical developmental periods, in particular in males.

# Modulation of human cognitive processes by sex steroid hormones:

Estrogens were shown to modulate hippocampus-dependent learning in women and non-human primates (Hampson, 1990; Lacreuse, 2006., and review by Hamson *et al.*, 2016). For instance, the excitability of the hippocampus is increased with increased endogenous levels of estradiol as in rodents, and this seems to be correlated with changes in synapse density, synaptic proteins, and long-term potentiation.

Testosterone also modulates cognitive functions in men. Hypogonadism affecting young adults, chemically castrated individuals or older men is associated with spatial, visual, verbal and episodic memory defects. Testosterone levels have been linked to performance in visual and episodic memory tasks, with hypogonadic and elderly men performing poorly in such tasks. Spatial abilities were shown to be reduced in men undergoing combined flutamide (anti-androgen) and leuprolide (GnRH agonist decreasing LH and thus testosterone secretions) treatment (Cherrier *et al.*, 2009). Whether gonadal testosterone is converted into neural estradiol, which then regulates learning and memory processes needs to be better studied in men. However, one study showed that inhibition of aromatase (blocking the conversion of testosterone to

estradiol) prevented testosterone-induced improvement of verbal memory in older men (Cherrier *et al.*, 2005).

Given these similarities in the modulatory effects of sex steroids on cognitive functions between rodents and humans, it is likely that the MoA observed in rodents occurs in humans and has similar effects on these processes in humans to those in rodents.

Whether developmental exposure to BPA could affect these processes in humans is more difficult to answer given the lack of studies supporting potential effects of sex steroids in the developing human brain. One study addressing the prenatal vs juvenile exposure to BPA in non-human primates found no effect on working memory for the juvenile exposure, but no tests were performed in the group prenatally exposed to BPA. The significant impact of BPA on synaptic plasticity at the prenatal and adult stages and a deficit in working memory performance in adult monkeys suggest cognitive disruption in humans, given the analogies between primates: monkeys and humans share some uniquely primate morphological, endocrine and cognitive traits (Lacreuse and Herdon, 2009). In support of this hypothesis, the *in vivo* study conducted on adult female African green monkeys showed that a subcutaneous exposure to BPA (50  $\mu$ g/kg/d) counteracts the synaptogenic effect of estradiol in the hippocampus and prefrontal cortex (Leranth *et al.*, 2008). Finally, during adolescence, it has been shown, by using high-resolution structural MRI scans, that sex steroids are associated with cerebral gray matter morphology in a sex specific manner (Koolschijn *et al.*, 2014).

There are therefore indications that this MoA may be relevant to humans during developmental exposure.

## 4.4.4 Summary and discussion

On the basis of i) the significant amount of *in vivo* and *in vitro* animal data showing impairment of learning and memory by exposure to BPA and the potential alteration of cellular and molecular mechanisms underlying these processes through disturbance of the estrogenic pathway, ii) the similar types of signalling pathways underlying human cognition and iii) the numerous data showing sex steroid regulation of these behaviours, exposure to BPA could also alter human cognitive abilities.

The main evidence is summarised in Table 19 and Table 20 below.

Table 19: Summary table of the ED-mediated MoA of BPA on learning and memory

Level	Molecular	Cellular	Organ/function
Alteration and available evidence	Alteration of the expression level of NMDAR, kinases, enzymes, involved in neurotransmitter. Regulation through estrogenic pathways	Modification of spine density or neurogenesis through estrogenic pathways	Alteration of learning/memory through estrogenic pathways
	<ul> <li>In vitro: Xu et al., 2010c</li> <li>In vivo: Xu et al., 2010b, 2011a; 2013b, 2014b, 2015b; Eilam-Stock et al., 2013; Jang et al., 2012; Matsuda et al., 2013; Wang et al., 2016; Zhang et al., 2014</li> </ul>	- In vitro: Xu et al., 2014a; Xu et al., 2010c - In vivo: Xu et al., 2015a and b; Inagaki et al., 2012; Leranth et al., 2008; Kumar et al., 2014; Kim et al., 2011	- <i>In vivo</i> : Xu <i>et al.,</i> 2010b; Xu <i>et al.,</i> 2011a; Inagaki <i>et al.,</i> 2012; Xu <i>et al.,</i> 2014b; Xu <i>et al.,</i> 2015b

Table 20: Overview of the elements supporting identification of alteration of learning/memory as an ED-mediated effect of BPA

Adverse effect	ED MoA	Human relevance
Alteration of learning and memory associated with cerebral modifications consisting of reduced expression levels of receptors (NMDAR), kinases, enzymes involved in neurotransmitter regulation, and synaptic proteins as well as decreased spine density or neurogenesis	Main evidence provided by inhibition of the functional and structural effects with ER antagonist or the suppression of estrogeninduced effects in ovariectomised animals.	Similar signaling pathways (e.g. NMDAR) in cognitive function in humans and rodents. Sex steroids known to modulate cognitive function in humans and rodents. Some indications for the effect (Elsworth et al. 2013 and 2015) or the estrogenic MoA (Leranth et al., 2008) in primate studies.

## 4.4.5 Summary tables of studies

The following table is presented by type of exposure (developmental, prepubertal and pubertal, adult), species studied (mice, rats, then other species) and year of publication.

Table 21: In vivo studies investigating the effect of BPA on learning and memory

Reference	Study design	Route of exposure Dose	Effects on learning and memory	Investigation of brain tissues and brain area relevant to the behaviour
Developmen	tal exposure (ges	tation and/or lactati	on)	
Mouse studies	5			
Xu et al., 2010b	ICR mice GD7-PND21	Oral (gavage) 0.05, 0.5, 5, 50 mg/kg/d	F1 male pups at PND 21/26 or 56/61  Morris water maze test:  longer distance in all BPA-exposed group to find the hidden platform in the 4 consecutive days (both ages)  average escape length extended by BPA at 5 or 50 mg/kg/d in PND21 mice and at 0.5, 5, or 50 mg/kg/d in PND 56 mice  yercentage of time spent in the target quadrant at 0.5 or 5 mg/kg/d (both ages)  Step down passive avoidance test:  Dose-dependent ⅓ in error frequency to step down from a platform after received footshock  Dose- dependent ⅓ in the latency of the step down 24 h after the training  Significant for both effects from 5 mg/kg in PND21 offspring and 50 mg/kg in PND56 offspring)  Global effect: impairment of spatial memory and avoidance ability	<ul> <li>Hippocampus</li> <li>Decrease of the expression of GluN1, GluN2A and GluN2B subunits of NMDA receptors.</li> <li>Decrease in the expression of estrogen receptors ERβ.</li> </ul>
Jasarevic et al., 2011	Outbred deer mice 2 weeks prior mating to end of lactation	Oral (diet) 7 mg/kg bw/d or EE	PND60 (Barnes maze), males and females  Control males exhibited shorter latency to exit the maze at day 2 than BPA, EE or females in all diet groups,  Control males and EE-exposed females committed fewer errors than the BPA-group  Failure to acquire spatial-oriented strategy in males exposed to BPA or EE.  Jatency to exit the maze and more rapid	

			acquisition of spatial oriented strategy in females typical of males with EE but not with BPA Global effect: altered spatial learning abilities in males, female abilities become more like those of control males	
Viberg <i>et</i> <i>al.</i> , 2011	NMRI mice (10- day old) Single exposure	Oral (gavage) 0.32, 3.2 or 4.8 mg/kg bw	3 month old males - no effect in the Elevated plus- maze. 4 month old males - no effect in the Morris water maze.	
Nakamura et al., 2012	ICR mice GD0 to PND21	SC 20 µg/kg/d	PNW 12/13, males and females no effect on spatial memory as tested in the Morris water maze.	
Jang <i>et al.</i> , 2012	C57BL6 mice GD 6 – GD 17	IP 0.1, 1 or 10 mg/kg bw/d	**B-week old F2 mice (females) - No effect on latency after repeated trials or after reverse trial with modification of platform location in the Morris water maze - Impaired memory retention: ン cross-over latency times at 1 and 10 mg/kg in the passive avoidance test indicating that memory retention of electric shock is impaired	### Hippocampus  8-week old F2 mice  UBrdU-positive cells in the hippocampal DG 10 mg/kg  UBrdU-positive cells in the hippocampal DG 10 m
Jasarevic et al., 2013	Outbred deer mice 2 weeks prior mating to end of lactation	Oral (diet) 0.05, 5 or 50 mg/kg of feed	Approx. PND60 (after sexual maturity), males and females  - Males at the two highest doses exhibit impairment in spatial learning assessed in the Barnes maze, similarly to EE.  - No effect in females in contrast to EE  Global effect: spatial memory impaired in males	
Matsuda et al., 2013	C57BL/6J mice GD 10-PND20	SC 250 ng/kg bw/d	PNW4 (males and females) - Enhanced fear memory in females using contextual fear conditioning PNW 9 (males and females) - No effect Global effect: enhancement of contextual memory of fear conditioning in females	<ul> <li>¬ serotonine metabolites in the brain (hippocampus, striatum, midbrain, pons, medulla oblongata of juvenile females (PNW4) but not adult females (PNW9) or juvenile or adult males</li> <li>¬ expression of Tph2, Slc6a4 and Maoa mRNA in the hippocampus of juvenile females.</li> </ul>
Kumar et al., 2014	Swiss albino mice GD7- PND21	Oral (gavage) 50 µg/kg bw/d	F1 pups at PNW 8 (only male tested)  - Effect on the escape latency, ↗ distance taken to find hidden platform on days of training, ↘ time spent in the target quadrant in the probe test after removal of the platform in the Morris water maze Global effect: impairment of spatial memory	Cerebral cortex and hippocampus F1 male pups at PNW 3 or 8  - Upregulation of Nrxn1 and Nlgn3 mRNA and protein level for both areas and ages  - 对 dendritic spine density for both areas and ages

Kubo <i>et al.</i> , 2001	Wistar rats GD0 - PND21	Oral (water) 1.5 mg/kg bw/day	F1 pups at PNW 7 (males and females) - lower avoidance memory in females and suppression of sexual dimorphism (step-through passive avoidance test)	F1 pups at PNW 7 - larger locus coeruleus in exposed females than in control males
Poimenova et al., 2010	Wistar rats GD1 - weaning (42 days)	Oral 40 µg/kg bw/day	F1 pups at PND 46 (males and females) - → spatial memory in both sexes (Y-maze test)	F1 pups at PND 46  -   → levels of corticosterone and \( \subseteq \text{levels of GR in males in basal state and in the 2 sexes after stress}  - No effects on the MR receptor in normal conditions, but \( \sum \text{MR level in females in the 2 groups of females} \)
Stump et al., 2010	Sprague Dawley rats GD 0 to PND 21	Oral (diet) 0.15, 1.5, 75, 750, and 2250 ppm (mean target doses of 0.01, 0.1, 5, 50, and 150 mg/kg/d)	PND 22 and 62 (males and females)  No effect on learning and memory as tested using the Biel water maze	PNDs 21 and 72  No effect on brain and nervous system neuropathology and brain morphometry.
Goncalves et al., 2010	Wistar rats  Gestation (20 d), lactation (21 d) or gestation + lactation (41 d)	Oral (gavage) 40 µg/kg bw/day	F1 pups at PNW 16 (males and females)  - Alteration of both short-term and long-term conditioning memory in the inhibitory avoidance task in males and females treated during gestation and lactation and males treated during lactation only  - Altered short-term and long-term memory by treatment for object recognition in both sexes  - In the Morris water maze: ↗ latency time during training in animals exposed during lactation and ↘ duration on the platform in the retention test in animals exposed during gestation and lactation.  Global effect: impairment of inhibitory avoidance, object recognition and spatial memory	
Xu et al., 2011a	Sprague Dawley rats (18-day old) Single exposure 1 or 24 hours before behavioural testing	SC 50 µg/kg bw or 500 µg/kg bw	Only males were tested:  - A latency in retention test 1 hour after footshock in the passive avoidance task when BPA was administered 1 hour but not 24 hour before testing in male rats.  - Similar effect with EB  - Co-treatment of BPA with EB did not affect latency  Global effect: impairment of the short-term avoidance memory	Hippocampus  No effect on levels of NMDA receptor GluN1 or GluN2B by BPA or EB    Plevel of phosphorylated-GluN1 and phosphorylated GluN2B 1 hour after exposure to BPA or EB.  Co-administration of BPA and EB inhibited the effect.  In mitogen-activated extracellular signal-regulated kinase (ERK) by BPA or EB.  Pretreatment with ER antagonist ICI 182,780 inhibited BPA or EB effect on phosphorylated GluN1 and GluN2B and ERK.  An ERK-activating kinase inhibitor U0126 reduced BPA-or EB-induced phosphorylation of GluN1, GluN2B and ERK within 1 h.
Jones <i>et al.</i> ,	Long Evans rats	Oral (drinking	F1 pups at PND 90 to 150 (males and females)	

2012	GD7 - PND14	water) 5, 50, 500 or 5000 µg/kg/d	No effect on spatial memory in the Morris water maze (The data are not shown, only the statistics)	
Kuwahara et al., 2013	Sprague Dawley rats GD10 - PND14	Oral 50 or 500 μg/kg/d	Only males were tested:  - ↗ in time to reach the reward in the MAZE test in male offspring of dams exposed to 50 μg/kg (female not tested)  - No significant effect in time to reach escape in the MWM test  Longer latency during training session for the stepthrough passive avoidance test at 50 μg/kg/d. No alteration in retention  Global effect: impairment of spatial learning and memory.	
Xu et al., 2014b	Sprague-Dawley rats Exposure: Last GD to PND21	Oral (drinking water) 0.1 mg/L (approx 2.5 mg/kg bw/d)	PNW9 (only males were tested) - Increase of latency (Morris water maze) - Decrease of the time spent in the platform quadrant.  The ER antagonist ICI 182,780 abolished the effects of BPA on rat behaviour and memory performance  Global effect: Impairment of spatial memory.	Hippocampus Expression of ER at PND7, 11 et 21  - □ of the expression of ERa at PND7 et PND21  - □ of the expression of ERa at PND11  - No difference between both hemispheres. The ER antagonist ICI 182,780 abolished the effects of BPA. The effects are observed at mRNA, protein (W Blot) and immuno-histochemistry levels  - No effect observed on ERβ Translocation of ERa towards nucleus (co-labelling of ERa and NeuN, neuronal nuclear antigen)  - Alteration of the translocation of ERa towards nucleus.  - □ of the expression of ERa in total proteins at PND7 and PND21, and □ at PND11.  - □ of ERa in the nuclear proteins at PND7 and 11 (not easily observable at PND21 because of the strong decrease in ERa in controls).  - □ ratio Nuclear ERa/total ERa, at PND7 and 11, suggesting a failure in the translocation of ERa towards the nucleus. The ER antagonist ICI 182,780 abolished the effects of BPA. Phosphorylation of ERa  - □ phosphorylation of total and nuclear ERa at PND7 and PND11.  - □ phosphorylation of total and nuclear ERa-Ser118 at PND7 and PND11.  The ER antagonist ICI 182,780 abolished the effects of BPA.
Wang <i>et al</i> ., 2014b	Sprague Dawley rats	Oral (gavage) 0.05, 0.5, 5 or 50	F1 pups at PND 21 (males)  - 7 in working memory errors at all BPA doses and in	Hippocampus F1 pups at PND 21

	GD9-GD20	mg/kg bw/d	reference memory errors at all doses except 0.5 mg/kg in the radial arm maze	<ul> <li>Synaptic junction dose-dependently altered by BPA: enlarged synaptic cleft and thinned PSD as observed by electron microscopy</li> <li>¬ mRNA and protein expressions of synaptophysin, PSD-95, spinophilin, GluR1 and NMDAR1 in male offspring</li> </ul>
Wang <i>et al.</i> , 2016	Sprague Dawley rats GD9-GD20	Oral (gavage) 0.05, 0.5, 5 or 50 mg/kg bw/d	F1 pups at PND 21 (males)  - ¬ in exploration of the familiar object 1.5h after training (significant at 50 mg/kg)  - ¬ in short-term (1.5h, significant from 0.5 mg/kg) and long-term (24h, significant from 5 mg/kg) recognition indexes (object recognition task)  Global effect: impairment of object recognition	Hippocampus F1 pups at PND 21  - Inhibition of expression of protein levels Akt , phospho-Akt, p44/42 MAPK and phospho-p44/42 MAPK protein levels  - Inhibition of phosphorylation levels of CREB and BDNF
Johnson et al., 2016	Sprague Dawley rats GD6 - PND21	Oral (gavage) 2.5, 25 or 2500 µg/kg bw/d	F1 pups at PND 90 to 104 (males and females)  Performance in the Barnes maze:  - animals in the 2500 μg/kg group sniffed more incorrect holes (7 <sup>th</sup> session)  - females in the 2500 μg/kg group had an overall longer latency as evidenced by reduced likelihood of locating the escape box  - in the 2.5 μg/kg group, males had a reduced latency and females a longer but not significant latency  - No significant effect on search strategy  Global effect: impairment of spatial navigational learning and memory.	
Hass <i>et al.</i> , 2016	Wistar rats GD7-GD21, then PND1- PND22	Oral (gavage) 0, 2.5, 25 µg/kg, 5 or 50 mg/kg bw/d	F1 at the age of 4-6 months (males and females)  Performance in Morris water maze:  - Shorter swim length and slower swim speed in females at 25 μg/kg making their behaviour resemble male performance  - No effect on latency to reach the platform  - Significant sex differences for the 3 endpoints (swim length, swim speed and latency) were observed in controls but not in animals exposed to 25 μg/kg and 5 mg/kg BPA.  Global effect: altered spatial learning pointing toward masculinisation of the female brain	
Non-human p	rimate study			
Elsworth et al., 2013	Rhesus macaques GD 100 - GD 155	Oral exposure to 400 µg/kg/d or subcutaneous exposure to dBPA	Females No behavioural or cognitive test performed	Oral exposure: - serum unconjugated dBPA levels of 0.68 ± 0.28 ng/ml ⅓ in tyrosine hydroxylase (TH) immunoreactivity in the substantia nigra and ventral tegmental area Subcutaneous exposure: - serum level of 0.91 ± 0.13 ng/ml of unconjugated dBPA

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	l or pubertal expo	sure		
Mouse studies			<u>,                                      </u>	<u></u>
Kim <i>et al</i> ., 2011	Male C57BL mice (6-week old) Exposure for 2 weeks	Oral (gavage in corn oil) 1, 5 or 20 mg/kg bw/d	Males Morris water maze: ¬ latency time after repeated training to reach the hidden platform at 20 mg/kg/d.	Hippocampus - 以 BrdU-positive cells in the hippocampal DG at 20 mg/kg - オ in neurogenesis at 1 mg/kg No effect on neuronal loss or astrocyte activation.
Xu <i>et al.</i> , 2011b	Male and females ICR mice (4-week old) Exposure for 8 weeks	Oral (gavage in sesame oil) 40 μg/kg/d or 400 μg/kg/d	Males and female Elevated plus maze test: - Both doses of BPA reduced the number of open arm entries and the time spent in open arms in males but increased them in females (dose effect) Morris water maze test: - ¬ average escape pathlength in males at 40 μg/kg; no effect in females - No significant effect in the probe trial with hidden platform removed Step-down passive avoidance test: - ¬ latency to step down 24h after footshock in males at 40 μg/kg. Global effect: alteration of spatial learning and memory in males tending to abolish sex differences. Effect was more marked at the lowest dose.	
Rat studies	_		<u></u>	
Jain <i>et al.</i> , 2011	Male Wistar rats (6 to 8-week old) Exposure for 28 days	Oral (gavage in propylene glycol) 2 or 20 µg/kg bw/d	Males Passive avoidance test - □ mean initial step-down latency (both doses).  mean retention latencies following training at both doses (reversed by co-administration with of antioxidant N-acetylcystein (NAC))  Morris water maze - □ mean acquisition latency and prolongation in retention latencies at both doses (reversed by NAC).  Global effect: BPA affected behaviour by stress oxidation that was reversed by NAC (antioxidant)	Brain tissues         - ¬ in malonaldehyde (marker of lipid peroxidation)         - dose-dependent ¬ in GSH         - Effects reversed by co-administration of NAC
Diaz Weinstein <i>et</i> <i>al.</i> , 2013	Male and female sprague Dawley rats (7-week old) Exposure for 6 to 12 days	SC 40 μg/kg/d	Males and females  - □ spatial memory performance in the object placement test. Ability to discriminate between the old and the new locations appears disrupted.  Global effect: spatial memory impaired	

	Observation			
	began D6 post-			
	injection			
Kuwahara et al., 2014	Sprague Dawley	Oral: 0.05, 1 or 10 mg/kg performed	Only males were tested - No difference in latencies to reach the reward in the	
ui., 2011	(5-week old)	under light	MAZE test	
	Single exposure	anesthaesia with	- No effect on fear-motivated memory performance in	
		halothane	the step through passive avoidance test	
		Microinjection into		
		dorsal hippocampus	Global effect: no impairment of spatial learning and memory.	
Bowman et	Males and	SC	PNW 11 (males and females)	Hippocampus
al., 2015	females	40 μg/kg bw/d	- elevated plus maze test: או number of total visit and visit in the closed arm in males but not females	PNW 13
	adolescent Sprague Dawley		- object placement trial: \(\mu\) in time spent exploring	י spine density of both apical and basal dendrites on pyramidal cells in CA1 (both sexes).
	rats (6-week		but no effect on percentage of time spent exploring	- No effect in the mPFC areas of the hippocampus
	old)		object in the new location	
	Exposure: one		- object recognition trial : \( \mu \) percentage of time spent	
	week		with the object in the new spatial location in males	
	Observation at		but not females leading to an accentuation of the	
	adulthood (>11 wk)		sex dimorphism. Global effect: impairment of non-spatial memory, object	
	WK)		recognition in males but not in females. No effect on	
			spatial memory, object placement	
Non-human p	rimate study	·	, .p	
Elsworth et	Male and	Subcutaneous	Males and females	- plasma level of 13.1 ±1.4 ng/ml at day 30 after
al., 2013	females St. Kitts	exposure to dBPA	- No effect on working memory as tested in the 2-	implantation
	African green	resulting in a	well spatial delayed response task. Animals were	- no effect on dopaminergic (DA) neurons (TH
	juvenile	plasma level	used indifferently of their sex.	immunoreactivity) in the ventral mesencephalon, or on
	monkeys (14- 18 months old )	measured to be 13.1 to 16.8 ng/ml		number of spine synapses in the hippocampus and prefrontal cortex (while BPA treatment during fetal
	Exposure for 30	across 3 measures		development induced a decrease in midbrain DA
	days	in time.		neurons, see above Elsworth <i>et al.</i> , 2013)
Adult expos	lire			
Mouse studies				
Xu et al.,	Male and female	Oral (gavage)	PNW22 (males, females)	Hippocampus
2013b	ICR mice	0.4, 4 or 40 mg/kg	Morris water maze	- א numeric synaptic density in males
	Exposure for 12	bw/d	- 7 average escape pathlength to the hidden platform	- Negative effect on the structural parameters of synaptic
	weeks		in males but not in females	interface including enlarged synaptic cleft, reduced
			Step-down passive avoidance task	length of active zone and PSD thickness in males
			י א step-down latency 24h after footshock in males but not in females	Down regulation of expression of synaptic proteins and synaptic NMDA receptor subunit GluN1 and AMPA
			but not in lemales	receptor subunit GluR1 in males
			Global effect: sex specific response of males, linear and	Table Sasanie State III maios

			nonlinear dose-response was observed according to the task	
Zhang et al., 2014	Male ICR mice (10-week old), Exposure for 90 days	Oral 0.4, 4 or 40 mg/kg bw/d	Only males were tested - 7 in freezing time 1h and 24h after fear conditioning training at 4 and 40 mg/kg Global effect: enhanced acquisition and retention of fear memory	Hippocampus  - □ level of NMDA receptor subunit GluN1 and □ expression of histone deacetylase 2 before fear conditioning training  - Enhancement by BPA of changes in expression of GluN1, phosphorylated extracellular regulated protein kinases and histone acetylation induced by fear conditioning
Xu et al., 2015b	Female ICR mice (7-week old) undergoing ovariectomy or sham operation Exposure for 8 weeks	SC 40 or 400 μg/kg/d	<ul> <li>Only females were tested         <ul> <li>No significant effect in the escape path length or in performance in the probe trial in the Morris water maze in sham female mice exposed to BPA</li> <li>No effect on step-down latency in the step-down passive avoidance task in sham mice exposed to BPA</li> </ul> </li> <li>Both doses of BPA shortened the escape path length and increased performance in the probe test in OVX females and thereby eliminate or decrease the difference between the vehicle sham and the OVX mice.</li> <li>Co-treatment of BPA with EB partially eliminated EB-induced shortening of the escape path length of OVX mice.</li> <li>Both doses of BPA decreased the step-down latency in OVX females and thereby decrease the difference between the vehicle sham and the OVX mice.</li> <li>Co-treatment of BPA with EB partially eliminated EB-induced increased in step-down latency of OVX mice.</li> <li>Global effect: BPA suppresses the reduction in spatial and passive avoidance memory observed in OVX mice and inhibits the rescue effect of estrogen</li> </ul>	Hippocampus  No effect of BPA in numeric synaptic density and in structure of the synaptic interface of sham females  BPA decreased synaptic interface modification in OVX females and thereby decrease the difference noted between the vehicle sham and the OVX mice.  Co-treatment of BPA with EB inhibited EB-induced modification of synaptic interface in OVX mice  No effect of BPA in expression in proteins synapsin I and PSD-95 and NMDA receptor GluN2B in sham females  These proteins were inhibited by BPA in OVX females.  Co-treatment of BPA with EB inhibited EB-induced enhancement of the expression of these proteins
Rat studies Eilam-Stock	Male Sprague	SC	Only males were tested	Hippocampus and prefrontal cortex
et al., 2011	Dawley rats (approx. 10- week old) Single dose on the day of the test	40 μg/kg	- Y memory performance in the object recognition test - Y performance in the object placement test Global effect: impairment of visual and spatial memory	□ Udendritic spine density on pyramidal cells in CA1 and medial prefrontal cortex     □ Ulevel of PSD-95, a synaptic marker, in hippocampus and っ cytosolic pCREB, a transcription factor, in medial prefrontal cortex.

Inagaki et al., 2012	Female Sprague Dawley rats (3- month old), ovariectomised or intact animals Single dose 30 min before or immediately after the sample trial of the test	SC Ovariectomised animals: 0, 0.4, 1, 4, 40 or 400 μg/kg/d BPA, alone or in combination with 17β or 17α-E <sub>2</sub> Intact animals: 0 or 40 μg/kg/d BPA, alone or in combination with 17β-E <sub>2</sub>	Ovariectomised animals  - BPA alone did not significantly affect object placement (OP) or object recognition (OR) memory performance  - BPA inhibited E₂-induced memory enhancing effects from 1 μg/kg for OP memory and 40 μg/kg for OR memory  Intact animals tested at the different phases of the estrous cycle  - No influence of BPA on OP memory performance at any phase of the cycle  - Inhibition of OR memory performance in females in proestrus (when E₂ levels are the highest)  Global effect: BPA blocked E2-enhanced recognition memory and impacted OR memory performance during proestrus	Ovariectomised animals  Pre-frontal cortex  - No effect of BPA on E2-induced increase in spine density of pyramidal neurons  Hippocampus BPA altered E2-induced increase in spine density of pyramidal neurons
Fan <i>et al.</i> , 2013	Male Wistar rats (60-days old) Exposure for 10 weeks pre- mating	Oral (diet, in corn oil) 50 µg/kg bw/d	FO males (female not exposed or tested)  - ↗ time to find hidden platform and ↘ platform contact accuracy when platform was removed in the Morris water maze  Global effect: acquisition and retention of spatial memory affected in exposed adults (see above for result of paternal exposure on offspring)	Hippocampus F0 males - 以 AChE activity - Mild but significant association between decreased hippocampal AChE and cognitive decline
Non-human p	rimate study			
Elsworth et al., 2015	Adult St. Kitts African vervet monkeys Exposure for 30 days	SC with osmotic minipump to achieve 50 µg/kg bw/d deuterium- labeled BPA	Only males were tested:  deficit in working memory accuracy in the two- choice spatial delayed response task after 1 week of exposure but not after 4 weeks.	- ע number of spine synapses in the PFC and CA1 regions after 4 weeks of exposure, - partial recovery following BPA withdrawal

Table 22: In vitro and ex vivo studies investigating the BPA MoA in neural brain cells

Reference	Tissue and treatment period	Type of modification	Evidence for ED/other MoA
Xu <i>et al.,</i> 2014a	Cultured hippocampal neurons from postnatal 24-hr old SD rats After 6-day culture in standard medium, 24-hr culture with 1, 10 or 100 nM BPA In culture with ICI 182,780 (ER antagonist), U0126 (MEK antagonist) or MK-801 (non- competitive NMDA receptor antagonist), cultures were pretreated with BPA for 30 min. 17 b-E2 used as positive control	Morphological examination: dose dependent ⊅ total length of dendrite (ss from 10 nM), motility (ss from 10 nM) and density of dendritic filopodia (ss at 100 nM. Similar effect with 17β-E2.  ICI 182,780 completely eliminated the effect of BPA (or 17β-E2)  U0126 or MK-801 partly eliminated the effect of BPA (or 17β-E2)  ¬ in amount of F-actin (cytoskeleton critical in morphological changes during synaptic plasticity) present in dendrites filopodia from 10 nM that is inhibited by co-treatment with ICI 182,780, U0126 or MK-801 (similar with 17b-E2)  Expression of proteins of the Rho family (involved in intracellular actin regulation):  ¬ a expression of Rac1/Cdc42 from 10 nM, blocked by co-treatment with ICI 182,780 or U0126  ¬ u expression of Rhoa from 10 nM, partly blocked by co-treatment with ICI 182,780 or U0126 (similar with 17β-E2)	BPA may affect dendritic development through modification of expression of Rho proteins and enhancement of Factin cytoskeleton and involve ER- and ERK1/2 signaling pathways
Xu et al., 2010c	Cultured hippocampal neurons from postnatal 24-hr old SD rats After 7-day culture in standard medium, 30-min exposure to 1, 10, 100 or 1000 nM BPA In some culture pre- treatment with ICI 182,780 for 30 min. 17 b-E2 used as positive control	Morphological examination: - dose dependent ⊅ filopodia motility and density (ss from 10 nM) ICI 182,780 suppressed the effects of BPA On filopodia motility: - similar effect with 17β-E2 - BPA partly suppressed (ss) the effect of 17β-E2 Expression of NMDA receptors: - no effect of BPA after 30 min incubation on NMDA GluN1 and GluN2B expression - Similar absence of effect with 17β-E2 and co- treatment BPA+17β-E2. Phosphorylation of NMDA GluN2B (on Ser 1003): - ¬ level of pGluN2B by BPA alone or 17β-E2 alone - suppression of effect by co-treatment BPA+17β-E2 -suppression of effect by ICI 182,780 (ER antagonist) Levels of ERβ: - no effect of BPA after 30 min incubation - Similar absence of effect with 17β-E2 and co- treatment BPA+17β-E2.	BPA may rapidly affect dendritic morphology through ER- mediated pathways involving activation (phosphorylation) of NMDA receptor GluN2B (primarily expressed in immature synapses of the hippocampus during postnatal development).
Hasegawa et al., 2013	Cultured hippocampal slices from adult (12-week old) male Wistar rats 10 or 100 nM BPA perfused 30 min before measurements	Modulation of synaptic transmission through long-term depression (LTD) with custom multi-electrode probes in various hippocampus areas: In the CA1 area: - dose −response Ϡ in LTD from 10 nM - effect suppressed by 4-OH-tamoxifen (ERRγ antagonist) - no effect of co-administration of ICI 182,780 (ER antagonist) In the dentate gyrus - ⊔ LTD at 10 nM, no effect at 100 nM	BPA may affect synaptic transmission in adult through ERRγ- but not ER-dependent pathways

Table 23: In vivo studies reporting effects of BPA exposure on brain tissues

Reference	Study design	Route of exposure Dose	Investigation of brain tissues and potential indications of endocrine MoA
Developmen	ntal exposure		
Kundakovic et al., 2013	BALB/c mice (males and females) Exposure: GD0-GD19	Oral 2, 20, 200 µg/kg/d	Prefrontal cortex: Changes in Esr1 (ERα gene), Esr2 (ERβ, sexspecific), Esrry (sex-specific) Hippocampus: Changes in Esr2 (sex-specific) and Esrry
Kumamoto & Oshio, 2013	ICR mice Exposures twice at GD6- GD15	Oral 0.0.2 and 50 mg/kg BPA	Cerebrum (cerebral cortex and subcortical structures (hippocampus, basal ganglia, and olfactory bulb)  - Xist downregulated at PN21, 28 in BPA-50 and at PN28 in BPA-0,02. Tsix upregulated.  - Changes in the expression of X-linked genes.  - *Increased AR levels at PN2 in BPA-50, and lower at PN28 in BPA-50 and 0,02  - *Increased ERα levels at PN4 and 28 in BPA-50 and 0,02, and ERb at PN28 (BPA-50-General:  E2 levels lower at PN21 in BPA-0.02 and BPA-50 Anogenital distance shortened at PN4-21-28 in BPA-50; at PN21 in BPA-0.02
Xu <i>et al.</i> , 2010a	SD Rat Exposure: GD7-PND21	Oral intragastric 0.05, 0.5, 5, 50, 200 mg/kg/d	<ul> <li>low doses [0.05-50] strongly decrease the expression of GluN1, GluN2A and GluN2B subunits of NMDA receptors, but at the higher dose (200 mg) the expression of GluN1 only was strongly inhibited</li> <li>dose-dependent decrease in the expression of estrogen receptors ERβ.</li> <li>dose-dependent increase of aromatase cytochrome P450 (P450arom).</li> </ul>
Castro <i>et al.</i> , 2015	Wistar rat - Females Exposure: GD12-PND0 and PN1-PN21	SC 10 µg/kg/d BPA	Brain prefrontal cortex  - Important decrease of the expression of the gene and the protein of 5a-R2 and 5a-R3
Adult expos	ure	I.	
Xu <i>et al.</i> , 2015a	Adult mice (10- week old) Exposure: 12 Weeks	Oral intragastric 0.04, 0.4, 4, 40 mg/kg/d	<ul> <li>Hippocampus         <ul> <li>Decrease in the expression of ERβ protein in males (inverse U-curve dose response) but no effect in females.</li> <li>BPA up-regulated GABA(A)a2 receptor in females but down-regulated in males</li> <li>Decrease in the expression of ERβ estrogen receptors</li> </ul> </li> </ul>
Castro at	Wictor est	50	The effect on ERβ in males is associated with:  - An increase of anxiety  - A decrease of brain level of testosterone in males.  No effect on brain and serum estradiol in females  Note: [0.4-40mg] doses decrease  Testosterone in the brain
Castro <i>et</i> <i>al.</i> , 2013	Wistar rat Exposure: Adulthood (4 days)	SC 50 µg/kg/d BPA	Brain prefrontal cortex   Reduced mRNAs and protein levels of 5α-reductase 1 in females but not males   Increased mRNAs and protein levels of

			aromatase in males and females  No effect on tryptophane hydroxylase Tph1. Increase in the expression of genes coding for Tph1 and Tph2 in males and females. Increase in the expression of the protein Tph2 in males and females
Leranth et al., 2008	Adult female African green monkeys ovariectomised Exposure: 28 days	SC implants: vehicle, estradiol alone, BPA (50 µg/kg/d), or estradiol + BPA	- BPA alone had no effect - Estradiol induced a synaptogenic effect, in hippocampal regions (CA1, CA3 and DG) and prefrontal cortex, which is abolished by the continuous exposure to BPA

### 4.5 Metabolism and obesity

# 4.5.1 Overview of previous evaluation of BPA's effect on metabolism and obesity

Effects of BPA on metabolism and obesity have already been assessed in previous EU reports (ANSES, 2014; EFSA, 2015; ECHA, 2015). It is not the aim of this dossier to detail the studies that were considered in these previous reports and only the recent studies or studies that could be used to investigate the MoA will be further considered.

**The BPA restriction dossier (ANSES, 2014)** concluded for hazard assessment in animals that:

"Studies examining effects on enzyme activity, growth and metabolism suggest that rodents exposed in adulthood or during gestation undergo metabolic changes in various organs such as the liver, adipose tissue and pancreas. Moreover, a few authors have noted changes in the expression of protein-coding genes intervening in the cell signalling pathways involved in lipogenesis and carbohydrate metabolism. There is a trend showing in vivo effects on lipogenesis. In vitro mechanistic studies support these observations.

However, the effects on carbohydrate metabolism cannot be confirmed on account of insufficient repeatability.

Thus, in animals, BPA increases blood lipid levels, leads to excess body weight and enhances lipogenesis. The effects on lipogenesis (in vivo and in vitro data), after pre- or perinatal exposure or exposure in adulthood, are considered to be recognised. The effects on glucose metabolism after pre- or perinatal exposure to BPA are considered to be controversial.

Changes in lipid metabolism are effects that are taken into account for the risk assessment."

These studies are summarised in Table 24.

Table 24: Summary of the studies examining the effects of bisphenol A on metabolism as quoted in the BPA restriction Dossier (ANSES 2014)

Reference	Species/ strain	Routes of exposure	Dose Exposur e period	Effects NOAEL/LOAEL	
Alonso- Magdalena et al., 2010	Mice	Sub- cutaneous	0 - 10 and 100 μg/k g bw/day GD9 to GD16	In F1 offspring, 6-month males had \( \sigma\) glucose tolerance, \( \sigma\) insulin resistance, and \( \sigma\) plasma levels of insulin, leptin, triglycerides and glycerol, altered calcium signalling in islets of Langherans \( \sigma\) BrdU incorporation into insulin-producing \( \beta\) cells, whereas their surface was unchanged.  In mothers, \( \sigma\) insulin resistance induced by gestation and \( \sigma\) glucose tolerance.  Dose-dependent \( \sigma\) in plasma levels of insulin, leptin, triglycerides and glycerol.  \( \sigma\) insulin-stimulated Akt phosphorylation in gastrocnemius skeletal muscle and liver.  4 months post-partum: higher BW, higher concentrations of insulin, leptin, triglycerides and glycerol	
Ryan <i>et al.,</i> 2010	CD-1 mice	Oral	0.25 μg/kg bw/day GD0 to PND21	In F1 offspring, ≯ BW in males and females at 3 weeks ≯ body length in males at 4 weeks, these biometric differences disappearing in adulthood. No significant effects on glucose tolerance were observed.	
Somm et	Sprague	Oral	70 μg/kg	At birth: BPA treatment during gestation did not	

al., 2009	Dawley rats		bw/day GD6 - PND21	affect sex-ratio or litter size. Newborns (♀ and ♂):  ¬ weight  PND21  ¬ BW in females  Increased parametrial fat associated with adipocyte hypertrophy and overexpression of lipogenic genes and lipogenic enzymes  In the liver, increased RNA levels of C/EBP-a,  SREBP-1C, ACC and FAS k. Circulating lipids and glucose were normal.  4 to 14 weeks: no difference in BW observed between BPA-treated males and control animals on standard chow diet.  ¬ BW in BPA-exposed males fed a high-fat diet.  ¬ BW in females for the 2 tested diets. In males fed a high-fat diet, normal glucose tolerance test results.  Conclusion: Perinatal exposure to BPA.  ¬ Adipogenesis at weaning in ♀. In adult ♂, ¬ BW
Miyawaki et al., 2007	ICR mice fed a high-fat diet	Oral	0.26 and 2.72 mg/kg bw/d via drinking water GD10 until weaning	observed if high-fat diet.  PND31:  BW in BPA-exposed (low and high dose groups) females fed a high-fat diet.  adipose tissue weight in BPA-exposed (low dose group) females fed a high-fat diet.  BW and adipose tissue weight in BPA-exposed (high dose group) males fed a high-fat diet.  leptin in BPA-exposed (low dose group) females fed a high-fat diet  total cholesterol in BPA-exposed (low and high dose groups) females fed a high-fat diet.  No change in glycemia in females  non esterified fatty acids in BPA-exposed (low dose group) males fed a high-fat diet  triglycerides in BPA-exposed (low and high dose groups) males fed a high-fat diet.  glycemia in BPA-exposed (low dose group) males fed a high-fat diet.

**The EFSA opinion on the risks to public health related to the presence of BPA (2015) concluded**: "Of the reviewed human studies on metabolic effects only two were prospective while 22 were cross-sectional and thus not suitable on their own to study exposure-disease associations. Inconsistent with the results of cross-sectional studies one prospective study found that a higher BPA concentration in maternal urine during pregnancy was associated with a lower level of obesity in daughters. A causal link between BPA exposure and metabolic effects in humans cannot be established.

A number of studies in pre- and postnatally exposed rats and mice indicate that BPA exposure could have an effect on metabolic function as evidenced by effects on glucose or insulin regulation or lipogenesis, and body weight gain (short-term studies). Based on the results from other studies with a longer duration (e.g. 90 days) there is no convincing evidence that BPA is obesogenic after intrauterine exposure or in longer-term studies. Using a WoE approach, the CEF Panel assigned a likelihood level of "as likely as not" to metabolic effects of BPA. Since the likelihood level for this endpoint is less than "likely", this endpoint was not taken forward for assessing the toxicological reference point, but was taken into account in the

evaluation of uncertainty for hazard characterisation and risk characterization." See sections 3.7 and 4.3 of the EFSA (2015) opinion for more details.

In its opinion of **June 2015 (ECHA, 2015),** RAC adopted the following conclusions in relation to the analysis of the effect of BPA on metabolism and obesity:

"The Dossier Submitter derived a LOAEL of 0.26 mg/kg bw/day based on increased body weight and increased cholesterolemia in female mice in Miyawaki et al. (2007). The EFSA (2015) opinion concluded: see above

#### Conclusion

RAC in principle agrees with EFSA's conclusion on metabolism and obesity. Although RAC is of the opinion that the studies described are not sufficiently convincing for quantifying the doseresponse, RAC considers it prudent to take the metabolic effects into account in hazard and risk assessment (by accounting for them in the setting of Assessment Factors) and in health impact assessment."

Considering that the metabolic effects of BPA were taken into account in hazard and risk assessment at EU level under the restriction regulatory processes, it was decided not to further present and discuss the whole database in the main part of the present dossier.

Considering these previous assessments, metabolic effects of BPA could be evidenced by effects on glycaemia and its insulin regulation on the one hand or on lipogenesis and body weight gain on the other hand.

The pancreas is involved in the regulation of glucose homeostasis via two major hormones: insulin and glucagon, which are produced and released by the  $\beta$ - and the  $\alpha$ -cells of the pancreatic islets, respectively (Quesada et al., 2008; Rorsman and Braun, 2013). The pancreatic endocrine system also includes  $\delta$  cells secreting somatostatin (involved in the regulation of  $\alpha$ - and  $\beta$ -cell activities),  $\gamma$  cells, secreting pancreatic polypeptide involved in the regulation of both endocrine and exocrine pancreas secretions, and  $\epsilon$ -cells producing ghrelin, a protein that stimulates hunger (see **Figure 17**).

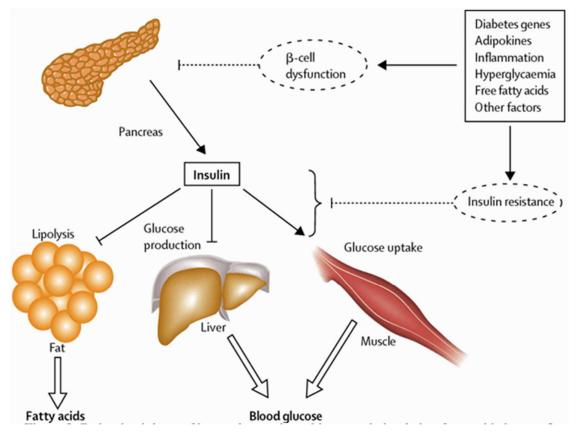


Figure 17: Pathophysiology of hyperglycemia and increased circulating fatty acids in type 2 diabetes from Stumvoll et al., 2005.

Adiponectin and leptin are protein hormones secreted from adipose tissue. They modulate metabolic processes and regulate energy balance. Leptin (secreted by adipocytes in proportion to fat mass) acts as a satiety hormone, whereas adiponectin (its expression by adipocytes is reduced in proportion to fat mass) is involved in glucose regulation and fatty acid oxidation in the liver, improves pancreatic  $\beta$  cell function, enhances peripheral insulin sensitivity, suppresses hepatic glucose production and reduces inflammation (Chakraborti, 2015). Both hormones play a major role in glucose and lipid metabolism, insulin resistance and obesity.

When insulin action decreases (e.g. with obesity), glycaemia remains at values exceeding physiological range between meals, resulting in an increase in insulin secretion by the pancreas to lower glycaemia. Normal glucose tolerance occurs as long as  $\beta$ -cells counteract hyperglycaemia through enhanced insulin secretion. First signs of insulin resistance arise in the liver, muscle and adipose tissues. In that case, hepatic production of glucose namely gluconeogenesis is no longer controlled appropriately by insulin, resulting in higher levels of glucose in the blood. Concomitantly, the glucose uptake in muscle, which is dependent on the insulin-regulated recruitment of the glucose transporter GLUT4 to the membrane, will be less effective. In the adipose tissue, lipolysis, usually inhibited by insulin, will be enhanced leading to elevated levels of free fatty acids in circulation (lipotoxicity).

A decrease in insulin secretion and/or in insulin action (insulin resistance) leads to an increase of lipolysis (and consequently an increase in plasma concentration of free fatty acids), an increase in hepatic production of glucose and a decrease in muscle glucose uptake (and consequently an increase in blood glucose concentration). These processes are auto-amplified since augmentation in free fatty acids in plasma and hyperglycemia increase insulin resistance and decrease insulin production in response to glucose.

In addition, adiponectin that is inversely secreted by adipocytes with enhanced fat mass will be less efficiently synthesised. Importantly adiponectin highly sensibilises metabolic tissues to insulin action. It reduces inflammation in the adipose tissue, suppresses glucose production and induces fatty acid oxidation in the liver and improves pancreatic beta cell function. Therefore, gradually, the body's ineffective use of insulin evolves as a tryptic of hyperglycemia, hyperinsulinemia and hypertriglyceridemia, in a vicious circle where hyperglycemia aggravates hyperinsulinemia and hyperinsulinemia aggravates hyperglycemia and hypertriglyceridemia. Eventually, type 2 diabetes develops with impaired  $\beta$ -cell function leading to persistent and progressive deterioration of glucose tolerance.

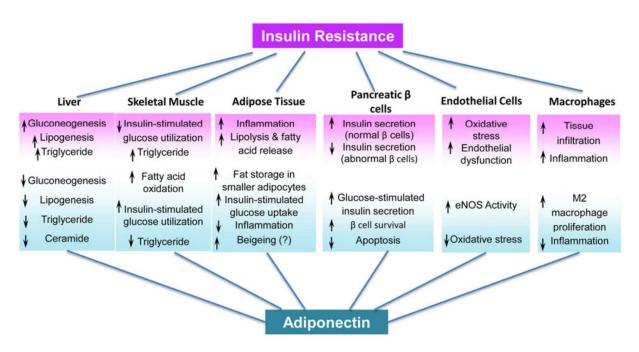
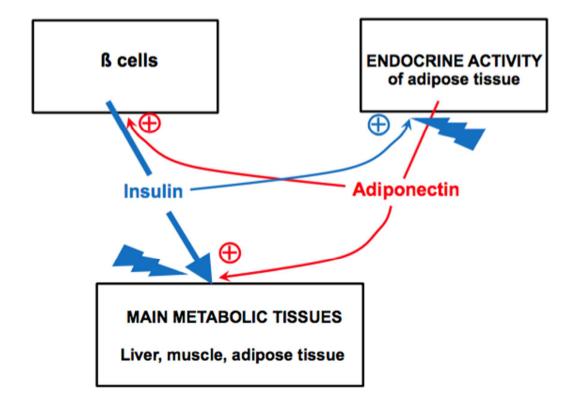


Figure 18: Summary of tissue-specific functions of adiponectin (Ruan and Dong, 2016).

Adiponectin and its downstream effector molecules are central for insulin sensitivity and homeostasis. Thus, dysregulation of adiponectin signaling will induce insulin resistance as follows. On the one hand, Adiponectin (shown in blue) acting via the adiponectin receptors (either I or II) expressed in liver, skeletal muscle, adipose tissue, pancreatic  $\beta$ -cells, endothelial cells or macrophages reduces metabolic effects favoring insulin resistance (e.g., inflammation in adipose tissue or in macrophages; lipogenesis and gluconeogenesis in liver) or enhances metabolic effects favoring insulin sensitivity (e.g.,  $\beta$  cell survival in pancreas, insulin-stimulated glucose utilisation in the skeletal muscle or uptake in the adipose tissue). On the other hand, insulin resistance (shown in purple) is favored in liver by enhanced gluconeogenesis or by enhanced inflammation in the adipose tissue to name a few of the effects (quoted from Ruan and Dong, 2016).



A self-amplifying loop of an initial increase in insulin resistance in type II diabetic processs.

Insulin acts on the endocrine activity of the adipose tissue to stimulate adiponectin secretion.

Adiponectin increases the efficiency of insulin on its target tissues (i.e. main metabolic tissues) and stimulates ß cells function.

Thus, an initial decrease in insulin efficiency (i.e. an increase in resistance to insulin) ( ) will provoke a reduction in the adiponectin secretion which will induce a supplemental increase in insulin resistance and a decrease of the ability of ß cells to secrete insulin in response to glucose. These last two changes will result in a further development of the type II diabetes.

Figure 19: Interaction between insulin and adiponectin.

In addition to the *stricto sensu* metabolic hormones (e.g., insulin, ghrelin, leptin, adiponectin), a large amount of hormones contribute to regulate energy homeostasis, among which are the sexual hormones, but also the glucocorticoid hormones or the thyroid hormones. Specifically, estradiol is involved in the regulation of metabolism through the modulation of food intake, body weight, glucose/insulin balance, body fat distribution, lipogenesis and lipolysis, and energy consumption. Moreover, estradiol exhibits other several important effects in the pancreas, including promoting insulin secretion and synthesis, enhancing  $\beta$ -cell survival, increasing islet oxygenation during transplantation and preventing lipotoxicity (Mauvais-Jarvis *et al.*, 2013). Estradiol-17 $\beta$  exerts a major role as evidenced with the phenotype of obesity and insulin resistance in both males and female mice deficient for the ERa (estrogen receptor alpha).

Lastly, it should be noticed that fetal pancreatic development occurs both in fetal and neonatal stages prenatally and post-natally in rodents whereas in humans, the major development is completed prenatally.

Overall, alterations in insulin secretion by  $\beta$ -pancreatic cells, or alterations of insulin action (signaling mechanisms) upon the insulin-sensitive organs, such as those leading to variations

in the expression levels of hepatic or adipose tissue markers known to reflect a state of insulin resistance, are considered by the experts as hallmarks of endocrine disruption mechanisms. This is especially true if there is a combination of effects, each leading to insulin resistance within the different insulin-sensitive tissues. Therefore, this dossier is based on a new evaluation of these specific aspects examining both *in vivo* and *in vitro* experimental studies.

In the following sections, available evidence of both types of effects (ß-cell dysfunction and insulin resistance), showing that BPA may have adverse effects and that these effects are related to an ED MoA in a causal way, will be considered.

Although the summary of the effects of BPA on metabolism and obesity presented above is mainly based on the literature collected for the restriction dossier, recent studies investigating "BPA and adipocytes" or "BPA and 3T3-L1 cells" or "BPA and obesity" or "BPA and steatosis" or "BPA and liver" or "BPA and pancreas" have been collected in Pubmed until May 2016.

### 4.5.2 Effects of BPA on glycaemia and insulin synthesis

### 4.5.2.1 Adverse effect of BPA on the endocrine function of the pancreas

This section focuses on the effect of BPA on the pancreatic insulin synthesis and secretion. However, some papers present the effect of BPA both on the pancreas and on the insulinsensitive tissues. In order to avoid redundancy, they will be presented in the present section (and summarised in Table 25):

Alonso-Magdalena et al. (2010) studied the effects of BPA on glucose metabolism in female mice, during gestation, and their male F1 offspring. BPA was administered sub-cutaneously to the mothers, from GD9 to GD16, at doses of 0, 10 and 100  $\mu$ g/kg bw/day. In the F1 offspring, 6-month old males had reduced glucose tolerance, increased insulin resistance, and higher plasma levels of insulin, leptin, triglycerides and glycerol. Moreover, the islets of Langerhans presented altered calcium signaling. The authors noted that bromodeoxyuridine (BrdU, a synthetic nucleoside commonly used in the detection of proliferating cells in living tissues) incorporation into insulin-producing  $\beta$  cells was reduced, yet their surface was unchanged. However, the latter results, although very likely, should be considered with caution, since they were obtained with cultured cells from exposed individuals. Therefore, taking into account isolation and culturing methods, cultured cells have different phenotypes than *in situ* cells. Such an approach is relevant when undertaking an instant analysis of the cellular state after rapid fixation and treatment of the tissues. However, it is not appropriate when examining differences in cell functioning between controls and individuals exposed to a stress agent.

**Ryan et al.** (2010) tested the hypothesis that prenatal and lactational exposure to BPA, at a dose consistent with environmental exposure (0.25  $\mu$ g BPA/kg bw/day), results in increased susceptibility to high-fat diet-induced obesity and glucose intolerance in CD-1 mice. F1 individuals were exposed to BPA (1  $\mu$ g/kg via the mothers' feed, equivalent to around 0.25  $\mu$ g/kg bw/day) from the embryonic stage GD0 to weaning (PND21). In the weaned F1 individuals, increased body weight was observed in males and females at 3 weeks and increased body length was observed in males at 4 weeks, these biometric differences disappearing in adulthood. No significant effects on glucose tolerance were observed. The authors concluded that the increased body length and weight were due to a faster rate of growth in the exposed mice rather than a state of obesity.

New experimental *in vivo* studies indicate that BPA exerts effects on the endocrine function of the pancreas (secretion of insulin). The potential impact of BPA on lipogenesis is confirmed by recent experimental (*in vivo* and *in vitro*) studies. These new data reinforce the observations

previously reported by ANSES. Therefore, the evaluation of the potential endocrine disrupting effects of BPA was conducted in order to determine to which extent BPA could interfere in the balanced interplay between insulin secretion and insulin action that controls glycaemia.

β-cell mass is critical for proper functioning of the endocrine pancreas as well as insulin biosynthesis and secretion. Therefore, BPA as an estrogeno-mimetic and, depending on the exposure periods, is likely to interfere with pancreas development and function.

Searching for new publications from 2013 to November, 2016 with the search terms BPA and PANCREAS generated 13 original articles, 3 *in vitro/ex-vivo* studies and 10 *in vivo* studies mostly on the mouse model. These papers are presented successively and summarised in Table 26.

### In vivo data and early life exposure

**Liu** *et al.* **(2013)** treated pregnant C57bl6 mice with either vehicle or BPA (100 μg/kg bw/day) on days 1–6 of pregnancy (P1–P6, preimplantation days); from day 6 of pregnancy (GD6) until postnatal day (PND) 0 (GD6–PND0, fetal exposure); from lactation until weaning (PND0–PND21, neonatal exposure); and from day 6 of gestation until weaning (GD6–PND21, fetal and neonatal exposure) via daily subcutaneous injection. Glucose homeostasis was impaired in males more than in females as assessed via handling glucose tolerance metabolic tests. After exposure to BPA, the β-cell mass increased while insulin secretion was either reduced or remained invariable. It suggested that β-cells were less functional in BPA-exposed mice. The alterations of insulin secretion rather than β-cell mass, were consistent with the development of glucose intolerance. Data also indicated that the fetal development stage may be a critical window of susceptibility to BPA exposure, knowing that fetal pancreatic development occurs both in fetal and neonatal stages prenatally and post-natally in rodents.

García-Arevalo et al. (2014) treated pregnant OF-1 mice with a subcutaneous injection of 10 µg/kg bw/day of BPA or a vehicle from day 9 to 16 of pregnancy. At weaning, males were either fed a normal chow diet (CD) or a high fat diet (HFD) resulting in 4 groups of 8 mice/group. Body weight was recorded weekly and male mice were sacrificed by 17 or 28 weeks. BPA-treated males had enhanced body weight whether fed a control or a HFD together with fasting hyperglycaemia, glucose intolerance and high levels of non-esterified fatty acids (NEFA) in plasma. Glucose-induced insulin secretion from isolated pancreatic islets was disrupted, particularly in the HFD-BPA group. The authors concluded that male offspring from BPA-treated mothers presented a form of diabetes which typically develops in later life and is associated with obesity.

**Garcia-Arévalo** *et al.* **(2016)** demonstrated that exposure of pregnant OF-1 mice to either vehicle (control) or BPA (10 and 100 μg/kg·d, BPA10 and BPA100) between GD9 and GD16 (subcutaneous injection) to cover the embryonic phase of pancreas development resulted in alteration of insulin secretion in the BPA10 male offspring with no change in the BPA100-exposed males at post-natal day 30. This was associated with an increase in pancreatic β-cell mass at PND0, PND21, and PND30 together with increased β-cell proliferation and decreased apoptosis. Transcriptomic analysis confirmed the differential expression of genes related to cell cycle and apoptosis. Importantly, treatment of pregnant mice with E2 (10μg/kg bw/d) between GD9 and GD16 also resulted in enhanced β-cell mass in the male offspring (PND30) as compared with controls, although it resulted from decreased apoptosis and not from changes in cell proliferation. Thus, it is proposed that modifications of the β-cell mass in the offspring as a consequence of estrogen signaling mechanisms initiated in fetal life and leading to an excess of insulin signaling during early life may contribute to impaired glucose tolerance during adulthood.

**Whitehead** *et al.* **(2016)** fed pregnant C57BL/6 mice with a BPA diet (25 mg BPA/kg bw/day diet roughly corresponding to 5 mg/kg bw/day) from embryonic day 7.5 (E7.5) to E18.5. At E18.5, fetal pancreas were collected and analysed for morphological changes in the endocrine

pancreas such as islet size, number and  $\beta$  and  $\alpha$  cell distribution. It was observed that BPA altered the differentiation program of pancreatic cells resulting in enhancement of the glucagon expressing cells and thus in a decrease of the insulin  $\beta$ - pancreatic cells. There was also a change in the localisation of the  $\alpha$ -cells, normally located in the periphery, as they were spread throughout the entire islet after BPA treatment. These data indicate that BPA may alter the differentiation program of the pancreas.

#### In vivo data on adult animals

**Alonso-Magdalena** *et al.* **(2015)** studied in OF-1 mice the metabolic status of mothers treated with BPA (10 and 100  $\mu$ g/kg bw/d) from day 9 to day 16 of gestation *via* subcutaneous route. Several months after delivery, it was shown that these female mice exhibited profound glucose intolerance and altered insulin sensitivity as well as increased body weight. Importantly, no effect was observed with non-pregnant mice. Mechanisms of action include reduced pancreatic  $\beta$ -cell mass as a consequence of decreased proliferation and increased apoptosis. Taken together, these data suggest that BPA exposure during gestation has long-term implications in glucose metabolism for the mother.

In the study from **Moon** *et al.* **(2015)**, 4 to 6-week-old C57BL/6 male mice on a high-fat diet (HFD) were treated with  $50\mu g/kg$  bw/day of BPA orally by gavage for 12 weeks. Although, body weight, percentage of white adipose tissue, and percentage of body fat did not differ between the treated and the control group of mice, long-term oral exposure to BPA along with an HFD for 12 weeks induced glucose intolerance in growing male mice. The origin of glucose intolerance did not result from any detrimental changes in the islet area or morphology or the insulin content of  $\beta$  cells. However, the authors observed decreased phosphorylation of AKT and GSK3beta in skeletal muscle indicative of insulin resistance which might be one mechanism by which BPA induces glucose intolerance. The effects are mostly subtle with the exception of the effects describing impairment of insulin signaling in the skeletal muscle.

**Moghaddam** *et al.* **(2015)** investigated the effects in adult male mice (strain not given) of BPA (0.5 and 2 mg/kg bw/day) dissolved in olive oil and injected intraperitoneally (n=6/group) for 4 weeks. BPA was found to enhance body weight with no differences between the 2 BPA-treated groups. BPA was also found to enhance glycaemia and plasma levels of triglycerides, LDL-C and cholesterol while HDL-C levels were significantly reduced. Importantly, in regard to the topic of this section, the authors evaluated the impact of BPA on the pancreas. Specifically, they studied oxidative stress. BPA injection increased malondialdehyde level and reduced the levels of glutathione (GSH) and the activities of SOD and CAT in the pancreas of the exposed compared to the control group. All these effects were significant and stronger in mice treated with the 2 mg/kg bw/day dose than in mice treated with the lower BPA dose. Taken together, it is suggested that BPA-induced hyperglycemia and hyperlipidemia may be associated with oxidative stress.

**Jayashree** *et al.* **(2013)** reported the effects of a single dose of BPA (20 or 200 mg/kg bw) diluted in corn oil and administered by gavage to male adult Wistar rats (n=6/group). Animals were sacrificed 30 days post dosing. The authors demonstrated enhanced serum insulin with the BPA dose but no change in fasting blood glucose level, indicative of an adaptive response of the pancreas to maintain glycaemia level. Glucose oxidation and glycogen content were found to be decreased in the liver of both high and low dose treated rats. In addition, there was impaired insulin response in the liver with decreased Akt phosphorylation, all indicative of hepatic insulin resistance. The authors also reported decreased testosterone levels which may also contribute to insulin resistance. However, unfortunately, there is no indication of the body weight of the animals and the doses used are high.

# 4.5.2.2 Mechanism of Action: secretion of insulin, insulin biosynthesis and secretion, and $\beta$ -cell survival

Estrogens signaling occurs via at least the a- and  $\beta$ -estrogen receptors and via GPR30 or GPER1, a membranous form of estrogen receptor, resulting in distinct effects depending on the receptor activated. Activation of ERa enhances glucose-stimulated insulin biosynthesis, promotes  $\beta$ -survival from apoptotic stimuli and prevents lipotoxicity. Activation of ER $\beta$  enhances glucose-stimulated insulin secretion (GSIS). Activation of GPER1 protects from apoptosis and enhances GSIS without affecting its biosynthesis (Tiano and Mauvais-Jarvis, 2012). For example, using  $\beta$ -cells and islets of Langerhans recovered from wild type (WT) and ER $\beta$ -/- mice, it was shown that ER $\beta$  was involved in the BPA-mediated rapid regulation of KATP channel activity, potentiation of glucose induced-[Ca2+]i signals and insulin release (Soriano et al., 2012).

Primary cultures of pancreatic islets recovered from C57BL/6 male mice were prepared by pancreas bile duct perfusion and collagenase P digestion (**Carchia** *et al.*, **2015**). The authors describe mitochondrial dysfunction and alteration of cell viability in pancreatic islets exposed to low dose of BPA ( $1 \times 10^{-9}$  M) through mechanisms involving oxidative stress and enhanced cell apoptosis. The *in vitro* results were confirmed *in vivo* in diabetic mice transplanted with pancreatic islets previously treated with BPA. The transplant with BPA-treated islets was unable to restore normal glycemic level neither in BPA treated nor in normal water administered mice at any time. Overall, it can be concluded that BPA exposure leads to disruption of insulin synthesis through enhanced oxidative stress and cell apoptosis.

**Gong et al. (2013)** explored the hypothesis that BPA could impair  $\beta$ -cell function through misfolding islet amyloid polypeptide (IAPP) into toxic oligomers causing apoptosis of  $\beta$ -cells. IAPP is a peptidic hormone co-secreted with insulin as pro-peptides and involved in glycemic control. Using an artificial micelle system, and INS-1 cells as an *in vitro* culture system of  $\beta$ -cells, it was demonstrated that BPA increases the INS-1 cell apoptosis caused by exogenous addition of IAPP. BPA treatment also resulted in enhanced levels of reactive oxygen species (ROS) and cell apoptosis. Effects were dose-dependent from 5 to 50 μM with first effects seen at 10 μM. This study is of interest because it is the only paper on BPA and IAPP which is a hormone secreted by the pancreas. However, BPA doses used are quite high and there is little information on the number of experiments performed.

**Song et al. (2012)** isolated islets from pancreas of male Sprague Dawley rats and islet morphology and  $\beta$ -cell function in the isolated rat islets was assessed after exposure to different estrogenic compounds including BPA, E2 and DES. Selected concentrations were from 0.1 to 250 μg/L. It was demonstrated that BPA, E2 and DES impacted cell viability as well as the  $\beta$ -cell insulin content, the number of insulin granules, and the area and density of mitochondria in these cells. Glucose-stimulated insulin secretion (GSIS) and expression levels of genes involved in  $\beta$ -cell function were analysed by qPCR. All the data converge at demonstrating impairment of both  $\beta$ -cell morphology and function after exposure to all three molecules although through distinct mechanisms, i.e. BPA is not mimicking all effects induced by E2 or DES. Importantly, the relationship between the doses and  $\beta$ -cell alteration was an inverted U-shape for BPA while dependent on the dose for E2 and DES. It is suggested that mitochondrial dysfunction could be an early event in the BPA-induced impairment of  $\beta$ -cells.

Two other chemicals have been tested within this experimental protocol. A total of 5 experiments were performed. It is not completely clear how the authors have taken into account the decrease of cell viability in the measurement of insulin secretion and the mRNA expression. Despite the fact that E2 while highly cytotoxic at the highest dose is also dose-dependently enhancing insulin secretion; at this highest dose, BPA is decreasing insulin secretion.

These studies described above are reported in Table 26.

#### Epigenetic data

**Mao** *et al.* **(2015)** investigated epigenetic changes following BPA exposure (via oral administration) of Sprague Dawley rats. BPA exposure during early life can result in generational transmission of glucose intolerance and  $\beta$ -cell dysfunction in the offspring through male germ line, which is associated with hypermethylation of the IGF-2 gene in islets. The changes of epigenetics in germ cells may contribute to this generational transmission. This study is one of the few investigating epigenetic changes with BPA exposure.

However, more studies are required before conclusions can be made on the BPA-induced epigenetic changes.

# 4.5.2.3 Summary of the plausible link between adverse effects and endocrine MoA: Effects of BPA on insulin synthesis

Different studies, mostly from Nadal's group, have pointed to that BPA could impair glucose homeostasis through targeting both a (Alonso-Magdalena et~al., 2005) and  $\beta$  pancreatic cells (Garcia-Arevalo 2014, 2016; Alonso-Magdalena et~al., 2006; 2008; 2015; Soriano, 2012). Mechanisms of action were found to involve an estrogen receptor dependent mechanism, the  $\beta$ , the  $\alpha$  or the membranous form depending on the species (mouse, rat, human) leading to changes in either cell proliferation and/or apoptosis and altering the  $\beta$ -cell mass.

Involvement of estrogen receptor-dependent mechanisms was highlighted using positive controls, estradiol- $17\beta$  or DES.

Two studies identified detrimental effects of BPA on insulin secretion and sensitivity with intolerance to glucose but the authors reported no changes in  $\beta$ -cell mass (Liu *et al.*, 2013; Moon *et al.*, 2015).

Other groups demonstrated that BPA exposure could provoke mitochondrial dysfunction decreasing ATP production and insulin release and generating oxidative stress leading to apoptosis of the pancreatic cells (Carchia *et al.* 2015; Song *et al.*, 2012; Makaji *et al.*, 2011). However, these authors did not examine whether E2 or DES had identical effects to those reported with BPA.

A total of 13 publications have been considered Most of them are *in vivo* studies with early life exposure. Altogether, it is demonstrated that BPA exposure during early life when pancreas differentiation occurs with  $\alpha$ - and  $\beta$ -cells differentiating into glucagon and insulin secreting cells, respectively, impairs pancreas development. Later in adult life, consequences are firstly hyperinsulinemia until the pancreas is exhausted and diabetes occurs. One paper did not identifyadverse effects in the pancreas but reduced insulin signaling in muscles as the event initiating glucose intolerance in the BPA-exposed animals (Moon *et al.*, 2015). Oxidative stress leading to  $\beta$ -cell apoptosis also happens in case of exposure during adult life as well as impaired insulin-signaling transduction with reduced phosphorylation of AKT, for example. A paper from Soriano *et al.* (2012) recapitulates several findings from Nadal's laboratory indicating that BPA may act through the distinct estrogen receptors including the nuclear  $\alpha$ -and  $\beta$ -receptors as well as the membranous form of estrogen receptor. In as much as these receptors signal distinct but overlapping mechanisms all converging to protect  $\beta$ -cell survival and  $\beta$ -cell mass as well as insulin biosynthesis and secretion, the harmfulness of BPA for pancreas development is likely to be due to an endocrine disrupting MoA.

Overall, it is suggested that the pancreas is targeted by BPA exposure and that mechanisms could differ depending on whether exposure occurs during fetal life or in adulthood. Fetal differentiation of the pancreas appears highly sensitive to BPA exposure based on the outcomes surveyed e.g.  $\beta$ -cell proliferation and apoptosis. Limited data exist on the impact of BPA on  $\alpha$ -cells and glucagon secretion. Conclusions point to BPA as a disruptor for pancreas morphology and function during fetal life resulting in alterations of insulin synthesis and/or release.

### 4.5.3 Effects of BPA on insulin resistance

4.5.3.1 Adverse effect of BPA on insulin-sensitive organs including liver and adipose tissue

The available *in vivo* experimental studies retrieved since 2013 are detailed below and reported in a tabular format in Table 25.

Angle et al. (2013) treated pregnant CD-1 mice with BPA at doses ranging from 5 to 50,000  $\mu$ g/kg bw/day, from GD9 to GD18 that resulted in average unconjugated BPA between 2 and 200pg/ml in fetal serum (AUC<sub>0-24h</sub>). BPA was fed to pregnant females once daily using a micropipetter and the volume was adjusted to 30  $\mu$ l in corn oil. There were significant effects in adult male offspring: an age-related change in food intake, an increase in body weight and liver weight, abdominal adipocyte mass, number and volume, and in serum leptin and insulin, but a decrease in serum adiponectin and in glucose tolerance. For most of these outcomes inverted U-shape dose-response curves were reported by the authors but a more refined statistical analysis did not confirm these observations. This study is convincing because of the multiple doses with multiple outcomes surveyed including a positive control (DES) with 9-14 pregnant mice and 13-17 male offspring per group. A 0.1  $\mu$ g/kg/day dose of DES resulted in some but not all low-dose BPA outcomes (eg. food consumption, renal and gonadal fat pad weight, more adipocytes and impaired glucose tolerance). However, the non-monotonic dose-response relationships suggested by the authors need to be confirmed.

Delclos et al. (2014) summarised the results of the US FDA/NCTR 2013 study. This US FDA/NCTR 2013 study was a large-scale animal study on continuous exposure to BPA from GD6 to postnatal day 90, when a number of Sprague dawley rats were euthanised and tissues were harvested. It complied with the criteria of Good Laboratory Practice (GLP) and was conducted in accordance with the specific guidelines of the NTP. The study included a wide range of tested doses  $(2.5 - 8 - 25 - 80 - 260 - 840 - 2,700 - 100,000 - 300,000 \mu g/kg$ bw/day) administered by gayage using a stomach tube (mothers or pups after 5 days) or orally for newborns from the first day after birth. This descriptive study relied primarily on the observation of overall morphological-histological and biochemical criteria. Two positive control groups for the characterisation of estrogenic effects were included in the study: treatment with 0.5 and 50 µg ethinyl œstradiol (EE2) per kg bw/day. Two negative control groups were also included (vehicle and without treatment). Although the US FDA/NCTR 2013 study did not specifically deal with metabolism and obesity as such, the following parameters were studied: weight changes of the exposed animals, levels of thyroid hormones (on postnatal days 15 and 90), cholesterol, serum triglycerides, glucose, insulin and leptin. The US FDA/NCTR 2013 study concludes that BPA has no effects on the assessed parameters for doses ranging from 80 μg/kg bw/day to 2,700 μg/kg bw/day, with the exception of an increase in AST (aspartate aminotransferase) in females on postnatal day 90 for the dose of 2,700 µg/kg bw/day; this effect was not found at higher doses (100,000 and 300,000 µg/kg bw/day).

Van Esterik et al. (2014) exposed C57BL/6 mice during gestation and lactation to 8 doses of BPA ranging from 0 to 3 mg/kg/day (4 mice per dosage group). Since BPA was introduced in the standard diet, these values are based on a body weight of 25g and a diet intake of 4.5 g per day which may be a problem because during gestation and lactation there is a high range of bw changes and food intake. Male and female offspring were surveyed for 20 weeks without further exposure to BPA. For every dose group on average 8 mice per sex (range 4–10, evenly recruited from available litters) were included for follow-up through juvenile and adult stages.. Overall the authors describe sex-dependent effects with bw increases in males possibly related to an increased overall body size rather than an increased fat mass and altered energy balance as evidenced by a dose-dependent decrease of circulating glucagon. However, these data are difficult to reconcile with the stable plasma levels of insulin and glucose; and food consumption could not be reliably surveyed because of spillages. In addition, there was an over-

representation of small litters with the highest doses of BPA introducing a bias.

In females, given that no bw changes were detected up to the age of 17 weeks, the authors decided to challenge all female mice with a high-fat diet for 6 weeks. In contrast to males, the authors observed decreased bw with BPA, decreased leptin levels, decreased fatty acids and triglycerides which are consistent with the reported increase in energy expenditure (increased locomotor activity and of Ucp1 expression in brown adipose tissue (BAT)).

Overall, the authors conclude that "Although these results suggest that BPA can program for an altered metabolic phenotype, the sexual dimorphism of effects and diversity of outcomes among studies similar in design as the present study do not mark BPA as a specific obesogen. The consistency within the complex of observed metabolic effects suggests that upstream key element(s) in energy homeostasis are modified. Sex-dependent factors contribute to the final phenotypic outcome."

In the study of Moon *et al.* (2015) already cited in the previous section (see 4.5.2.1), the authors observed decreased phosphorylation of AKT and GSK3beta in skeletal muscle indicative of insulin resistance which might be one mechanism by which BPA induces glucose intolerance.

The aim of the following study from **Veiga-Lopez** *et al.* **(2016)** was to assess the effects of prenatal exposure to BPA on postnatal metabolic outcomes, including insulin resistance, adipose tissue distribution, adipocyte morphometry, and expression of inflammatory markers in adipose tissue, as well as to assess whether postnatal overfeeding would exacerbate these effects. Female sheep (groups of 6-9) were daily injected subcutaneously with BPA (0.05, 0.5, 5 mg/kg/day from day 30 to day 90 of gestation, term 147d). In study 1, metabolic tests were made in pre- and post-pubertal F1 sheep at the age of 17 months. The authors described intolerance to glucose and reduced insulin sensitivity in post-pubertal F1 but no effect in prepubertal sheep. In study 2, F1 sheep were fed a high fat diet starting at the age of 14 weeks ending by 19 months of age.

The authors observed glucose intolerance and insulin resistance in the high-fat fed animals. BPA did not impact these parameters. The population of adipocytes was also analysed; normally there is a bimodal repartition with a population of small adipocytes and a population of large adipocytes full of lipids. Interestingly, there was a shift in response to BPA towards more hypertrophic adipocytes, also evidenced in response to the high-fat diet. Notably, there was no further aggravation in sheep exposed to BPA and fed the high-fat diet. This indicates that both challenges (BPA and High-fat diet) lead to similar defects.

Yang et al. (2016) prepared primary cultures of adipocyte progenitors from the stromal vascular fraction (SVF) recovered from the white adipose tissue (WAT) of C57BL/6 male mice. They found that addition of 50  $\mu$ M BPA but not of lower doses on confluent cells resulted in an increase in the expression of of C/EBPa, PPAR $\gamma$ , FABP4 through a mechanism involving GR (use of RU486 as a glucocorticoid antagonist). This study is presented together with in vivo data showing enhanced body weight and fat mass by oral intake of BPA for 30 days in both sexes when fed a chow diet but not a high fat diet (n=9-12/group). BPA doses ranged from 5 to 5000  $\mu$ g/kg/day. In addition, mice exhibited increased circulating inflammatory factors and leptin plasma levels; and there was local inflammation in the WAT. Glucose tolerance was not changed and insulin sensitivity was not studied.

**Biasiotto** *et al.* **(2016)** treated a group of pregnant C57BL/6 mice with BPA daily by gavage at the doses of 0.5, 5, 5, 500  $\mu$ g/kg/day (consistent with the 0.85  $\mu$ g/L of BPA found in municipal water) and gavages continued on F1 males until the age of 140 days with an interruption from birth to weaning. Groups ranged from 16 to 19 mice. Mice were fed a standard diet. The number of pregnant females is not given but it can be estimated to be around 5-6 per dose level. Fat mass and body weight enhanced starting from day 90 in male

mice dosed with BPA 5  $\mu$ g/kg/day probably linked to the enhancement in the epididymal weight. Several genes including PPAR $\gamma$ , ATGL, HSL and LPL were analysed in epididymal fat and liver and they were found to be significantly enhanced in BPA-treated samples over control even though differences were subtle.

#### **Human studies**

In the study from **Yang** *et al.* **(2016)** reported above, the authors have studied the association of BPA levels (subdivided in quartiles) with leptin and TNFa plasma levels within a cohort of 228 subjects. Leptin levels reflect adiposity because the hormone is mostly produced by adipocytes. TNFa is a marker of inflammation. The authors observed that urinary BPA concentrations were associated with leptin and TNFa plasma levels in lean female subjects but not in lean male subjects. No association was found in overweight/obese subjects of both sexes between leptin and TNFa plasma levels and the urinary BPA quartiles. The authors concluded that BPA may interact with body mass index (BMI) and/or diet composition to affect adiposity or inflammation.

**Menale** *et al.* **(2016)** prepared adipocytes from subcutaneous explants recovered from children undergoing orchidopexy surgery. Adipocytes were treated with BPA (1, 10, 100 nM) and adiponectin and resistin were measured by RT-qPCR as an index of insulin sensitivity. The authors demonstrated a significant down-regulation of adiponectin at the 2 highest doses of BPA. In addition, resistin could only be quantified in BPA-treated cells. These findings are indicative of a reduced sensitivity to insulin. Clinical and biochemical features of 141 obese children were collected. Serum resistin and adiponectin were evaluated. Insulin resistance and urinary BPA levels were assessed. The authors found a direct association between urinary BPA levels and HOMA (r = 0.23; p: 0.0069). The association remained significant when adjusted for BMI, sex, age (135 children, 80 boys, mean age of 10.5 +/- 2.3 years). The authors also observed a strong inverse association between BPA and adiponectin (r = -0.48; p < 0.0001). In their conclusion, the authors suggested the involvement of BPA in the development of insulin resistance in childhood obesity highlighting that urinary BPA levels are directly associated with insulin resistance regardless of BMI.

### 4.5.3.2 Mechanism of action

Literature search analysis of the *in vitro* experiments was subdivided considering, on the one hand, studies using the 3T3-L1 cell line, which is a mouse cell line very commonly used and human mesenchymal cells or cultured cells from human adipose explants on the other hand. These studies are described below and reported in Table 26.

### In vitro studies using murine 3T3-L1 cells

Three different experimental protocols were developed by **Biemann** *et al.* **(2012)** to cover the phases of undifferentiated growth, induction of differentiation and terminal differentiation of adipocytes which are occurring *in vivo*. The authors demonstrated that BPA ( $10 \mu M$ ) had an inhibitory effect during the undifferentiated growth and no effect at the 2 other phases. Three independent experiments were performed. The authors conclude that "These findings indicate BPA as an EDC, which reduces the determination of multipotent stem cells to the adipogenic lineage. (...) Regarding potential mechanisms of action estrogenic signaling triggered by BPA seems to be a plausible mechanism to reduce the commitment of Mesenchymal Stromal Cells (MSC) into adipocytes (...) in accordance with the general conclusion from in vivo studies that obesity is associated with reduced estrogen signaling and that estrogens may act antiadipogenic". The conclusions drawn by the authors are in accordance with the findings described in the study.

**Pereira-Fernandes** *et al.* **(2013)** developed a screening system for obesogenic compounds and the obesogenic properties of BPA were evaluated using the 3T3-l1 model **(Pereira Fernandes** *et al.*, **2014)**.

First study: The aim was to develop a reproducible and standardised protocol for the adipocyte differentiation assay to use as an *in vitro* tool for obesogenic compounds screening. It was based on PPARy transactivation and antagonist studies considering that PPARy signaling is a major regulator of differentiation. The culture system concerned the differentiation step and not the proliferation step. Cells were confluent in all the plates and BPA (or other compounds to be tested) was either added alone or in the presence of insulin to evaluate interactions. Outcomes included a lipid accumulation fluorescent test and the development of a PPARy CALUX cell line. Positive controls included the reference compounds rosiglitazone (ROSI) and TBT which are PPARy agonists and T0070907, a potent PPARy antagonist. Exposure to 12.5, 25, and 50  $\mu$ M of BPA resulted in enhanced lipid accumulation by less than 1.5 fold-change. There was a 2 fold-change at the highest doses when BPA was combined with insulin. Addition of ROSI 100 nM resulted in a 5 fold-change and 3.5 fold-change if combined to insulin. Finally, a very weak, although significant, increase in PPARy activity (<1.2 fold-change *versus* >15 with ROSI 1  $\mu$ M) was detected in cells treated with the highest dose of BPA. The authors conclude that there was weak obesogenic activity for BPA in their model.

Second study: The authors used the 3T3-I1 model to analyse and compare the transcriptomic profile of cells exposed to various compounds including ROSI, TBT and BPA. The authors stated that: "Based on the transcription data, BPA was the most distinct compound of this group, indicating that this compound might act through a different mechanism of action. Indeed BPA is the sole compound of this cluster that only weakly activated the PPAR receptor. Together all these studies show that the obesogenic mechanism of action of BPA remains enigmatic, but seem to be different from the frequently observed PPAR mediated obesogenicity. Including other obesogenic compounds with low PPAR activation capacity in future microarray experiments could further confirm the distinction of the BPA gene expression profile compared with PPAR agonists"

In conclusion, the authors considered BPA as weakly obesogenic with a mechanism of action distinct from PPARy-mediated.

**Valentino** *et al.* **(2013)** investigated the impact of nM doses of BPA (1 and 10 nM) on differentiated 3T3-L1 cells. While markers of differentiation (Glut4, PPARγ) did not change with BPA exposure, the authors described reduced glucose utilisation and insulin signaling (measured by phosphorylation levels of IR, AKT/PKB and phosphor ERK), and leptin mRNA levels, all converging to BPA impairing insulin action.

**Atlas et al. (2014)** investigated the impact of BPA on differentiated 3T3-L1 cells 2 days post-confluence in the presence of insulin 100nM. No dexamethasone (DEX) was included in the differentiation medium. BPA was added at 0.1 nM, 1 nM and 10 nM. DEX 1 nM and 250 nM were used as a positive control. Within these conditions, addition of BPA resulted in a dose-dependent increase of lipid accumulation, FABP4, adipsin. There was no induction of C/EBPa or PPARy in contrast to DEX (also shown using reporter genes coupled to luciferase). However, BPA could potentiate the transcriptional complex containing GR and C/EBP at the promoter of FABP4. Enhancing FABP4 is clearly indicative for an adipogenic mechanism of action but it is not PPARy- mediated.

**Héliès-Toussaint** *et al.* **(2014)** investigated the impact of 1fM, 1 pM, 1 nM and 1 microM (100  $\mu$ M was found cytotoxic) on confluent and differentiated 3T3-L1cells. Findings include enhanced lipid accumulation (as shown with DES and ROSI), increased lipolysis (not seen with DES), no effect on glucose uptake (seen with ROSI not with DES) and no effect on Leptin (DES has a negative effect and ROSI a positive effect); a very light effect on Srebp1C, PPAR $\gamma$  and FABP4 mRNA levels and enhancement of ERR $\alpha$  and ERR $\gamma$  mRNA levels (not shown for DES or ROSI). The authors conclude that BPA could activate adipocyte differentiation through binding to ERR $\alpha$  or ERR $\gamma$ . Based on the data shown in the paper, it is not possible to determine if BPA modulated insulin sensitivity as no effect was described on glucose uptake.

**Ariemma et al. (2016)** investigated the effects of adding BPA (1nM) from plating throughout the phase of proliferation and differentiation of the 3T3-L1 cells. They demonstrated enhanced FABP4 mRNA and protein; PPARy and C/EBPa were also enhanced, so were leptin and IL6. Glucose utilisation and insulin signaling were reduced. All data converge to show adipocyte metabolic dysfunction and inflammation and decreased insulin sensitivity in 3T3-L1 cells.

**Biasiotto** *et al.* **(2016)** investigated the response of 3T3-L1 cells to BPA and found that it promoted adipocyte differentiation at the concentration of 50 and 80μM. BPA effect in 3T3-L1 cells was associated with the specific activation of the ERa in undifferentiated cells and the ERβ in differentiated cells. BPA also activated the PPARγ upregulating a minimal 3xPPRE luciferase reporter and the PPARγ-target promoter of the Fabp4 (alias aP2) gene in adipose cells, while it was not effective in preadipocytes (undifferentiated cells). The pure estrogen receptor agonist DES displayed an opposite action to that of BPA inhibiting PPARγ activity in adipocytes, preventing cell differentiation, activating ERa in preadipocytes and inhibiting ERa and ERβ regulation in adipocytes. Three independent in vitro experiments were performed. The in vitro protocol is not classical with a mix of the proliferative and differentiation phases. Initial plating of cells is very low with  $10^3$  cells per 12-well culture plate (instead of 50,000 cells).

**Ahmed et al. (2016)** treated 3T3-L1 cells with BPA 25  $\mu$ M. It resulted in enhanced lipid accumulation and increased mRNA and protein expression of key adipogenic markers (P<0.05) including lipoprotein lipase, adipocyte protein 2, PPAR $\gamma$ , perilipin, and adipsin. Furthermore, using transcriptional assays, BPA was found to modestly activate PPAR $\gamma$  using a PPRE (PPAR $\gamma$  response element)-dependent luciferase construct by 1.5-fold (P<0.05). Co-treatment of cells with the selective PPAR $\gamma$  antagonist GW9662 inhibits BPA-, ROSI- but not DEX-dependent adipogenic differentiation, indicative that BPA requires PPAR $\gamma$  and not GR to induce adipogenesis.

The paper of **Dai et al.** (2016) was not read (not available). The abstract indicates that the authors treated 3T3-L1 adipocytes for 0, 2, 6, 12 and 24 h with BPA at 80  $\mu$ M in serum-deprived medium. The sensitivity of adipocytes to insulin was measured by the ability of insulin to phosphorylate the insulin receptor substrate 1 (IRS-1) and the AKT protein. Western blotting analysis indicated constant levels of AKT and IRS1 proteins but decreased levels of the phosphorylated forms in the BPA-treated cells.

### In vitro studies using murine primary cells culture

Yang et al. (2016) have prepared primary cultures of adipocyte progenitors from the stromal vascular fraction (SVF) recovered from the white adipose tissue (WAT) of C57bl6 male mice. They found that treatment of confluent cells with BPA (50  $\mu$ M) but not of doses of BPA lower than 50  $\mu$ M resulted in a stimulation of C/EBPa, PPARy, FABP4 through a mechanism involving GR (use of RU486). This study is presented together with *in vivo* data (see 4.5.3.1).

## <u>In vitro studies using human cells (either cell lines or cultured from explants)</u> <u>published from 2013 to the end of May, 2016:</u>

In the study from Chamorro-Garcia et al. (2012), murine and human multipotent mesenchymal stromal stem cells (MSCs) and murine 3T3-L1 cells were used to study the adipogenic capacity of BPA (1, 10, 100, 1000, 10000 nM) assessed by lipid accumulation measurement (oil red O staining and FABP4 mRNA and protein levels). BPA enhanced significantly lipid accumulation and FABP4 at the 3 highest doses in 3T3-L1 cells. In contrast, no effect was observed with the 2 MSCs populations used.

Wang et al. (2013) investigated the impact of 3 doses of BPA 10 nM, 1  $\mu$ M, 80  $\mu$ M in adipocytes recovered from omental biopsies of children (boys and girls neither overweight nor

obese) with an age range of 3 to 13 years for girls (10 for boys). The authors demonstrated enhanced 11-bHSD1 mRNA and activity in adipocytes treated with BPA 10nM and 80  $\mu$ M, and enhanced PPAR $\gamma$  and LPL mRNA levels. The enzyme 11-bHSD1 converts the inactive cortisone into the active hormone cortisol (corticosterone in rodents) in adipose tissues and promotes adipogenesis. To determine mechanisms of action of BPA, they used human cell lines isolated from human visceral fat tissue. The authors demonstrated promotion of adipogenesis (lipid accumulation) and enhanced 11-bHSD1, PPAR $\gamma$  and LPL mRNA levels. Addition of CBX, the 11-bHSD1 inhibitor or of RU486 to inhibit glucocorticoid signaling prevented partially the BPA-induced effects on 11-bHSD1 mRNA levels.

**Valentino et al. (2013)** investigated the impact of nM doses of BPA (1 and 10 nM) on human adipocytes prepared from a biopsy of subcutaneous WAT. The authors also used 3T3-L1 cells (see above). While markers of differentiation (Glut4, Pparγ) did not change with BPA exposure, the authors described reduced glucose utilisation and insulin signaling (measured by phosphorylation levels of IR, AKT/PKB and phospho ERK), and leptin mRNA levels, all converging to BPA impairing insulin action.

In **Boucher** *et al.* **(2014)**, primary human pre-adipocytes were differentiated in the presence of 50  $\mu$ M BPA or 1  $\mu$ M DEX for 48 hours and gene expression microarray analysis was developed to determine potential mechanisms of BPA. Transcriptomic profiling shows enrichment in genes involved in adipogenesis and in other specific genes depending on treatment. Specifically BPA treatment resulted in enrichment of genes associated with the SREBF1 but also mTOR and Thyroid receptor/RXR signaling.

**Ohlstein et al.** (2014) demonstrated very strong effects of low doses of BPA (100 nM and 1 microM) on adipogenic differentiation of human adipose stromal cells with a 6-fold increase of PPAR $\gamma$ , 65-fold increase of Era and 20-fold increase of ER $\beta$ , effects that were inhibited if pretreating cells with 100 nM ICI, indicating mechanisms largely under estrogenic regulation. However, it seems not plausible to get such extremely large changes as observed in this study.

**Menale et al.** (2015) prepared adipocytes from subcutaneous explants recovered from children undergoing orchidopexy surgery (total of 8 explants). Adipocytes were either treated with E2 (1nM) or with BPA (1, 10, 100 nM) and gene profiling performed. The analysis of deregulated genes in response to E2 allowed the identification of a small group of genes that are expressed in an opposite manner from that of adipocytes treated with BPA. In particular, BPA increases, whereas E2 decreases the expression of pro-inflammatory cytokines and the expression of FABP4 and CD36, two genes involved in lipid metabolism. In addition, using a human pancreatic cell line, the authors found that BPA could decrease the expression of PCSK1, a gene involved in insulin production. These results indicate that BPA can exhibit opposite effects of those induced by E2.

**Menale et al.** (2016) have prepared adipocytes from subcutaneous explants recovered from children undergoing orchidopexy surgery. Adipocytes were treated with BPA (1, 10, 100 nM) and adiponectin and resistin were measured by RT-qPCR as an index of insulin sensitivity. The authors demonstrated a significant down-regulation of adiponectin at the 2 highest doses of BPA. In addition, resistin could only be quantified in BPA-treated cells; all these observations indicate a reduced sensitivity to insulin.

# 4.5.3.3 Summary of the plausible link between adverse effects and endocrine MoA: Effects of BPA on insulin sensitivity

Only studies showing reduced levels of adiponectin, which is a marker of insulin sensitivity have been considered here, as well as studies describing reduced insulin sensitivity coupled or not with reduced glucose tolerance, as they demonstrate reduced capacity to respond to hyperglycemia.

Angle *et al.* (2013) demonstrated metabolic disruption through endocrine disruption in male mice exposed *in utero* to BPA with alteration in serum leptin, insulin and adiponectin and in glucose tolerance. They have also shown that some of the effects are mimicked by DES which is indicative of estrogen dependency.

Veiga-Lopez *et al.* (2016) demonstrated in *in utero* BPA -exposed female sheep, intolerance to glucose coupled with insulin sensitivity in post-pubertal, but not pre-pubertal F1 sheep illustrating an endocrine MoA for BPA.

Finally, a study performed on 141 obese children concluded that there was a strong inverse association between BPA and adiponectin (Menale *et al.*, 2016).

Taken together, using distinct animal models and a cohort of obese children, these 3 recent studies established that BPA acts on the efficiency of insulin on its target cells, and thus *via* an ED MoA.

However, two other studies from Van Esterik  $et\ al.\ (2014)$  and Delclos  $et\ al.\ (2014)$  did not provide evidence for strong effects indicative of BPA as an endocrine disrupter. Two of the studied doses in Delclos  $et\ al.\ (2014)$  are the same as those in the study by Miyawaki  $et\ al.\ (2007)\ (260\ and\ 2,600\ \mu g/kg\ bw/day)$  used as a key study in the ANSES restriction dossier (2014). However, these two studies differ on many methodological points, which may explain why BPA had no effects on metabolism at these two doses in the Delclos study (2014) whereas Miyawaki  $et\ al.\ (2007)$  reported effects on body weight, adipose tissue weight, serum leptin levels, triglyceridemia, non-esterified fatty acids and glucose. These differences involve the animal model (rats in the Delclos study and mice in the study by Miyawaki  $et\ al.\ (2007)$ , the exposure route, the administration mode and vehicle used (gavage versus drinking water), the exposure period (post-coitum day 6 to postnatal day 90 versus post-coitum day 6 to postnatal day 30), age of examination (adult versus juvenile stage), and diet (standard diet versus highfat diet (30% kcal) in Delclos and Miyawaki study respectively). Furthermore, the animals were subject to fasting from the day before the study for Miyawaki  $et\ al.\ (2007)$ , whereas this indication is not given in Delclos  $et\ al.\ (2014)$ .

Thus, it can be concluded that there is some evidence that BPA elicits endocrine mechanisms of action to disrupt glucose homeostasis.

Mechanisms of action of BPA have mostly been examined using the 3T3-L1 cells: there are several papers indicating an alteration of endocrine activity with reduced insulin sensitivity upon exposure to BPA. For example, it has been demonstrated that exposure to BPA reduced estrogen signaling (Biemann *et al.*, 2012); reduced glucose utilisation and insulin signaling (indicative of resistance to insulin) (Valentino *et al.*, 2013); potentiated the transcriptional complex containing GR and C/EBP at the promoter of FABP4 (indicative of enhanced adipogenesis) (Atlas *et al.*, 2014); promoted adipocyte differentiation through the specific activation of the estrogen receptors  $\alpha$  or  $\beta$  (Biasiotto *et al.*, 2016); and altered the ability of insulin to phosphorylate downstream effectors strongly indicating that BPA was acting as an EDC (Dai *et al.*, 2016).

In other studies, BPA has been shown to decrease insulin sensitivity as a consequence of enhanced inflammation (Ariemma *et al.*, 2016). It is also debated whether BPA activates PPARy which is a master transcription factor of adipogenesis. For example, it has been shown that BPA could weakly activate the PPARy receptor (Pereira-Fernandes *et al.*, 2013; 2014); others demonstrated that BPA required PPARy to induce adipogenesis (Ahmed *et al.*, 2016) or initiated adipocyte differentiation through binding to ERRa or ERRV (Héliès-Toussaint *et al.*, 2014); but with no indication of insulin sensitivity.

It has been demonstrated that metabolic actions of estrogen receptor beta  $(ER\beta)$  are mediated by a negative cross-talk with PPARy resulting in improved insulin sensitivity in male mice

knockout for ER $\beta$ , although obese because of enhanced activation of PPAR $\gamma$  (Foryst-Ludwig *et al.*, 2008).

**Human cells** (either cell lines or cultured explants) have also largely been used to explore the effects of BPA. Down-regulation of adiponectin release, a marker of insulin sensitivity, was demonstrated by Menale *et al.* (2016) using adipocytes from subcutaneous explants recovered from children undergoing orchidopexy surgery. Alteration of glucocorticoid signaling was evidenced with adipocytes recovered from omental biopsies of normal weight children (Wang *et al.*, 2013) as well from explants of C57bl mice treated in vivo with BPA (Yang *et al.*, 2016). Reduced glucose utilisation coupled to alteration of insulin signaling was observed in human subcutaneous adipocytes (Valentino *et al.*, 2013). A transcriptomic analysis pointed also to alteration of the thyroid receptor signaling pathway (Boucher *et al.*, 2014) or to a decrease in the expression of PCSK1, a gene involved in insulin production (Menale *et al.*, 2015).

In conclusion, the *in vitro* studies on adipocyte differentiation and function point to an alteration of endocrine mechanisms (e.g., adiponectin release, insulin signaling cascade effectors). It is not clear whether BPA activates PPARγ and/or other nuclear receptors. The importance of cross-talk between nuclear receptors must be kept in mind.

### 4.5.4 Human data

In the previous risk assessments (ANSES, 2014, EFSA 2015; ECHA, 2015) it was concluded that based on available epidemiological studies, an association between BPA exposure and metabolic effects in humans cannot be established. Most of the studies were indeed cross-sectional and thus not suitable on their own to establish a causal inference and only few were prospective studies.

In the EFSA risk assessment, five studies examining urinary BPA and diabetes outcomes (Ning et al., 2011; Shankar et al., 2012; Silver et al., 2011; Lakind et al., 2012; Kim and Park, 2013) were reported (EFSA, 2015). All were cross sectional by design and relied on spot urine sampling for BPA exposure assessment. The study by Wang et al. (2012a) found that in addition to being associated with increased prevalence of obesity, higher urinary BPA was also associated with increased prevalence of insulin resistance in 3390 Chinese adults aged 40 years or older. Ning et al. (2011) studied 3423 Chinese adults and defined type-2 diabetes from fasting- and 2-h glucose tolerance test (GTT) and serum insulin levels. Increased risk of type-2 diabetes was seen for participants in the second and fourth BPA quartiles, but not in the third. A study in 1210 nationally representative Korean adults aged 40-69 years found no association between urinary BPA and self-reported type-2 diabetes (Kim and Park, 2013).

Two cross-sectional studies used NHANES data (Shankar *et al.*, 2011; Silver *et al.*, 2011). Shankar *et al.* (2011) examined 3967 adults in pooled data from 2003 to 2008 and examined type-2 diabetes diagnosed by fasting glucose levels and glycosylated haemoglobin according to the latest American Diabetes Associations guidelines. The risk of type-2 diabetes increased with increasing quartiles of BPA in a dose-dependent manner.

**Silver** *et al.* **(2011)** examined 4389 adults and also used pooled data from 2003 to 2008, and defined diabetes 2 as glycosylated haemoglobin  $\geq$ 6.5% or if participants used diabetic medication. A weak association between BPA and type-2 diabetes mellitus was seen in 2003-08 pooled data. Breaking down by year, the association was only significant in 2003/04, not 2005/06 or 2007/08. Results were similar when glycosylated haemoglobin was used as a continuous outcome. It is unclear whether the studies by Silver *et al.* (2011) and Shankar *et al.* (2011) report the same association or are independent studies. Both studies used a population in which the association was already described before by Lang *et al.* (2008) and Melzer *et al.* (2012).

Lakind et al. (2012) conducted a re-analysis of the associations between BPA exposure and

chronic disease outcomes, including diabetes, using four available NHANES data sets, including the same data used in the studies above. Scientifically and clinically supportable exclusion criteria and outcome definitions were applied. All analyses were adjusted for creatinine, age, gender, race/ethnicity, education, income, smoking, heavy drinking, BMI, waist circumference, calorie intake, family history of heart attack, hypertension, sedentary time, and total cholesterol. When the a-priori selected methods were used to address the research question, no associations were found between urinary BPA and diabetes. The authors concluded that the discrepancy between their findings with regard to diabetes and those reported previously (Lang et al., 2008; Melzer et al., 2010) was largely explained by the choice of case definition. The Lakind et al., 2012 study did not support the associations and causal inferences that were suggested in the previous studies, and highlighted that data from cross sectional studies like NHANES surveys are inappropriate for drawing conclusions about relations between short-lived environmental chemicals and chronic diseases.

Since the publication of the previous risk assessment reports, several human studies investigating association between BPA exposure and metabolic outcomes were published. A systematic review of the literature published in 2013, 2014 and 2015 was performed to check if the conclusions, which were reported in these reports, are still valid. Among all publications retrieved, 4 were reviews and therefore excluded (Liu *et al.*, 2015; Ranciere *et al.*, 2015; Song *et al.*, 2015; Chrysant, 2015).

The other studies were assessed based on the title and summary and criteria were used to classify studies that could be used in a weight of evidence approach to assess a causal link between BPA exposure and effects on metabolism.

#### These criteria are:

- Type of study: cross-sectional studies are not designed to assess a causal link and therefore were not further considered. Only prospective studies may be used to discuss a causal link.
- Validity of BPA exposure estimation: most of the studies relied on spot urine BPA measures which are not appropriate to assess properly exposure, as explained above. The study of Agay-Shay *et al.* (2015) is the only one with 2 measures. Therefore this criterion was not used to exclude studies. This consideration should be taken into account when analyzing the retrieved studies.
- Assessment of the outcomes: some studies consider biological parameters (leptin, adiponectin), other considered diseases (hypertension, type 2-diabetes) or anthropometric characteristics such as BMI and body weight. Some of these parameters may be considered as adverse outcomes whereas others are only indicative of health status without any direct link with a disease.
- Size of the samples and representativity of the population compared to the general population.

The following studies that are based on a cross sectional design were not further assessed. However they may be used to elaborate some hypothesis.

The study of Leclerc *et al.* (2014) was excluded due to the small sample size and lack of representativity of the population included in the study. The studies from Aekplakorn et al. (2012, 2015), Ko *et al.* (2014), Choi *et al.* (2014), Khalil *et al.* (2014), Beydoun *et al.* (2014), Ronn *et al.* (2014), Xiong *et al.* (2015), Xue *et al.* (2015), Lee *et al.* (2015), Lin *et al.* (2015) and Savastano *et al.* (2015), were excluded because they were strictly transversal studies. The case-control study from Ahmadkhaniha *et al.* (2014) was also excluded due to the small sample size and limited information on the control group and their health status.

Four prospective cohort studies should be further considered.

In the study from Shapiro *et al.* (2015), no statistically significant associations were observed between BPA with impaired glucose tolerance (IGT) or gestational diabetes mellitus (GDM).

In the study from Hu et al. (2015), the authors showed that serum BPA could predict the progression of chronic kidney disease (CKD) in patients with type 2 diabetes mellitus (T2DM).

In the nested case-control studies within the cohorts of the Nurses' Health Study (NHS) and of the Nurses' Health Study II (NHSII), Sun *et al.* (2014) reported some diverging results concerning the association between BPA exposure and type 2 diabetes. In the NHSII, BPA levels were not associated with incident T2DM in multivariate-adjusted analysis until BMI was adjusted: odds ratio (ORa) comparing extreme BPA quartiles increased from 1.40 (95% CI: 0.91,

Concerning anthropometric characteristics, the study from Agay-Shayet *al.*, (2015) in which measurements of BPA in two maternal pregnancy urine samples were performed did not show any association with body weight in the children until 7 years old. Song *et al.* (2014) reported a slight but significant increase in body weight. The study population was from the controls in a prospective case-control study of T2DM in the NHS and NHSII. A total of 977 participants provided first-morning-void urine samples in 1996-2002. Urinary concentrations of BPA were measured using liquid chromatography-mass spectrometry. Body weights were self-reported at baseline and updated biennially thereafter for 10 years. On average, the women gained 2.09 kg (95% confidence interval (CI), -2.27 to 6.80 kg) during the 10-year follow-up. In multivariate analysis with adjustment of lifestyle and dietary factors, in comparison with women in the lowest quartile of BPA concentration, those in the highest quartile had 0.23 kg per year (95% CI, 0.07-0.38 kg per year) greater weight gain during the 10-year follow-up (P-trend=0.02).

Finally, using 188 mother-child pairs from the CHAMACOS prospective study (Volberg *et al.*, 2013), BPA was measured in urinary spot samples during early and late pregnancy and in 9-year-old children. BPA concentrations during late pregnancy were associated with increased plasma lectin in boys and BPA concentrations during early pregnancy were associated with plasma adiponectin in girls. By contrast, no associations were found between BPA concentrations in 9-year old children and adiponection or leptin at the same age.

In conclusion, in view of the limitations of using urinary BPA concentrations as a surrogate of exposure, the problems of interrelated dietary exposures, mostly cross-sectional designs and inconsistency of the results between cross-sectional and prospective studies, the conclusions that can be drawn concerning the relationship of BPA exposure and the reported findings are limited.

Notwithstanding this, there are indications from cross-sectional studies that higher BPA may be associated with increased body mass in children, and indication from a prospective study (Volberg *et al.*, 2013) that prenatal BPA exposure may be associated with reduced body mass and lower plasma adiponectin levels in girls and with higher plasma leptin levels in boys. There are no indications of note for other hormonal or metabolic endpoints. A systematic literature review of the epidemiological literature on the relation of BPA with obesity and markers of glucose metabolism and diabetes concluded that assertions about a causal link between BPA and obesity or diabetes are unsubstantiated (Lakind *et al.*, 2014).

### 4.5.5 General conclusion on metabolism and obesity

In conclusion, based on animal studies (rodents and non-rodents) after prenatal and/or perinatal or adult exposure, there is now evidence that BPA may increase the incidence of type-2 diabetes  $\emph{via}$  an ED MoA. In particular, BPA has been shown to alter insulin secretion and/or release by  $\beta$ -pancreatic cells, or insulin signalisation (signaling mechanisms) within

insulin-sensitive organs (i.e., liver, muscle, adipose tissues). This resulted in variations in the expression levels of hepatic or adipose tissue markers which are indicative of a state of insulin resistance. These effects are considered by the experts as hallmarks of endocrine disruption mechanisms, especially if there is a combination of effects each leading to insulin resistance within the different insulin-sensitive tissues. In addition, while most studies were performed on males, a few studies have also examined the impact of BPA either on both sexes or on females. However, more studies should be undertaken before one can conclude on a sexspecificity or not of the metabolic impact of BPA.

Recent experimental *in vivo* and *in vitro* studies indicate that these effects may involve ERa, ER $\beta$  or GPR30 pathways. Other hormones such as leptin and adiponectin, which are involved in resistance to insulin and lipogenesis, are also modified following BPA exposure. This shows that BPA could interfere in the balanced interplay between insulin secretion and insulin action that controls glycaemia.

Overall, it is suggested that the pancreas is targeted by BPA exposure and that mechanisms could differ depending on whether exposure occurs during the fetal life or in adulthood. Fetal differentiation of the pancreas appears highly sensitive to BPA exposure based on the outcomes surveyed e.g.  $\beta$ -cell proliferation and apoptosis. Limited data exist on the impact of BPA on  $\alpha$ -cells and glucagon secretion. Conclusions indicate that BPA can elicit histopathological modifications during the fetal life, with consequences on insulin synthesis rate and/or release.

Moreover, most of the *in vitro* studies showing adverse effects of BPA on adipocyte differentiation and function point to alteration of endocrine mechanisms (e.g., adiponectin release, insulin signaling cascade effectors). It is not clear whether BPA activates PPARγ and/or other nuclear receptors. Cross-talk between nuclear receptors may explain these uncertainties.

Even if available epidemiological studies are inconclusive, these effects are considered relevant for humans because similarities exist in homeostatic regulation of insulin production and sensitivity between animals and humans and because of *in vitro* experimental data using human cells or tissue.

## 4.5.6 Summary tables of studies

Table 25: In vivo experimental study on effects of BPA on glycaemia and insulin synthesis.

Ref.	Species Strain Model	Routes	Dose Exposure period	Group size	Outcomes reported				Conclusions of the authors	Comments of the expert
Angle et al., 2013	Pregnant CD1 mice	ation with a pipette not by	5, 50, 500, 5000, 50000 BPA. 0.1 DES ug/kg/d From GD9 to GD18. 30 μl of BPA	9-14 pregnant mice and 13-17 male offspring	age-related change in food intake, an increase in bw, liver weight and abdominal adipocyte mass (nb, volume),	increase in serum leptin and insulin; decreased in serum adiponectin and in glucose tolerance	non-monotonic dose-response curves with no effect of the highest dose	not all effects were seen with DES; statistic tests should be looked at	Metabolic disruption in male mice due to fetal exposure to low but not high doses of BPA with effects on bw, food intake, adipocytes, leptin, adiponectin, insulin and glucose regulation.	convincing study because of the multiple doses (TDI up to 10-fold the NOAEL ), multiple outcomes, positive control (DES), 9-14 dams, 13-17 males/ group
Jayas hree et al., 2013	Wistar rat	oral	Control and BPA 20 mg/kg and 200 mg/kg 4 weeks	n= 6/group	<ul> <li>         ⊅ blood insulin with BPA dose         dependently         No effect on glucose but ↓ in         glucose oxidation         ↓ in glycogen content         ↓ testosterone with BPA     </li> </ul>		↓ insulin receptor and Akt mRNA and protein kinase B with BPA 200 mg/kg >GLUT2 mRNA and its protein with BPA 20 and 200		The authors conclude that BPA impairs hepatic glucose oxidation and glycogen content through defective signal transduction	There is no indication of the BWof the animals and the doses used are extremely high
Liu et al., 2013	Pregnant C57bl6 mice	Sc BPA (100 µg/kg/d) or vehicle	- GD1-GD6, (preimplantat ion exposure); - GD 6 - PND0 (fetal exposure); - PND0- PND21 (neonatal exposure); and - GD6- PND21, fetal and neonatal exposure)	N = 15-30 mice/ group	Glucose homeostasis and Insulin release *gluc tolerance in F1 females - PND0-PND21: no effect of BPA - GD6-PND21: no effect of BPA -GD6-PND0: gluc intolerance at 3 and 6 months - GD1-GD6: impaired gluc tolerance at 6 m but not at 3m - no difference in gluc tolerance at 8m in all groups * gluc tolerance in F1 males - GD1-GD6: no effect of BPA -GD6-PND21: gluc intol at 3m - GD6-PND0: gluc intol at 3m, 6m		Insulin sensitivity (IpITT)  * in females: -GD1-GD6: no effect - PND0-PND21: no effect - GD6-PND0: ↓ insulin sensitivity at 3m and 6m  * in males: - GD1 -GD6: no effect - GD6-PND21: ↓ insulin sensitivity at 3m - PND0-PND21: ↓ insulin sensitivity at 3m - PND0-PND21: idem - GD6-PND0: idem and until 8m		Glucose homeostasis was impaired in GD6-PND0 mice from 3 to 6m of age, and until 8m in males. In PND0-PND21 and GD6-PND21, BPAtreated group, only the 3m old male F1 developed the glucose intolerance. Moreover at the age of 3m, perinatal exposure to BPA resulted in the γ in β-cells mass mainly due to the coordinate changes in cell replication, neogenesis and	Glucose homeostasis was impaired in males more than in females as assessed through handling glucose tolerance metabolic tests. After exposure to BPA, the b-cell mass > while insulin secretion was either \u2214 or remained invariable. It suggested that beta cells were less functional in BPA-exposed mice. The alterations of insulin

					* insulin release in F1 females: - GD6-PND0: ↓insul release at 3m but improved at 6m and normal at 8m - GD1-GD6: ↓insul release at 6m - PND0-PND21: no effect - GD6-PND21: no effect  * insulin release in F1 males: - GD6-PND0: ↓insul release at 3m and 6m - PND0-PND21: ↓insul release at 3m but ≠ at 6m - GD6-PND21: ↓ insul release at 3m but ≠ at 6m - GD1-GD6: no effects  Normal at 8 m in all groups	Insulin secretion after glucose stimulation in vitro   in GD1-GD6 females  in GD6-PND0 females and males and in PND0-PND21 and GD6-PND21 males  No effects at 8m in all groups  Islet morphologic analysis  β-cells mass:  - GD1-GD6: no effect in m and f  -GD6-PND21: /* 'β-cells mass in m and f  - GD6-PND0: : /* β-cells mass in m only  - PND0-PND21: /* β-cells mass in m only  β-cells turnover:  - GD6-PND0 ↓ in females  - PND0-PND21 ↓ in males  - GD6-PND21: idem	apoptosis.	secretion and insulin sensitivity, rather than b-cell mass, were consistent with the development of glucose intolerance.  Data also indicated that the fetal development stage may be a critical window of susceptibility to BPA exposure.
Van Esteri k et al., 2014	C57bl6 mice	In diet (fed a BPA- containing CD)	3, 10, 30, 100, 300, 1000, 3000 µg/kg/d During gestation and lactation	(4 females /group); groups were made of 8 mice per sex (4- 10)	in males: increased body size (not fat mass) with altered energy balance (glucagon increase) (but overrepresentation of small litters)	in females: no effect, so the authors decided to give a High Fat diet to all mice by week 17 and for 6 weeks. It resulted in decreased bw, leptin levels, fatty acids and triglycerides consistent with increased locomotor activity and of Ucp1 in the BAT	the authors conclude "Although these results suggest that BPA can program for an altered metabolic phenotype, the sexual dimorphism of effects and diversity of outcomes do not mark BPA as a specific obesogen. The consistency within the complex of observed metabolic effects suggests that upstream key element(s) in energy homeostasis are modified. Sex-dependent factors contribute to the final phenotypic outcome. »	conclusions consistent with the data

Garci a- Aréva lo et al., 2014	Pregnant OF1- mice	SC	Vehicle or BPA 10 μg/kg/d From E9 to E16	n= 8 mice/group 4 groups: males exposed to CD and CD-BPA or to HFD and HFD-BPA during 13 or 24 weeks	Effects of BPA on weight, food intake, gonadal and retroperitoneal fat pad weight and plasma NEFA  At birth: - ↓ bw in BPA rats - slight bw in BPA mice compared to CD  17 weeks: HFD-BPA ↓ perigonadal and retroperitoneal fat pad weight compared to HFD  28 weeks: CD-BPA ⊅ perigonadal fat pad weight compared ton control  No effect on food consumption in ND groups but ≯ in HFD-BPA compared to HFD  Glucose tolerance and insulin sensitivity Fasting hyperglycemia, glucose intolerance and high levels of NEFA in CD-BPA, HFD and HFD-BPA mice compared to control  no effect in ipITT	Effect of BPA on glucose stimulated secretion (GSIS) and insulin content  Disruption of glucose stimulated insulin release, particularly in HFD-BPA group  At 17 weeks: ✓islet insulin content in BPA and HFD −BPA mice compared to control but not at 28 weeks  Effects of BPA on gene expression in white adipose tissue, liver and skeletal muscle  - ↓mRNA expression of genes involved in fatty acid metabolism in white adipose tissue, comparable to HFD; -upregulation of Ppary and Prkaa1 genes in the liver;  - ↓expression of Cd36	Impaired tolerance to glucose in BPA treated mice compared to control. The phenotype of the BPA group resembles that of HFD mice.  Male offspring from BPA-treated mothers presented a form of diabetes which typically develops in later life and is associated with obesity	The experiment is well conducted. Only males but not females were studied. Mechanisms of action are not described nor did the authors used a positive control or inhibitor to explore the possible involvement of estrogeno-mimetic actions of BPA.
Moon et al., 2015	Growing male C57 BL/6 mice	Oral during 12 week High Fat Diet (HFD)	BPA 50 μg/kg/d	4 groups: CD with or without BPA and HFD with or without BPA: CD; CD BPA HFD; HFD BPA n= 5 per group and experiments repeated 3 times	BW, % of WAT and % of body fat did not differ between BPA and control group.  **glucose intolerance in HFD BPA mice (IpGTT)	Long-term exposure to BPA impairs insulin signaling: ↓Akt and GSK3β phosphorylation in skeletal muscle from BPA mice but not in hepatic of adipose tissue  No changes in islet area or morphology or insulin content of β-cells.	Long-term oral exposure to BPA along with HFD for 12 weeks induced glucose intolerance and insulin resistance in growing male mice.	The study is well conducted. The effects are mostly subtle with the exception of the effects describing impairment of insulin signaling in the skeletal muscle.
Alons o-	OF-1 mice	sc	Pregnant mice:	n= 10-30/ group depending on the	BPA 10 and 100 in BPA <b>pregnant</b> mice:	BPA 10 and 100 in BPA treated in non pregnant mice:	Exposure to low doses of BPA during pregnancy	The study is well conducted with a high

Magd alena et al., 2015			Vehicle (control<0) or BPA, 10 (BPA10) and 100 µg/kg/d (BPA100) From E9 to E16 Nonpregnant mice:	tests and ages	PND 3 months: no ≠ in glucose homeostasie (no effect on ipGTT) – no effect on ipITT PND 4 months: effects on glycemia homeostasie, slight effect on ipITT in BPA 100 PND 5 months: ≠effects on glycemia homeostasie; effect on ipITT in BPA 100 PND 6 months: strong effects on glycemia homeostasie; effect on ipITT in BPA 10 and 100 ★ BW and perigonadal fat pad weight PND 7 months: ↓ plasma insulin levels in BPA10 and BPA 100 compared to control ←↓ glucose – stimulated insulin secretion ↓ β-cells mass in BPA 10 and BPA 100 ←↓ proliferation; ↓ expression of Ccnd2 (in BPA100) but no effect on Cdk4 ★ expression of p16, no ≠in p53 gene expression, ↓ expression of cyclin D2 and CDK-4 proteins in BPA10 mice ★ expression of p16 and p53 proteins	PND 3, 4, 5 or months: no effect on ipGTT – no effect on ipITT  No change in glucosestimulated insulin secretion  Oxidative stress	- is associated with maternal alterations of glucose homeostasis and insulin sensitivity in the long term (glucose intolerance and insulin resistance) - disrupts pancreatic β-cells function 7 months after delivery - has impact on β-cell mass, proliferation and cell death	number of animals and long expertise on the domain. The study is highly indicative that BPA could be considered as pancreas toxic during pregnancy
Mogh adda m et al., 2015	Adult Male mice	ip.	Control and BPA 0.5 and 2 mg/kg 4 weeks	n= 6/group	BW with BPA     blood glucose with BPA dose dependently     *triglycerides, total cholesterol, LDL-C and ↓ HDL-C with BPA dose dependently	Oxidative stress parameters: - serum: ≯malondialdehyde  ↓ GSH with BPA dose dependently - pancreas: ≯malondialdehyde  ↓ GSH, TAS and SOD/CAT activities with BPA dose dependently No effect in proteins content	These results suggest that BPA exposure might induce hyperglycemia and its complications in adult male mice by induction of oxidative stress	Strain of mice not specified; composition of food not given; some authors not referenced.

Garci a- Aréva Io et al., 2016	OF-1 mice	SC	Vehicle (control <0); E2 10 μg/kg·d (control >0) or BPA, 10 (BPA10) and 100 μg/kg/d (BPA100), From E9 to E16	Control: n =73; BPA 10: n= 63; BPA 100: n= 56; E2: n= 18 (10 control + 8 treated)	↓ insulin secretion at P30 in the male offspring exposed to BPA10		Transcripton differential exp genes related apoptosis		modifications of the beta cell mass in the offspring as a consequence of estrogen signaling mechanisms initiated in fetal life at a wrong timing and leading to an excess of insulin signaling during early life which may contribute to impaired glucose tolerance during adulthood	This study is of good quality with a high number of animals. It was performed in a group with well recognised expertise in the pancreas and BPA.
White head et al., 2016	mice	Diet	BPA: 25 mg/kg/d E7.5 to E18.5		At E18.5 :  number of islet (ICCs) in fetal pa		Immunohisto analysis: BPA expression and glucagon-expr	✓ glucagon	BPA promotes islet differentiation or delays conversion of ICCs into mature cells , indicating alterations in glucagon expression in islets and ICCs	These data indicate that BPA can alter the differentiation program of the pancreas.
Veiga - Lopez et al., 2016	Female sheep		0.05, 0.5, 5 mg/kg/d D30-D90 (term 147d)	(groups 6-9)	study 1: metabolic tests in pre and post-pubertal F1 17 months; no effect in prepubertal sheep but intolerance to glucose and reduced insulin sensitivity in post-pubertal F1		study 2: overf weeks till 19 n glucose intole insulin resistar adipocytes are response to BI HFD but no int	nonths; HFD: rance and nce; BPA: hypertrophic in PA and to the	the authors conclude that exposure to BPA during fetal life at levels found in humans can program metabolic outcomes that lead to insulin resistance, a forerunner of type 2 diabetes, with postnatal obesity failing to manifest any interaction with prenatal BPA relative to insulin resistance and adipocyte hypertrophy.	the study is of good quality and the conclusions of the authors are fully consistent with the data yielded
Yang et al., 2016	C57 bl6 mice (5- week old)	In diet fed for 30 days with CD or HFD containing	50, 500, 5000, 50000 μg/d	(9-12/group)	10% increases in bw of both males and females; no dose-effect	increases of fat mass, inguinal WAT and epididymal WAT with higher number of large	almost no effect is seen in the groups of mice fed the high-fat diet	in vitro data and a human study complete the publication	The authors demonstrated the non monotonic dose effects of BPA on adiposity and chronic inflammation in 5-week-old mice	study of good quality

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		BPA so that exposure is of 50, 500, 5000, 50000 µg/d				adipocytes vs small adipocytes; no effect on GTT; increases of C/EBPa, PPARy, FABP4, Scd1, Srebp1c (dpdt on doses) in WAT				
Biasio tto et al., 2016	Pregnant C57bl6 mice	Gavage (daily)	doses: 0.5, 5, 5, 500 µg/kg/day from the 2nd wk of pregnancy; continued until 140 pnd except from birth to weaning	Groups ranged from 16 to 19.	Mice were fed a sinumber of pregnagiven but it can baround 5-6.	int females is not	Fat mass and BW enhanced starting from day 90 in mice dosed with BPA 5 microg/kg/da y probably linked to the enhancemen t in the epididymal weight.	little although significant increases of Pparg, ATGL, HSL and LPL in epididymal fat and liver	This study encompasses different protocols which together provide evidence that BPA acts as an ED acting through a nuclear receptor mediated mechanism	Study of good quality. Only males were studied.

Table 26: Summary of the available recent in vitro studies on bisphenol A on insulin

Ref.	Model	ВРА	Outcome s: TG	Glucose	GLUT	FABP4	Adipon ectin /	Possib	le MoA (	MOA)		Insulin sensitivity (IS)	Comments of the authors	Expert's comments
	3T3-L1 cells or MSCs	doses	accumul ation	uptake	4	(aP2)	leptin	ER	Pparg	GR	РІЗК			
Biemann <i>et</i> <i>al.,</i> 2012	mesenc hymal stem cells	10 μΜ	if treatment covered the proliferati on phase	no effect added at during the process	confluen	ce or							BPA is an EDC interfering with estrogenic action and reducing the commitment of MSCs into adipocytes	the study is of good quality
Pereira- Fernandes et al., 2013 2014,	3T3-L1 cells	12.5, 25, 50 μΜ	enhanced (x1.5) and x2 if combined to insulin versus x5 with ROSI 1 microM						very weak activit y versus ROSI 1µM				BPA has a very weak PPARg activity	the study is convincing
Valentino et al., 2013	3T3-L1 cells	1 and 10 nM		¥	not ⊅		ゝ of Leptin				→ (WB Phosp ho)	∡ IS	∖ of IS	BPA is reducing insulin sensitivity
Atlas <i>et al.,</i> 2014	3T3-L1 cells	0.1 nM, 1 nM, 10 nM	enhanced in the absence of DEX (presence of Ins)			dose- dpdt /			Not directl y				BPA can interact with the transcriptional machinery at the promoter of FABP4	BPA is astimulating FABP4 but the weak activation of Pparg by BPA cannot explain the FABP4 increase
Hélies- Toussaint et al., 2014	3T3-L1 cells	1 fM, 1 pM, 1 nM, 1 μΜ	✓ with 1fM, 1pM, 1nM	no effect	not ⊅ but Glut1 is increa sed	1pM / srebp1c, pparg, aP2	no effect on leptin	ERRa (not ERRg) / at 1 pM, 1nM					BPA could activate adipocyte differentiation through binding to ERRa or ERRg.	It is not possible to determine if BPA modulated insulin sensitivity ( no effect on glucose uptake).
Ariemma et al., 2016	3T3-L1 cells	1 nM added during all		`	not ⊅	7	no change in adipoQ;		7	GR not regard ed; ↗ of	→ (WB Phosp ho)	∖ of IS	Glucose utilisation and insulin signaling were	BPA is reducing insulin sensitivity. However,

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		phases				increase of leptin			C/EBP a			reduced. All data converge to show adipocyte metabolic dysfunction and inflammation; and decreased insulin sensitivity in 3T3-L1 cells.	inconsistency with regards to leptin (same lab as in Valentino 2013) .
Biasiotto et al., 2016	3T3-L1 cells either cultured with BPA in basal medium (BM) or BPA + MDI	10, 50, 80 μM	⊅ at 50 and 80 μM	z ii	n MDI		⊅ ERa in BM, Erb in MDI (ICI 182,78 0)	⊅ in MDI (TO)				specific activation of ERa in undifferentiate d cells and ERβ in differentiated cells. BPA also activated PPARγ; opposite action of DES	not classical in vitro protocol. Initial plating of cells is very low with 103 cells per 12-well culture plate (instead of 50,000 cells).
Dai <i>et al.,</i> 2016	3T3-L1 cells	80 µM	Socs3 expressio n (socs3 contribut es to IR and leptin R							√ (WB Phosp ho)	∿ of IS	→ of IS	of IS as can be concluded from the reading of the abstract (only the abstract is available)
Ahmed, and Atlas, 2016	3T3-L1 cells	0.01 to 25 μM	z lipid accumula tion (2.5>25 μΜ)	,				⊅; further ⊅with ROSI				BPA requires PPARy to induce adipogenesis	the use of GW9662 to demonstrate the involvement of PPARg in BPA induced effects is convincing (RT- qPCR and Western-blot analysis)

## 4.6 Summary and discussion of human health hazard assessment

This dossier has summarised the available evidence showing that BPA can affect a number of physiological functions and systems of a mammal organism through ED pathways. More specifically, BPA alters the reproductive function, mammary gland development, cognitive functions and metabolism through an ED MoA.

Most importantly, although the steps of the respective mechanisms of action are specific for each effect, the disruption of the estrogenic pathways is a common MoA consistently involved in each of the four effects. The primary target of BPA is however still not known with certainty. BPA binds to estrogen receptors (ER) but with a weak affinity. In addition, BPA binds also to other types of ER such as GPER or ERR- $\gamma$  with a higher affinity and then these receptors may also be involved in BPA-mediated effects, particularly at low doses. The complexity of the toxic response to BPA also suggests multiple MoA that may interact. However, some evidence detailed in each section of this report for each effect specifically enables to establish that the estrogenic pathway is central and common in the MoA. Other findings also point toward alternative elements involved in BPA-mediated effects, e.g. epigenetic modifications. It is not known whether these modifications may be estrogen-dependent but the potential role of additional mechanistic pathways does not contradict the importance of the estrogenic pathways in the BPA-mediated effects.

Estrogens are known to be central in the regulation of the sexual function and system but are also known to interact with many other physiological functions and developmental processes including neurobehaviour or metabolism. The pattern of the effects observed with BPA is therefore consistent with an ED MoA through estrogenic pathways.

There is recent emerging evidence that BPA may have immunotoxic effects (Ménard *et al.*, 2014a and 2014b). The variability of the effects makes the interpretation and the transposition of these effects to humans uncertain. It is however noted that the role of estrogens has been often reported in immunocompetence and in the development of innate and adaptive immune response (Fish *et al.*, 2008). It therefore cannot be excluded that the range of effects related to the ED-properties of BPA may be wider than those described in this dossier.

Effects on environment are not addressed in this report. However, it is of importance to note that estrogen-related modes of action are reported as one of the predominant MoA potentially involved in the reproductive disturbance reported in several taxonomic groups. In particular, some data suggest an estrogen-agonist MoA in fishes (severe effect on reproduction affecting population (Yokota *et al.*, 2000) and vitellogenin induction (Kashiwada *et al.*, 2002)), in amphibians (skewed sex ratio towards females (Levy *et al.*, 2004) and vitellogenin induction (Oehlmann *et al.*, 2009)) and in molluscs (stimulatory effect of BPA on egg production antagonised by anti-estrogen (Oehlmann *et al.*, 2006)) and possibly an estrogen-like action in echinoderms (abnormal larvae development (Roepke *et al.* 2005)).

Altogether, there is consistent evidence showing that BPA affects the reproductive function, mammary gland development, cognitive functions and metabolism and these alterations are mediated through disruption of estrogens and estrogenic pathways. These effects are considered serious health effects. Although they may be exerted through direct exposure (alteration of estrous cycles and of memory/learning performance), they are also observed for all four endpoints after developmental exposure with consequences later in the life of offspring.

# 5. Environmental hazard assessment

Not assessed for this dossier addressing the identification of the substance as SVHC as an endocrine disruptor for human health in accordance with Article 57 (f) of REACH.

## 6. Conclusions on the SVHC Properties

#### 6.1 CMR assessment

Not relevant for the identification of the substance as SVHC in accordance with Article 57 (f) of REACH.

It is however noted that BPA is included in the Candidate List as a SVHC under Article 57 (c) by decision of ECHA ED/01/2017 of the 4 January 2017.

Indeed, Bisphenol A is covered by index number 604-030-00-0 of Regulation (EC) No 1272/2008 in Annex VI, part 3, Table 3.1 (the list of harmonised classification and labelling of hazardous substances) and it is classified in the hazard class reproductive toxicity category 1B (H360F 'May damage fertility') since its 9<sup>th</sup> Adaptation to Technical Progress (ATP) (Commission Regulation (EU) 2016/1179).

#### 6.2 PBT and vPvB assessment

Not relevant for the identification of the substance as SVHC in accordance with Article 57 (f) of REACH.

## **6.3 Assessment under Article 57(f)**

## 6.3.1 Summary of the data on the hazardous properties

This dossier has summarised the available evidence that BPA can affect a number of physiological functions and systems of mammalian organisms through an endocrine disruptive pathway. In particular, BPA can alter reproductive function, mammary gland development, cognitive function and metabolism through ED MoA.

#### Alteration of estrous cycles

In humans, one epidemiological study investigated the link between BPA and the characteristics of menstrual cycles. An association with shorter luteal phases was observed. However, no definitive conclusion in humans can be drawn on the basis of a single study (Jukic *et al.*, 2015). In animals, although there was some disparities between studies, reliable results were repeatedly reported in several experimental studies in rats and mice showing an adverse effect of BPA on the estrous cycle, including irregular and prolonged cycles and an alteration of estrous cycles dynamic by BPA after different periods of exposure:

- after exposure of adult females (Tyl et al., 2008; Laws et al., 2000; Lee et al., 2013) and
- after exposure during the developmental phase of the reproductive system, i.e. in utero (Honma et al., 2002; Nikaido et al., 2004; Wang et al., 2014a), perinatal (Rubin et al., 2001; Mendoza et al., 2011; Patisaul et al., 2014; Delclos et al., 2014), postnatal (Nah et al., 2011; Adewale et al., 2009; Fernandez et al., 2009) or prepubertal exposure (Zaid et al., 2014)

This effect was recognised by RAC in its opinion in support of classification of BPA as Repr 1B – H360F as summarised in the RAC opinion on restriction of BPA (ECHA, 2015):

"RAC's opinion (RAC, 2014) was based on adverse effects, such as disturbances in the estrous cycle, at a dose of 600 mg/kg bw/day (Tyl et al., 2008) and at a dose of 100 mg/kg bw/day (Delclos et al., 2014)."

Proper cyclicity is considered essential to reach successful ovulation. An alteration of cyclicity may therefore directly induce at least subfertility through disturbed (delayed or absent) ovulation. As synthetised by Kortenkamp *et al.*, (2012) an association between menstrual cycle characteristics (e.g. length of the cycle, duration of menstrual bleeding), sub-fecundity and spontaneous abortion has been observed in humans and menstrual characteristics have been associated with chronic diseases, including breast and ovarian cancer, uterine fibroids, diabetes and cardiovascular disease. Alteration of cyclicity is therefore considered as an effect clearly fulfilling the criteria of adversity.

In relation to alteration of cycle following adult exposure, a negative effect of BPA on ovarian estrogen production is clearly demonstrated in rodents in vivo (Lee et al., 2013) in accordance with the effects observed on rodent and human ovarian cells in vitro (Kwintkiewicz et al., 2010; Mansur et al., 2016). Whereas the effect of BPA on thecainterstitial cells depends on the test model and the protocol, BPA constantly reduces the conversion of androgens into estrogens in granulosa cells. This reduction is, at least in part, a consequence of a decreased transcription of CYP450arom either via a direct effect in the granulosa cells or via changes in intrafollicular signaling factors that regulate follicular growth and endocrine activity. In both primates and non-primate mammals, follicle selection, growth, and maturation, as well as ovulation, oocyte quality, and subsequent corpus luteum function, all depend on subtle sequential actions of gonadotropins and intra-ovarian regulators. Furthermore, the ovary and the hypothalamo-pituitary system are in permanent endocrine dialogue with each other. Consequently, any disturbance in the cycle implies some disturbances in the endo/para/autocrine activities of the ovary and/or the hypothalamo-pituitary system. Given the regulatory scheme of the estrous cycle, an alteration in the preovulatory follicle steroidogenic activity - as observed with BPA - is very likely to be associated with a disruption of the estrous cycle as shown in the Lee et al. study (2013). These results demonstrate a clear endocrine MoA, namely the alteration of the ovarian steroidogenic activity, underlying estrous cycle disruption in adult rodents.

Although most of the reported evidence relies on rodent studies, there are *in vitro* data showing the same negative effect of BPA on the estrogen production in the human follicle cells. Furthermore, an indication of a negative association between the ability of the follicle to produce estrogens and exposure to BPA was observed in women (Mok-Lin *et al.*, 2010; Ehrlich *et al.*, 2012). Lastly, the role of estrogens in the maintenance of the cycle is similar in rodents and humans. Thus, we conclude that it is likely that BPA may alter the ovarian cycle in humans through the disruption of the endocrine activity of the ovarian follicle.

Alteration of cycle further to developmental exposure to BPA through endocrine or neuroendocrine mechanisms is less clearly demonstrated than after adult exposure. Nevertheless, many studies show that the basic (neuro) endocrine mechanisms underlying the finely tuned regulation of the gonadotropic function underlying the estrous cycle can be altered in response to exposure to BPA, after developmental exposure. BPA has been shown to affect the hypothalamic expression of kisspeptin, a key neuropeptide in the regulation of the hypothalamic-pituitary-gonad (HPG) axis that is essential to later achieve the release of hormones at the appropriate time and concentrations during the cycle. In particular, studies by Monje *et al.*, (2010) and Fernandez *et al.*, (2009) provide a link between neuroendocrine changes and alteration of the cycles through concomitant observation of an alteration of hormones of the HPG axis and a cycle disturbance. The affected targets are similar to a large extent to targets affected by either estrogen agonist or estrogenic positive controls. Overall, animal and *in* 

*vitro* data support the hypothesis of an endocrine-relative MoA of BPA to induce perturbation of estrous cyclicity after developmental exposure.

In humans, the role of kisspeptin in the neuroendocrine control of the HPG axis has been demonstrated. In addition, the recent study by Kurian *et al.* (2015) on mid to late pubertal ovarian intact female rhesus monkeys suggests that persistent exposures to BPA could impair the female reproductive function by directly influencing the hypothalamic neuroendocrine function as evidenced by an alteration of kisspeptin release and GnRH pulsatility. Therefore, it can be considered that BPA-induced alterations of the hypothalamic kisspeptin/GnRH system are also relevant in humans and may impact estrous cyclicity long time after exposure in an irreversible manner.

# • Alteration of mammary gland development leading to an increased susceptibility to carcinogens through morphological modifications

Very few epidemiological studies on risk of breast cancer after exposure to BPA early in life are available. Based on those studies, which were mainly case-control or cross-sectional studies, a link between exposure to BPA and human breast cancer could not be determined (Aschengrau *et al.*, 1998, Yang *et al.*, 2009, Sprague *et al.*, 2013 and Trabert *et al.*, 2014). It should be noted that the existing studies evaluate exposure to BPA on the date of the study whereas the effects looked at are long-term effects appearing several decades after exposure. Therefore, a more biologically relevant study design would be a longitudinal epidemiological study measuring BPA *in utero* as breast cancer most likely takes years to develop.

The mammary gland develops in three distinctive life stages: fetal, peri-pubertal, and pregnancy (Fenton, 2006) with hormonal implication such as mammary epithelial estradiol signaling, progesterone (PR) and prolactin (PL) receptor involvement depending on the developmental period. Moreover, mammary epithelium responds differently to hormonal stimulus depending on its developmental stage.

In terms of hormonal regulation, there seems to be substantial similarities across species. In most mammals, the ovaries first secrete estrogens in response to increased secretion of gonadotropins, and sexual maturity coincides with the establishment of cyclic peaks of ovarian progesterone secretion. Proliferative activity is observed in the human breast epithelium (Masters *et al.*, 1977; Longacre and Bartow, 1986) concomitantly with the peak of progesterone during the luteal phase. Thus rodents and human mammary epithelia may indeed be similarly regulated, at least with regards to hormonal control of cell proliferation.

Although the anatomy of the human breast is more complex than the rodent mammary gland, some human breast structures (Terminal Ductal Lobular Unit or TDLU) are structurally similar to rodent structures (Terminal End Bud or TEBs) during the same life stages. These structures are undifferentiated and highly proliferative, and as such they are sensitive to the effects of carcinogens and other chemicals.

There is evidence from rodents and non-human primate studies that prenatal and postnatal exposure to BPA causes endocrine modifications in the mammary tissue, ultimately increasing its susceptibility to chemical carcinogens, as previously reported (ANSES, 2013b and review by Soto *et al.*, 2013).

Based on BPA data analysis, there is evidence from rodent and non-human primate studies that prenatal and post-natal exposure to BPA causes, depending on the period of exposure, modifications in the mammary tissue such as an increased number of TEBs relative to the ductal area, a fewer apoptotic TEB cells, an increased lateral branching and ductal hyperplasia, an increased cell proliferation and a decreased apoptosis in the glandular epithelium, a ductal (and occasionally lobuloalveolar) and an intraductal

hyperplasia. This ultimately increases its susceptibility to chemical carcinogens, as previously been reported (ANSES, 2013b and review from Soto *et al.*, 2013) and the number of DCIS in rats treated with BPA (Durando *et al.*, 2007, Jenkins *et al.*, 2009 and 2012, Betancourt *et al.*, 2010). Delclos *et al.* (2014) observed ductal hyperplasia accompanied with a very limited number of mammary adenocarcinoma. Overall, based on all available *in vivo* studies, the structural similarity between humans and rodents, and some common hormonal controls of cell proliferation, suggested there is substantial evidence from rodent studies that early-life exposures to BPA lead to increased susceptibility to breast cancer in humans.

In vitro experiments conducted on normal-like (MCF-10F or MCF-12A) breast cells show an increased cell proliferation due to exposure to BPA and the engagement of ERa and its co-activator (Sengupta et al., 2013). In breast cancer cells and cancer-associated fibroblasts (CAF) that lack the classical ER, it was shown that G protein-coupled estrogen receptor (GPER) is required for growth effects and migration (Pupo et al., 2012). In vitro 3D models for breast glandular structure development, using non-transformed breast epithelial MCF-10F and MCF-12A cells, indicated that BPA may alter the ductular and alveolar patterns in the collagen matrix, with a similar pattern as E2 (Fernandez and Russo, 2010). When BPA or  $17\beta$ -estradiol treatment was combined with ER and GPER inhibitors (ICI 182 780 and G15, respectively), the effects (deformed acini) were reversed suggesting a role of ER and GPER in the estrogenic disruption of acinar formation operated by BPA. As GPER is required for growth effects and migration in cancer (SKBR3) cells and in CAFs and since proliferative effects induced by BPA were cancelled when GPER expression was silenced by shGPER, it can be concluded that BPA induces stimulatory effects as a GPER agonist in these breast cancer cells and CAFs.

Some studies using transcriptional analyses on the stromal and epithelial compartments isolated from the fetal mammary gland (mouse) demonstrate that exposure to BPA in dams alters the mesenchymal and epithelial transcriptomes related to proteins involved in apoptosis (increased expression of the anti-apoptotic gene, Birc2, Abl1), myoepithelial differentiation, changes in the composition of ECM and in adipogenesis (Wadia *et al.*, 2013).

A series of studies from the same scientific group, Mandal's team (Bhan *et al.*, 2013, 2014a and 2014b) described how BPA can alter the epigenetic programming of the promoter of HOTAIR and EZH2. The expression of these genes has been shown to be EE2 dependent. In addition, these genes have been described as being involved when cell proliferation increases, during increased invasiveness seen in some breast tumors and to contribute to breast cancer progression.

Other studies support the possibility that BPA, through interaction with the nuclear ERs (Murray et al., 2007; Wadia et al., 2013) or GPER and indirectly with PR (Munoz de Toro et al., 2005; Jenkins et al., 2009; Ayyanan et al., 2011) modulates estrogenic and progestin agonist activities. Furthermore, exposure to BPA during fetal life provokes an increased expression of both RankL (a critical connection between progesterone and epithelial cell proliferation) and Wnt4 (involved in progesterone-induced side branching in early adult life) at adult stage (see Jenkins et al., 2009 and Ayyanan et al., 2011, respectively).

In conclusion, available data support the plausibility that BPA, through interaction with the nuclear ERs, or GPER and indirectly with PR, modulates estrogenic and progestin agonist activities. Emerging epigenetic studies have reported changes related to estrogen-dependent genes (such as EZH2 and HOTAIR), as well as HOX genes (involved in embryogenesis and post-natal development) associated with the BPA induced abnormal development and cancer increased susceptibility of mammary gland.

## Alteration of learning and memory performances

Epidemiological evidence on the potential role of exposure to BPA early in life on learning and memory performances is still insufficient to conclude. However, a substantial majority of rodent studies (total of 26/35) reported impaired spatial and non-spatial memory following exposure to BPA, regardless of the period of exposure. Nonetheless, the overall evidence has recently substantially increased and altogether strongly points toward the conclusion that BPA alters memory in rodents. In addition, as the 74% of studies reporting impaired cognitive behaviours were also performed under various experimental conditions (various doses, routes and periods of exposure or tested species), this means that this impairment is a robust BPA-induced effect. Another interesting finding, which can be drawn from this analysis, is the sex-dependent effect observed in several studies. Furthermore, although not systematically assessed in all these studies, the neural mechanisms associated with the behavioural alterations consist of a reduction in the level of expression of NMDAR subunits, kinases, enzymes involved in neurotransmitter regulation, and synaptic proteins as well as decreased spine density or neurogenesis. Such molecular, cellular and structural changes are fully relevant and could underlie the impaired learning and memory performances observed in the same animals.

Finally, although there is a limited number of studies conducted in non-human primates, these studies have shown that BPA during the prenatal stage of development has detrimental effects on the midbrain dopaminergic system and on spine synapses in the hippocampus, while it has no effect when applied at a juvenile stage (Elsworth *et al.*, 2013). In addition, adult BPA-exposed monkeys displayed a significant cognitive impairment (Elsworth *et al.*, 2015). It is also interesting to note that down-regulation of estradiol-induced increase in spine synapses in the hippocampus and prefrontal cortex in adult ovariectomised monkeys exposed to BPA was also reported in another study of the same laboratory (Leranth *et al.*, 2008).

Overall, BPA has been demonstrated to alter memory and learning after developmental, pubertal or adult exposure, based on multiple converging experimental studies reporting this functional effect as well as molecular and cellular changes in the brain in line with the functional changes observed pointing to a clear and coherent adverse effect.

The possibility that BPA alters the cellular and molecular pathways involved in learning and memory processes through disruption of estrogen-dependent pathways was first suggested in the study of Xu et al. (2010a and 2010b) reporting decreased expression of ERβ. In the study of Xu et al. (2014b), the link between BPA-induced effects on learning and memory processes and estrogenic pathway disruption was clearly established from the demonstration that the ER antagonist ICI 182,780 reversed both BPA-induced effects on ERα (modulation and regulation) and memory. Three other studies performed in adults indicated that BPA is also able to interfere with estradiol-induced effects on behaviour and spine density in rodents (Xu et al., 2015b; Inagaki et al., 2012) and on synaptogenesis in nonhuman primates (Leranth et al., 2008). Additional evidence was provided by in vitro studies showing that BPA-induced effects on NMDAR signalling and synaptic proteins were reversed by the ER antagonist. In one in vitro study, BPA-induced disruption extended to other non-classical estrogen receptors (ERRγ).

The modulatory effects of estrogens on cognitive processes and behaviour in adults are now well established. Although they were studied more extensively in females (Galea *et al.*, 2013; Pawluski *et al.*, 2009), the importance of the estrogenic pathway in the regulation of cognitive behaviour and synaptic plasticity has also been reported in male rodents (Picot *et al.*, 2016). Detailed effects of adult estrogens on learning and memory and the mechanisms underlying these effects in both males and females are described in

recent reviews (Frick *et al.*, 2015; Hamson *et al.*, 2016). Sex differences in hormonal impregnation during the critical periods of development and their influence on cognitive performance have been described by Roof and Havens (1992). This may explain the sex differences observed in the expression of cognitive behaviours and their alteration by BPA.

Altogether, these data highly support the plausibility that alteration of learning and memory by BPA is mediated by disturbance of the estrogenic pathways. Further details presented in the dossier also show that other steroid pathways might be involved.

Cognitive function in humans involves signaling pathways, which seem similar to those described above in rodents. The involvement of NMDAR signaling pathway in memory processes in healthy and diseased brain has been largely reviewed (e.g. in Gilmour *et al.*, 2012; Campos *et al.*, 2016; Arnsten *et al.*, 2016). Estrogens were shown to modulate hippocampus-dependent learning in women and non-human primates (Hampson, 1990; Lacreuse, 2006 and review of Hamson *et al.*, 2016). Testosterone also modulates cognitive functions in men. Given these similarities in the modulatory effects of sex steroids on cognitive functions between rodents and humans, it is likely that the MoA observed in rodents occurs in humans and affects similarly these processes in humans as in rodents. In support of this hypothesis, the *in vivo* study conducted on adult female non human primates showed that a subcutaneous exposure to BPA (50  $\mu$ g/kg/d) counteracts the synaptogenic effect of estradiol in the hippocampus and prefrontal cortex (Leranth *et al.*, 2008).

Based on i) the significant amount of *in vivo* and *in vitro* animal data showing impairment of learning and memory following exposure to BPA and the potential alteration of cellular and molecular mechanisms underlying these processes through disturbance of the estrogenic pathway, ii) the similar types of signaling pathways underlying human cognition and iii) the numerous data showing sex steroid regulation of these behaviors, exposure to BPA could also alter human cognitive abilities through disturbance of estrogenic pathways.

### · Effects on metabolism

Based on animal studies after prenatal and/or perinatal or adult exposure, there is now evidence that BPA may increase the incidence of type-2 diabetes via an ED MoA. In particular, BPA has been shown to alter insulin synthesis and/or release by  $\beta$ -pancreatic cells, or insulin signalisation (signaling mechanisms) within insulin-sensitive organs (i.e., liver, muscle, adipose tissues). This resulted in variations in the expression of hepatic or adipose tissue markers which are indicative of a state of insulin resistance. These effects are considered by experts to be hallmarks of hormonal adverse effects, especially if there is a combination of effects, each leading to insulin resistance within the different insulin-sensitive tissues. In addition, while most studies were performed on males, a few studies have also examined the impact of BPA on either both sexes or only females. However, more studies should be undertaken before concluding on a sex-specificity or not of the metabolic impact of BPA on insulin synthesis/release and signalisation.

Recent experimental *in vivo* and *in vitro* studies indicate that these effects may involve ERa, ER $\beta$  or GPR30 pathways. Other hormones such as leptin or adiponectin, involved in resistance to insulin and lipogenesis, are also modified following BPA exposure. This shows that BPA could interfere in the balanced interplay between insulin secretion and insulin action that controls glycaemia.

Overall, it is suggested that the endocrine pancreas is targeted by BPA and that mechanisms could differ depending on whether exposure occurs during the fetal life or in adulthood. Fetal differentiation of the pancreas appears highly sensitive to exposure to BPA based on the outcomes surveyed, e.g.  $\beta$ -cell proliferation and apoptosis. Limited

data exist on the impact of BPA on a-cells and glucagon secretion. In addition, BPA can elicit histopathological modifications during the fetal life with consequences on insulin synthesis rate and/or release.

Moreover, most of the *in vitro* studies showing adverse effects of BPA on adipocyte differentiation and function indicate alteration of endocrine mechanisms (e.g., adiponectin release, insulin signaling cascade effectors). However, uncertainties remain as whether BPA activates PPARy and/or other nuclear receptors (possible cross-talk between nuclear receptors).

Even if available, epidemiological studies are inconclusive. These effects are considered relevant for humans because similarities exist in homeostatic regulation of insulin production and sensitivity between rodents (mostly investigated in the experimental studies) and humans and because of *in vitro* experimental data using human cells or tissues.

#### Overall conclusion on ED-mediated effects of BPA

This dossier displays extensive evidence showing that BPA can affect several physiological functions and systems of mammal organisms through ED pathways. As explained earlier, BPA alters the reproductive function, mammary gland development, cognitive functions and metabolism through an ED MoA.

Most importantly, although the steps of the respective mechanisms of action are specific for each effect, the disruption of the estrogenic pathways is a common MoA consistently involved in each of the four effects. The primary target of BPA is however still not known with certainty. BPA binds to estrogen receptors (ER) but with a weak affinity. In addition, BPA binds also to other types of ER such as GPER or ERR-γ with a higher affinity and then, these receptors may also be involved in BPA-mediated effects, particularly at low doses.

The complexity of the toxic response to BPA also suggests multiple MoA that may interact. However, some evidence detailed in each section of this report for each effect, specifically enables to establish that the estrogenic pathway is central and common in the MoA. Other findings also point toward alternative elements involved in BPA-mediated effects, e.g. epigenetic modifications. It is not known whether these modifications may be estrogen-dependent but the potential role of additional mechanistic pathways does not contradict the importance of the estrogenic pathways in the BPA-mediated effects.

Estrogens are known to be central in the regulation of the sexual function and system but are also known to interact with many other physiological functions and developmental processes including neurobehaviour or metabolism. The pattern of the effects observed with BPA is therefore consistent with an ED MoA through estrogenic pathways.

There is recent emerging evidence that BPA may have immunotoxic effects (Ménard *et al.*, 2014a and 2014b). The variability of the effects makes the interpretation and the transposition of these effects to humans uncertain. It is however noted that the role of estrogens has been often reported in immunocompetence and in the development of innate and adaptive immune response (Fish *et al.*, 2008). It therefore cannot be excluded that the range of effects related to the ED-properties of BPA may be wider than those described in this dossier.

Effects on environment are not addressed in this report. However, it is of importance to note that estrogen-related modes of action are reported as one of the predominant MoA potentially involved in the reproductive disturbance reported in several taxonomic

groups. In particular, some data suggest an estrogen-agonist MoA in fish (severe effect on reproduction affecting population (Yokota *et al.*, 2000) and vitellogenin induction (Kashiwada *et al.*, 2002)), in amphibians (skewed sex ratio towards females (Levy *et al.*, 2004) and vitellogenin induction (Oehlmann *et al.*, 2009)) and in molluscs (stimulatory effect of BPA on egg production antagonised by anti-estrogen (Oehlmann *et al.*, 2006)) and possibly an estrogen-like action in echinoderms (abnormal larvae development (Roepke *et al.* 2005)).

Altogether, there is consistent evidence showing that BPA affects the reproductive function, mammary gland development, cognitive functions and metabolism and these alterations are mediated through disruption of estrogens and estrogenic pathways. These effects are considered serious health effects. Although they may be exerted through direct exposure (alteration of estrous cycles and of memory/learning performance), they are also observed for all four endpoints after developmental exposure with consequences later in the life of the offspring.

## 6.3.2 Equivalent level of concern assessment

#### 6.3.2.1 Human health

There is currently no guidance on how to assess that an adverse health effect represents an equivalent level of concern (ELoC) to a CMR substance, thereby fulfilling criteria for SVHC identification according to article 57(f) of REACH for human health. A discussion paper of ECHA is however available (ECHA, 2012) with a specific focus on sensitisers. However, the criteria identified in this document to evaluate the EloC are considered fully relevant for the present case. These factors are listed below and are used in the present analysis:

- Health effects:
  - o Type of possible health effects
  - o Irreversibility of health effects
  - o Delay of health effects
- Other factors:
  - Quality of life affected
  - o Societal concern
  - o Is derivation of 'safe concentration' possible?

#### Type of effects

The type of effects of BPA is comparable to CMR substances that have the potential to induce serious and permanent organ dysfunction. This section focuses on whether the ED-related effects of BPA are serious effects. Persistency of the effect is addressed in the following section related to irreversibility.

In relation to its ED-mediated MoA BPA exerts a range of effects described in our dossier that are considered as **serious**:

- BPA can affect the proper regulation of the reproductive function and can be involved in alteration of fertility, as reflected by its harmonised classification Repr 1B. On this basis, it unequivocally fulfills the criteria of seriousness, as well as per se of ELoC compared to CMR.
- BPA properties altered the development and the structure of the mammary gland and BPA is considered to present a substantial risk factor for increasing the susceptibility in induction of breast cancer tumours. BPA has no harmonised classification for carcinogenicity but, its capacity to increase sensitivity to other breast carcinogens, by its proliferative action in relation to its specific ED MoA, is considered as a serious effect. The carcinogenic effect of BPA has been discussed

and recognised: "BPA may be reasonably anticipated to be a human carcinogen in the breast and prostate due to its tumor promoting properties" (Seachrist *et al.*, 2016). BPA has already a harmonised classification for a CMR endpoint. Therefore, this hazard has not been evaluated further and the possibility of proposing a harmonised classification for this endpoint judged too resource-consuming compared to the risk management added-value.

- BPA is also implicated in alteration of learning and memory as well as of brain histological structures which is also considered to be a serious effect. As summarised in Annex 2.2 of the recent JRC report (JRC, 2015) discussing ELoC for neurotoxicants (and immunotoxicants), all these changes are recognised as adverse changes by groups of experts on neurotoxicity within the WHO Environmental Health Criteria Programme and EFSA. They are also considered as serious effects eligible for a STOT RE classification according to CLP
- The effects of BPA on metabolism are associated with serious chronic pathologies such as type-2 diabetes after prenatal and/or perinatal or adult exposure in animals (rodents and non-rodents). The two major pathophysiologies of concern for diabetes mellitus (type 2) are a decreased insulin secretion and an increased insulin resistance. We have shown in this report that BPA exposure may act on both pathways. Diabetes and obesity are associated with serious co-morbidities and reduced life span. Moreover, BPA exposure during pregnancy induces an alteration of glucose homeostasis in dams, which is characterised by the development of severe glucose intolerance and aggravated insulin resistance, hyperinsulinemia and hyperleptinemia. This is a situation similar to that of gestational diabetes mellitus (GDM). GDM is associated with increased risks of birth complications (complications related to excessive birth weight, preterm birth and respiratory distress syndrome, hypoglycemia) and of type 2 diabetes later in life. The effect of BPA on metabolism is therefore considered as serious. In addition, there are no criteria on how to consider these effects are available for classification & labelling. Despite the seriousness of the effect, a CLH dossier for this endpoint is not foreseen.

Altogether, the ED MoA of BPA is unequivocally considered to raise concern in relation to multiple serious health effects.

## Irreversibility of the effects and delay of health effects

Some of the ED-related effects of BPA have been identified after concomitant exposure. However, these ED-related effects are also all characteristically observed after developmental exposure to BPA, with consequences that are observed later in life (estrous cycle, mammary gland, neurotoxicity, insulin synthesis and resistance), without a direct exposure. As the effects appear long time after the exposure, they are indeed considered as **permanent and irreversible**.

It is also noted that some studies although limited in number have reported transgenerational effects of BPA on cognitive function.

This delay between exposure and manifestation of the effects creates difficulties in detecting the link between exposure and effect. It raised a difficulty in the protocol design, more specifically in human epidemiological studies. An absence of observed effects does not mean necessarily an absence of effect. In this case, risk management measures and prevention of exposure may not be taken in time and individuals might be exposed for a long time before action is taken. It is especially applicable to developmental effects for which exposure occurs during the developmental period and they are therefore very difficult to characterise *a posteriori*. This relates to the concept of

developmental origins of health and disease (DOHaD). Moreover, the latency and irreversibility of the effects may also pose some ethical concern due to the fact that they may also affect future generations that might not be protected. Due to the difficulty posed by latency both in terms of detection of the effect and responsibilities for later generations, it is considered as an additional concern.

## Quality of life affected

Altogether, the adverse effects of BPA such as demonstrated in our dossier may concur to a **reduced quality of life**.

- The effects of BPA on the reproductive function encompass a wide range of manifestations that can significantly impact the quality of life. Puberty is a difficult period of life associated with significant psychological distress and a need for social identification. Modification of the onset of puberty can increase these difficulties. The disruption of ovarian cycles may occur triggering various outcomes in women: extension or shortening of the menstrual cycle, erratic period cycles, irregularity of menstruation flows, etc. These outcomes may have various and more or less serious impacts on everyday life such as abnormal bleeding (menstruation flow), disruption of fertility (due e.g. to fewer ovarian cycles and thus a lower probability of getting pregnant in the case of elongation of cycles), disruption of sexuality, discomfort and inconvenience, and generally a lower quality of life. This kind of disruption may occur from puberty to menopause. The magnitude of adverse impacts depends on the severity of the effects likely to appear, from a slight extension of ovarian cycles to complete amenorrhoea. Ultimately, potential infertility is associated with a major source of worries and psychological distress for the individuals affected. It may involve medical assistance that involves constraining and time demanding protocols, most often over a certain period of time to reach success, if any.
- The increased vulnerability of the mammary gland due to developmental and/or peri-post-natal exposure increases the risk of developing breast cancer at a later stage in life. The health impacts associated with the effect of BPA on mammary glands thus correspond to increased occurrence of breast cancers; the structural changes of mammary glands induced by BPA exposure being considered as precursors to breast cancer. Breast cancer is an uncontrolled growth of breast cells leading to a malignant tumor. Initially, breast cancer may not cause any symptoms. A lump in the breast may be too small to feel or to cause any unusual changes (swelling of all or part of the breast, breast pain, nipple turning inward, etc.). Breast cancer can be treated by surgery, medical therapies (radiotherapy, chemotherapy) and/or medication such as anti-estrogen, cytotoxic, and endocrine drugs. Medicines can also be prescribed to treat patients, in order to mitigate potentially severe side effects of medical therapies. Besides treatments, breast cancer might cause many other adverse consequences for patients such as absenteeism, social isolation, psychological depression, anxiety, and more generally a lower quality of life; the worst adverse effect being death. Subsequent to the breast cancer itself, additional physical and psychological suffering may be imposed by breast reconstructive surgery, if any.
- A decrease of cognitive functions can have an impact on daily life by impairing the ability of an individual to effectively and autonomously cope with daily tasks and difficulties. It may occur through many various forms such as disorientation, weak memory, disrupted learning capacities, etc. In children and adolescents it may also affect school performance and may contribute to situations of school failure that may impact childrens' self-esteem as well as compromise future educational and occupational achievement. It may also create a significant source of worries in parents. Emerging evidence of anxiety is also associated with

- possible impact on well-being and social abilities of these individuals. At a population level, this effect could lead to a global displacement of the Gaussian IQ distribution impacting the average IQ of the population.
- BPA exposure may enhance diabetes risk due to an increase in insulin resistance.
  Diabetes is a metabolic disease accompanied by important co-morbidities and
  reduced life span. In addition, it generally requires daily glycemic control as well
  as careful adherence to specific dietary habits that may in particular affect also
  social life. The quality of life may also be altered by symptoms of low or very high
  blood glucose and fears about potential or real complications.

#### Societal concern

The adverse effects of BPA such as demonstrated in our dossier all raise indisputable **societal concern**.

- Regarding reproduction, there is a general societal concern associated with possible conception difficulties. Infertility rates have remained stable (Kortenkamp et al., 2012) in the last decades ranging from 3.5 to 16.7% in developed countries (Boivin et al., 2007) but the demand for assisted reproductive technologies (ART) treatment in Europe - as expressed in treatment cycles performed in European countries - has increased by 59% in the five years from 1997 to 2002 (HEAL 2014). Although increasing age at the time of conception may be a preponderant factor to explain conception difficulties, a potential role of EDCs is generally considered as plausible as well (Marques-Pinto et al., 2013). In addition, there is an increasing concern related to disorders associated with the female reproductive system such as endometriosis and PCOS which might additionally affect fertility. Additionally to ethical and demographical concerns associated with conception difficulties, those adverse effects affecting fertility might be costly for the society as a whole. Indeed, recent literature provides evaluations of such costs although there is no specific estimate of the burden that may be potentially related to BPA. For example, the HEAL study (HEAL 2014) concluded that EDC-related male and female infertility direct cost associated with ART is between 48 and 155 million € per year in the EU28 as quoted by Rijk et al. (2016)<sup>18</sup>. Likewise, Norden et al. 2014 study estimated the direct (healthcare) and indirect (productivity losses) cost of EDCs-related male infertility cases associated with fertility treatments such as ICSI treatments to be about 72.3 million € for the EU28 (Norden, 2014).
- The risk factors for breast cancer are numerous and there is no specific estimate of the breast cancer burden related to BPA. Within the EU, breast cancer is the most common form of cancer in women, accounting for 28% (WHO website<sup>19</sup>). Breast cancer is the cancer type causing most cancer related deaths in women (17.2% of the total) (WHO website<sup>20</sup>, 2010). Breast cancer is likely to occur from puberty to end of life. The age-standardised incidence of breast cancer is reported to be around 90 per 100,000 in EU women (89.7 per 100,000 women in Western Europe by WHO<sup>21</sup> and 94.2 per 100,000 EU women by IARC (Ferlay *et al.*,

<sup>&</sup>lt;sup>18</sup> Proportion of 2 to 5% of the total annual ART cycle cost in EU28 (€2.4 - 3.1 billion) attributed to EDC exposure

<sup>&</sup>lt;sup>19</sup> http://www.euro.who.int/en/health-topics/noncommunicable-diseases/cancer/news/news/2012/2/early-detection-of-common-cancers/breast-cancer, accessed on 3 February 2017

<sup>&</sup>lt;sup>20</sup> http://www.euro.who.int/en/media-centre/sections/press-releases/2010/02/up-to-40-of-cancer-cases-could-be-prevented, accessed on 3 February 2017

http://www.who.int/cancer/detection/breastcancer/en/index1.html

2013)). The life time risk $^{22}$  for women of getting breast cancer in EU28 is around 1 in 8 women, or approximately 13% (e.g. 12.5% in France, 12.90% in the UK $^{23}$ , 14.3% in the Netherlands (Paap *et al*, 2008)). In terms of societal burden, recent literature shows that breast cancers also represent significant costs. The HEAL study (HEAL 2014) concluded that EDC-related female breast cancers direct (healthcare) costs are between 128 and 320 million € per year in the EU28. Including also indirect (productivity losses and informal care) costs, the total societal burden accounts for between 320 and 800 million € per year in the EU28. How large a part of this burden can be attributed to BPA itself has however not been calculated.

- In its state-of-the-art-report Kortenkamp et al. (2012) reports that although population-based statistics are not generally available for neurodevelopmental outcomes, surveys indicate that, for example in the US, several hundred thousand children have disabling childhood mental health conditions including mental retardation, learning disabilities, autism and attention deficit hyperactivity disorder (ADHD). Learning difficulties may affect up to 10% of school children. Endocrine disruption is unlikely to be the only cause of such trends but may importantly contribute. In its assessment of health costs associated with ED chemicals, Rijk et al. (2016) stated that neurodevelopmental and behavioural diseases and disorders include several pervasive disorders that persist for a lifetime, thereby leading to prolonged costs. They concluded that these disorders comprise the largest contributors to the total EDC-associated socio-economic cost estimates. Especially the contribution of IQ loss dominates the cost, accounting for between 32 and 184 billion € per year for the EU28 (indirect cost). HEAL 2014 also provided an evaluation of neurological disorders affecting child brain development and behavior due to EDCs but only focusing on autism and ADHD. The study reported a direct cost of between 4.5 and 11.3 billion € per year for the EU28, autism being the major contributor. Bellanger et al. (2015) estimated a cost of €80-400 million for EDC-related autism spectrum disorders. Which part of this burden can be attributed to BPA itself has however not been calculated and no BPA-specific data are available to our knowledge. The emergence of an elevated prevalence of neurobehavioural disorders in particular in children (that may be partly linked to EDCs) as well as their important costs for the healthcare systems in particular, and for society at large when also considering possible indirect costs of future decreased productivity, is therefore a major societal concern.
- The number of people with diabetes has doubled since 1980, rising from 33 million in 1980 to 64 million in 2014 in Europe (WHO 2016). The European prevalence of diabetes among adults over 18 years of age has risen from 5.3% in 1980 to 7.3% in 2014 and the associated mortality has also increased. Likewise, according to WHO (2016), the (age-standardized) mortality rate due to high blood glucose for adults over 20 years old (both sexes) is estimated to be 55.7 per 100,000 in Europe and around 10% of all deaths can be attributed to high blood glucose in 2012 (both sexes). Due to these increasing prevalences, there is a growing societal concern about diabetes and related-diseases. For example, HEAL 2014 estimated the direct and indirect cost of diabetes due to EDCs between 6 and 15 billion € per year for the EU28. How large a part of this cost can be attributed to BPA itself has however not been calculated.

<sup>&</sup>lt;sup>22</sup> The lifetime risk of cancer is an estimation of the risk that a newborn child has of being diagnosed with cancer at some point during its life.

<sup>&</sup>lt;sup>23</sup> http://www.cancerresearchuk.org/cancer-info/cancerstats/incidence/risk/statistics-on-the-risk-of-developing-cancer

## Is derivation of a 'safe concentration' possible?

- With regard to alteration of the reproductive function and of estrous cyclicity in particular, when considering oral studies, it is noted that effects are detected in some studies at relatively high doses (100, 300 and 600 mg/kg in Laws et al., 2000, Delclos et al., 2014 and Tyl et al., 2008, respectively) and at relatively low doses in other studies (0.0005, 0.001, 1.2 and 1.2 mg/kg in Wang et al., 2014, Lee et al., 2013, Rubin et al., 2001 and Mendoza-Rodriguez et al., 2011, respectively). The uncertainty in the dose response was acknowledged by RAC in its restriction opinion (ECHA, 2015) that concluded that "effects on the reproductive system have been observed at and below the range where kidney effects occur, RAC considers it prudent to take them into account in hazard and risk assessment and in health impact assessment. RAC however acknowledges that the available information does not allow a quantification of the dose-response relationship."
- Regarding the mammary tissue proliferative effect, RAC agrees "that BPA has been shown to have a proliferative effect on mammary tissue at doses below the doses causing general toxicity (such as kidney weight changes). The effects on mammary gland development should be taken into account in hazard and risk assessment and in health impact assessment. In line with EFSA (2015), no individual study is however considered robust enough by RAC to serve as critical study for the identification of a starting point for DNEL derivation. Therefore the effects will be accounted for in the setting of Assessment Factors." When considering oral studies, it is noted that most of the oral studies identified a rather low NOAEL/LOAEL couple of 25/250 µg/kg/d (see Moral et al., 2008; Jenkins et al., 2009; Betancourt et al., 2010). Whereas in Delclos et al., 2015, doses from which BPA causes hyperplastic ductal lesions were difficult to estimate.
- A similar conclusion as for reproductive toxicity was reached by RAC regarding neurobehavioural effects. In addition, a number of recent studies have confirmed since then, that an alteration of cognitive performance is observed in the vast majority of the studies at doses below 9 mg/kg (point of departure used for DNEL derivation - BMDL10 for kidney effects in mice) and the form of the doseresponse relationship is still under discussion. Several studies consistently identify effects from/at 40-50 µg/kg/d. In some studies, a single dose of exposure was used which does not allow to assess the dose-response relationship. In a metaanalysis, Peluso et al. (2014) concluded that the occurrence of spatial skill changes displayed an inverted U-shaped curve and emphasised possible low dose effects. The study by Inagaki et al. (2012) well illustrates that the time of exposure may be a more critical parameter than the dose of exposure. An effect on object placement memory was only observed in adult females during proestrus and not during the other phases of the cycle. This result was consistent with other results in the study showing that the effect is dependent on the concomitant presence of estrogens. Therefore, other parameters than the dose could impact the 'safe concentration'.
- With regard to metabolism, in the restriction dossier (ANSES, 2014), the effects of BPA on metabolism were considered as critical effects to be used for the risk assessment. The study from Miyawaki et al., 2007 was selected as the key study for this type of effect and a LOAEL of 260 μg/kg bw/day was derived based on the results reported by the authors. This LOAEL was below the NOAEL used by EFSA and RAC to derive a safe level with BPA. The recent studies that have been published and assessed for this dossier (and presented above see Table 25) also reported effects of BPA at doses equivalent or below the LOAEL from Miyawaki et al. (2007). Moreover in some studies, in vivo and in vitro, non monotonic dose response relationship have been reported which may complicate the

determination of a safe level for this end point (Garcia-Arevalo  $et\ al.$ , 2016; Angle  $et\ al.$ , 2013; Song  $et\ al.$ , 2012). The level of sensitivity will also depend on the window of exposure and the susceptibility of the exposed population. Based on the available data, it will be difficult to define a safe level with a sufficient level of confidence. In addition, it is noted that our knowledge of the ED-related effects of BPA may not be complete. In particular, there is some emerging evidence of an effect of BPA on the immune function that has been recently investigated and also raises concern on possible effects at low doses (5  $\mu$ g/kg and possibly 0.5  $\mu$ g/kg in Menard  $et\ al.$ , 2014a and 2014b).

Altogether, the database shows important uncertainties in establishing a quantitative dose-response, as well as safe levels, with some studies identifying effects at doses below the point of departure used for DNEL derivation.

The synthesis of these elements for each effect as well as for ED-mediated effects of BPA in general are presented in Table 27 below.

#### Overall assessment

Overall, the effects of BPA that are linked to an ED MoA are considered to meet the criteria of ELoC. Therefore it is justified to identify BPA as an SVHC according to article 57(f) for effects on human health exerted through an ED MoA.

It is noted that the MSC has agreed in December 2016 to identify BPA as an SVHC according to article 57 (c) in relation to its recent harmonised classification Repr 1B-H360F and BPA is included in the Candidate List as an SVHC under Article 57 (c) by decision of ECHA ED/01/2017 of the 4 January 2017.

The reproductive toxicity of BPA is also one of the effects linked to the ED properties of BPA and it is considered that the additional identification of the ED MoA for this effect, as well as for the three other effects, through SVHC 57(f) criteria is fully justified as this additional identification has specific regulatory consequences:

- So far, agreeing on a SVHC-57(f) property for a substance is the only way to identify it as an ED. Indeed, there is no other regulation allowing an identification/ classification of a substance as an ED.
- In the prioritisation process for inclusion into Annex XIV, a score is calculated. The different inherent properties have different weight attributed and 57f (ED) is given a medium weight, higher than the low score allocated to 57 a, b or c. In the scoring system a higher score is given to a combination of inherent properties only when PBT or vPvB properties are involved. For all the other properties, and our case, identifying BPA as an ED in addition to e.g. reproductive toxicity will not result in summing the different scores or increasing the existing score: The score for ED alone will apply (ECHA, 2014b). Technically, there is therefore no double counting in relation to prioritisation. Additional identification as an ED is therefore important for the prioritisation process without any double counting effect.
- If a new identified SVHC property (57(f) endocrine disruption) is to be included in the entries of Annex XIV, a concentration limit of 0.1% would apply to the use of the substances in mixtures, with regard to authorisation obligations, whereas currently a higher concentration limit of ≥0.3% applies for substances identified under Article 57 (c).
- In accordance with Article 60(3), applicants may have to apply for authorisation under the socio-economic route, unless a threshold can be determined for which the risk for humans and the environment can be demonstrated to be adequately controlled for any exposure scenario. So far, the discussion on the capability of

scientists to determine a threshold for ED effects is still open. Specificities of EDs have been raised to justify that even if a threshold might exist at an individual level, it would be very difficult if not impossible to determine it at a population level. Indeed, critical windows of susceptibility and potential non-monotonic dose response are specificities rendering classical risk assessment complicated for EDs. These specificities are particularly relevant for BPA, as described above. More generally, with respect to risk assessment for authorisation applications, European Commission recognised in a report published in December 2016, that the possibility of demonstrating a threshold for EDCs is expected to be difficult given the current state of scientific knowledge, but cannot be excluded. As a consequence, European Commission confirmed that it will be up to the applicants to demonstrate a threshold for the uses of the substance they applied for (to be scrutinised then by RAC) and in case they cannot, they shall apply under the socio-economic route (European Commission, 2016).

To enable an appropriate assessment of potential future applications under the socioeconomic route, it is critical that the scope of the health effects associated with the ED MoA is appropriately defined and is as comprehensive as possible.

In addition, it makes much sense to present and analyse all the possible well-established effects linked to an ED MoA together as they present strong common features that highly support their overall plausibility.

Finally, it is noted that there is no legal provision in the REACH regulation that prevents the identification of a substance as an SVHC according to several criteria, whatever the possible link between them may be.

Table 27: Summary of factors to be considered in ELoC assessment for the different EDrelated effects identified

	Alteration of reproductive function	Disruption of mammary gland development	Effect on brain and behaviour	Effect on metabolism and obesity	Overall conclusion for BPA ED- related effects
Possible serious health effects?	YES Serious dysfunction as illustrated by its harmonised classification Repr 1B for fertility	YES ■ Factor of risk for breast carcinogenici ty through specific modification of tissue morphology and proliferative effects of BPA	YES • Serious dysfunction having consequence s on the cognitive performance	▼ES  Associated with serious chronic pathologies such as diabetes and obesity Diabetes is a major cause of blindness, kidney failure, heart attacks, stroke and lower limb amputation.	YES ■ Pattern of ED-related effects associated with serious dysfunction or pathologies clearly related to serious illness

Delay of health effects?	YES ■ Effects on the reproductive function can be induced after only development al exposure and do not systematicall y involve a direct exposure. In this case, they appear later in time when the reproductive system becomes functional.	PES Developmental and/or post- natal exposures can induce effects on the mammary gland which do not systematically involve a direct exposure. In this case, they appear later in time when breast (or mammary) tissues further develop.	YES • The effects of development al exposure impacts cognitive function later in life.	PES Developmental exposure has been shown to have consequences in relation to ED-related metabolism effects later in life	PES Development al exposure has been shown to have consequence s in relation to ED-related effects later in life. Future generations not protected.
Irreversibilit y of health effects?	YES Development al exposure will lead to late effects when exposure has stopped showing that these modifications are irreversible.	YES Development al and/or post-natal exposures may lead to late effects even when exposure has stopped suggesting that these modifications are irreversible.	YES Impairment of cognitive function can be induced after development al exposure. These modifications are irreversible as shown by the fact that they are visible when exposure has stopped.	YES  BPA-related effects may be induced after development al exposure without direct exposure	YES  • All ED- related effects may be induced after development al exposure without direct exposure.
Quality of life impaired?	POSSIBLY YES  Possible mental/psyc hological impact, modification of normal physiology.	POSSIBLY YES Impaired quality of life associated with breast cancer. Possible impact on well-being, social isolation, psychological depression,	YES  ■ Possible impact on ability to cope with daily tasks and difficulties, alteration of school performance, school failure affecting self-esteem and later achievement	YES  • •Diabetes is associated with serious comorbidities and reduced life span. Impact of monitoring and control of diabetes on daily quality of life.	YES Impaired quality of life associated with several ED-related effects of BPA

		anxiety.	, parental worries; possible impact on well-being and social abilities.		
Societal concern?	YES  • Widespread concern about fertility issues  • Cost implications for society in terms of healthcare	YES Concern about breast cancer.  Cost implications for society in terms of healthcare.	YES Increasing concern about neurobehavi oural disorders in particular in childhood High socioeconomic impact of neurobehavi oral effects	YES  Diabetes is a metabolic disease that represents a growing public health concern worldwide.  High socioeconomic impact of diabetes	YES  • Multiple ED- related effects associated with major societal/ethic al health concerns and socio- economic burden for the society as a whole
Is derivation of a 'safe concentratio n' possible?	POSSIBLY NO Difficulties in defining the 'safe concentratio n' based on inconsistent information across studies on dose response including low dose effects in some studies Critical windows of exposure may be more important than doses Derivation of safe concentratio n associated with large uncertainties	MOST PROBABLY NO Non Non Monotonic dose relationship observed in a few studies Critical windows of exposure may be more important than doses Derivation of safe concentratio n associated with uncertainties .	POSSIBLY NO Non monotonic dose relationship observed in some studies Critical windows of exposure may be more important than doses Derivation of safe concentratio n associated with large uncertainties and many co-factors to be still determined.	POSSIBLY NO Non monotonic dose relationship observed in some studies Critical windows of exposure may be more important than doses Derivation of safe concentratio n associated with large uncertainties and many co-factors to be still determined.	MOST PROBABLY NO Difficulty in the quantificatio n of doseresponse acknowledge d by RAC Possible immunotoxic effects not well investigated (low dose potential) Critical windows of exposure may be more important than doses Derivation of safe concentratio n associated with large uncertainties

## 6.3.2.2 Environment

Not relevant for this dossier

# 6.3.3 Conclusion on the hazard properties and equivalent level of concern assessment

Bisphenol A is proposed to be identified as a substance of very high concern in accordance with Article 57(f) of Regulation (EC) 1907/2006 (REACH) because it is a substance with *endocrine disrupting properties* for which there is scientific evidence of probable serious effects to human health which gives rise to an equivalent level of concern to those of other substances listed in points (a) to (e) of Article 57 REACH.

BPA has been shown to affect the reproductive function, mammary gland development, cognitive function and metabolism through pathways that commonly involve disruption of estrogenic regulation. The effects on female reproductive function include the induction, after both developmental and adult exposures, of cystic ovaries, changes in the uterus morphology, alteration of fertility parameters as well as estrous cycle disturbance. The estrous cycle is a perfectly synchronised and sequenced event that relies on a permanent endocrine dialogue between the ovary and the hypothalamo-pituitary system. Those pathways differentiate during fetal life and are largely influenced by numerous factors and in particular the steroid environment of the foetus. BPA at the adult stage alters the endocrine steroidogenic function of the ovary and more specifically the production of estrogens by the follicle, leading to disturbance in the estrous cycle. At the neuroendocrine level, BPA can also act during the perinatal/postnatal organisation or adult activation of the hypothalamus-pituitary system through changes in kisspeptin, gonadotrophin-releasing hormone (GnRH) expression, activity or liberation and sex steroid receptor expression that impact estrous cyclicity.

The effects on the mammary gland, depending on the period of exposure, include: modifications in the mammary tissue such as an increased number of terminal end buds (TEBs) relative to the ductal area, fewer apoptotic TEB cells, increased lateral branching and ductal hyperplasia, increased cell proliferation and decreased apoptosis in the glandular epithelium, ductal (and occasionally lobuloalveolar) and intraductal hyperplasia - ultimately increasing its susceptibility to chemical carcinogens. These effects were observed in rodent or in non-human primate following prenatal and/or post-natal exposure to BPA. Available data also support the plausibility that BPA, through interaction with the nuclear estrogen receptors (ERs), or G protein-coupled estrogen receptor (GPER) and indirectly with the progesterone receptor (PR), modulates estrogen and progestin agonist activities. Emerging epigenetic studies have reported changes related to estrogen-dependent genes (such as EZH2 and HOTAIR), as well as HOX genes (involved in embryogenesis and post-natal development) associated with BPA induced abnormal development and increased cancer susceptibility of the mammary gland.

BPA has been demonstrated to alter memory and learning after developmental, pubertal or adult exposure, based on multiple converging experimental studies reporting this functional effect as well as molecular and cellular changes in the brain (reduced expression of NMDAR, altered synaptogenesis). These effects are mediated through disturbance of estrogenic pathways as evidenced by the reversal of the functional, cellular and molecular effects of BPA by an ER antagonist and interference of BPA with estradiol-induced effects on behavior and spine density/neurogenesis.

The effects of BPA on metabolism in rodent and non-rodent after prenatal and/or perinatal or adult exposures include alteration of insulin secretion and/or release by  $\beta$ -pancreatic cell, or of insulin signalisation (signaling mechanisms) within insulin-sensitive organs (i.e., liver, muscle, adipose tissues) leading to variations in the expression levels of hepatic or adipose tissue markers which are indicative of a state of insulin resistance. It is therefore considered that BPA may increase the incidence of type-2 diabetes. Additionally, *in vivo* and *in vitro* experimental studies indicate that these effects may

involve ERa, ERβ or GPR30 pathways. Other hormones such as leptin and adiponectin, which are involved in resistance to insulin and lipogenesis, are also modified following BPA exposure. This shows that BPA could interfere in the balanced interplay between insulin secretion and insulin action that controls glycaemia. Most of the *in vitro* studies showing adverse effects of BPA on adipocyte differentiation and function point to alteration of endocrine mechanisms (e.g., adiponectin release, insulin signaling cascade effectors). Overall, it is suggested that the pancreas is targeted by BPA, the mechanisms could differ depending on the period of exposure (fetal life or adulthood) and that an ED MoA is involved. Lastly, mainly based on similarities in homeostatic regulation of insulin production and sensitivity between animals and humans, these effects are considered relevant for humans.

The steps of the respective mechanisms of action are specific for each effect. The complexity of the toxic response to BPA suggests multiple MoA that may interact but most importantly, the available evidence shows that disruption of the estrogenic pathway is central and consistently involved in each of the four effects.

In conclusion, on the basis of evidence available in relation to alteration of reproductive function, mammary gland development, cognitive function and metabolism, BPA can be considered an endocrine disruptor for human health.

It is not excluded that BPA may also alter other physiological functions, e.g. the immune function, through a similar ED MoA but the level of evidence is considered insufficient at the moment for this effect to be presented.

The range of effects induced by BPA in relation to its ED MoA is considered serious. All these ED-related effects are characteristically (but not only) observed after developmental exposure to BPA, with consequences that are observed later in life. As they appear a long time after the exposure, they are indeed considered permanent and irreversible. In addition, the effects of BPA are associated with conditions that may lead to a reduced quality of life. In particular breast cancers, neurobehavioural disorders and diabetes are observed with high prevalence and increasing trends during the last decades in Europe and raise indisputable societal concern, also in relation to their potential economic burden on the health systems. Finally, for each of the four effects, the database shows important uncertainties in establishing a quantitative dose-response as well as safe levels, with some studies identifying effects at doses below the point of departure used by RAC for DNEL derivation and on-going discussions on the shape of the dose-response relationship and the parameters impacting the dose-response (period of exposure and concomitant presence of estrogen in particular). For these reasons, the effects of BPA that are linked to ED MoA therefore give rise to an equivalent level of concern to those of other substances listed in points (a) to (c) of Article 57 of REACH.

Thus, BPA is proposed to be identified as a substance of very high concern in accordance with Article 57(f) of REACH for effects in relation to human health exerted through an ED MoA.

## Part II

# 7. Registration and C&L notification status

#### 7.1 **Registration status**

**Table 28: Registration status** 

From the ECHA dissemination site <sup>24</sup>						
Registrations	<ul><li>✓ Full registration(s)</li></ul>					

## 7.2 CLP notification status

**Table 29: CLP notifications** 

	CLP Notifications <sup>25</sup>
Number of aggregated notifications	46
Total number of notifiers	1837

http://echa.europa.eu/registration-dossier/-/registered-dossier/15752 (accessed on 13 June 2016)
 C&L Inventory database, <a href="http://echa.europa.eu/web/guest/information-on-chemicals/cl-inventory-database">http://echa.europa.eu/web/guest/information-on-chemicals/cl-inventory-database</a> (accessed 6 June 2016)

# 8. Total tonnage of the substance

Table 30: Tonnage status

Total tonnage band for the registered substance (excluding the volume registered under Art 17 or Art 18) <sup>26</sup>	1 000 000 – 10 000 000 t/pa
Tonnage information from public sources other than registration dossiers	1.6 Mt/pa in 2005 (ANSES 2011)

According to Plastics Europe<sup>27</sup> the production of polycarbonate, that constitutes the major use of BPA by volume, continues to increase due to the variety of applications of polycarbonate and its very good properties of impact resistance, transparency and smooth surface aspect. During the public consultation of the SVHC dossier regarding the identification of Bisphenol A for its reproductive toxicity (57(c)) in autumn 2016, Plastics Europe however commented that according to latest market data and perspectives, notably the IHS/CMAI World Report, October 2015, the production capacity for polycarbonate is expected to be flat until 2025. ANSES doesn't have access to this report and is not able to check this information.

## 9. Information on uses of the substance

The review of uses of BPA has been made on the basis of the aggregated registration dossiers<sup>28</sup>, the ECHA's dissemination site, the INERIS report published in 2010, the ANSES report published in 2011 compiling available knowledge about the uses of BPA, and on the basis of interviews of a few stakeholders.

Table 31 below summarises the uses from the registration dossiers. The conclusion on whether the use is in the scope of Authorisation or not is based on the Guidance for monomers and polymers (ECHA, 2012) and the general information available in the registration dossiers. Specific conditions and circumstances of use are not known and may impact the status of the substance as an intermediate or not as reflected in the ongoing acrylamide court case. Further analyses will be provided on the status of each use at later stages of the risk management process of the SVHC dossier.

Table 31: Uses from the registration dossiers

	Use(s)	Registered use	Use in the scope of Authorisation
Uses as	Manufacture of polycarbonate		
intermediate	Industrial use of bisphenol A for	Yes	No
(monomer or	manufacturing polymers <sup>o</sup>		

<sup>&</sup>lt;sup>26</sup> http://echa.europa.eu/registration-dossier/-/registered-dossier/15752 (accessed on 13 June 2016)

<sup>&</sup>lt;sup>27</sup> ANSES has interviewed Plastics Europe on May 2016 to collect data on uses, alternatives and exposure of Polycarbonate.

<sup>&</sup>lt;sup>28</sup> Aggregated on 13 May 2016

intermediate)	Manufacture of others substance		
,	Manufacture of epoxy resins#		
	Manufacture of coating materials#		
	Manufacture of epoxy resin hardeners#		
	Use of epoxy resin hardeners at		
	industrial sites#		
	Use of epoxy resin hardeners by		
	professional workers#		
	Formulation of epoxy resin hardeners		
	Manufacture of chemicals		
	Industrial repackaging of bisphenol A <sup>Δ</sup>		
	Industrial use of bisphenol A for		
Formulation or	manufacturing thermal paper	Yes	Yes
repacking	Industrial use of bisphenol A as antioxidant for processing PVC*	. 55	. 65
	Formulation of preparations <sup>o</sup>		
	Blending of polycarbonate		No
	Industrial manufacture of articles		
	made of polycarbonate		No
	Industrial use of bisphenol A for		
	manufacturing polymers*		Yes
	Use of bisphenol A as laboratory		
Uses at industrial	reagent <sup>o</sup>		Yes
sites	Industrial use of bisphenol A for	Yes	
	manufacturing chemicals*		Yes
	Industrial use of bisphenol A for		Vaa
	manufacturing thermal paper°		Yes
	Industrial use of bisphenol A as anti-		Yes
	oxidant for processing PVC°		163
	Other industrial use <sup>a</sup>		Yes
	Professional repackaging of bisphenol		Yes
	Aso	_	
Uses by	Professional Use of bisphenol A as antioxidant for processing PVC	V	Yes
professional workers	•	Yes	No
Workers	Professional use of thermal paperoo  Professional use of articles made of	_	INO
	PVC°		No
	Consumer use of bisphenol A in		
	thermal papero		
Consumer uses	Consumer use of bisphenol A in	Yes	No
	articles made of PVC°		
	Professional indoor use of articles		
	made of polycarbonate		
	Professional outdoor use of articles		
	made of polycarbonate		
	Consumer indoor use of articles made		
	of polycarbonate		
	Consumer outdoor use of articles made		
Article service life	of polycarbonate		
	Consumer use of articles made of	Yes	No
	polycarbonate <sup>o</sup>		
	Consumer use of articles made of epoxy resins <sup>o</sup>		
	Professional use of thermal paper <sup>o</sup>		
	Professional use of articles made of		
	PVC°		
	Consumer use of thermal paper <sup>o</sup>		
	Consumer use of articles made of PVC°		

\*BPA can be considered as an intermediate when it is transformed into BADGE that is the monomer in the production of the resin.

- $\triangle$ This use can be exempted from potential authorisation if it is conducted during the use of Bisphenol A as an intermediate.
- $^{\rm o}$  During the public consultation of the SVHC dossier regarding the identification of Bisphenol A for its reproductive toxicity (57(c)) in autumn 2016, the Bisphenol A-REACh-Consortium indicated that this use is not covered since the 12/2014 update of the Joint-CSR of the Lead Registrant. However this information came from the registration dossiers aggregated on 13 May 2016.
- During the public consultation of the SVHC dossier regarding the identification of Bisphenol A for its reproductive toxicity (57(c)) in autumn 2016, European Plastics Converters (EuPC) indicated that the use of BPA as an antioxidant in PVC is currently either fully phased out or in the process of being phased out. In a survey of industry, EuPC was unable to identify a company that was using BPA, while all responding companies did report that they had switched to another antioxidant. However this information came from the registration dossiers aggregated on 13 May 2016.
- During the public consultation of the SVHC dossier regarding the identification of Bisphenol A for its reproductive toxicity (57(c)) in autumn 2016, the Bisphenol A-REACh-Consortium indicated that the "Use of Bisphenol A as laboratory reagent" is not in general in the scope of a potential authorisation. According to them, the following existing applications are exempted:
  - Bisphenol A used as an intermediate in the laboratory, or
  - Bisphenol A used in the quality control during the manufacture of Bisphenol A, or
  - Quality control of Bisphenol A when it is used as an intermediate

 $\diamond$  These uses were reported in the aggregated dossier (13 May 2016) but BPA content in thermal paper will be restricted after 2 January 2020.

Based on ANSES's compilation of knowledge and not based on the registration dossier, the main uses are further discussed below with a specific interest in describing the range of target sectors for each use.

#### **Polymers**

#### Polycarbonates

Polycarbonate is an amorphous, clear polymer with high transparency, superior dimensional stability, good electrical properties, good thermal stability, and outstanding impact strength and ductility (ANSES, 2012).

Some polycarbonate thermoplastics are not based on BPA. These polymers are not in the scope of the dossier. Therefore, in the dossier, polycarbonate should be interpreted as BPA based.

BPA reacts first with sodium hydroxide to form a sodium salt of BPA, which then reacts with phospene to produce polycarbonate.

<sup>\*</sup> Relevant for uses that are not intermediate uses

Figure 20: Polycarbonate formation reaction (ANSES, 2011)

The bisphenol A polycarbonate belongs to thermoplastics. These polymers are clear, and display heat and impact resistance. However, they have limited resistance to chemical agents: hydrolysis of the polymer can occur at high temperature and alkaline pH, a reaction that can release BPA (ANSES, 2011). In this case, the status of BPA released from polycarbonate is questionable in relation to Authorisation.

Applications of polycarbonate are summarised in Table 32 below.

Table 32: Applications of polycarbonate (ANSES, 2011)

Sectors	Applications
Optical media	CDs, DVDs, Blu-Ray and other audio or video media/formats
Electrical and electronic goods	Electrical equipment such as sockets and switches  Housings for electronic equipment: injection-moulded items with polycarbonate mixtures, used mainly in the electrical and electronics industry (alarm devices, mobile phone cases, coils, screens, computers, domestic appliances, lamps, power sockets, cables, etc.)
Domestic appliances	Kettles (transparent water level gauge) etc.
Construction	Covers for solar panels  Roofs of stadiums and sports facilitites
Automotive and aeronautic and space applications	Certain items for motor vehicles (safety glazing, light reflectors, headlamps, bumpers, radiator and ventilation grilles, interior lighting, motorcycle windshields and helmets, car roof modules, decorative trimmings for dashboards or door interiors, etc.)  Windows for airplanes, covers for position lights,

	personal protection equipment for civil and military usage (e.g. helmet visors) <sup>29</sup>
Reusable containers and bottles	Food containers such as reusable water bottles, shockproof baby bottles*
	Tableware items such as plates, cups, water bottles
Medical and healthcare items <sup>30</sup>	Medical equipment: blood oxygenators, respirators, dialysers, incubators, breathing apparatuses, disposable instruments, non implantable medical devices, implantable urogynaecology medical devices, ventilation mask, humidifier, administration sets, drug containers, catheter, stopcock, injection caps, enteral nutrition kit, syringe components, filter needles, sharps collector accessories; Infusion (IV set components, needleless connectors, infusion disposables) Specimen collection (blood collection sets and accessories, urine cup, and plastic multilayer bottles) <sup>31</sup> Additional uses are referenced in the SCENIHR (2015) opinion on "The safety of the use of bisphenol A in medical devices".
Leisure and safety	Optical equipment (prescription glasses, protective glasses, frames, sunglasses, thermal camera lenses, etc.)
	Sporting goods and protective gear (e.g. hockey helmet visors)
Other	All applications that fall within the other categories, such as furniture and decorative items
*RDA is hanned from haby bottles in Europe according	ELLO

<sup>\*</sup>BPA is banned from baby bottles in Europe according to the EU Commission Regulation N°321/201132

In polycarbonate, BPA is a monomer that is considered as an intermediate and this use is not subject to authorisation under the REACH Regulation.

#### Other polymers

Based on previous work (INERIS, 2010; ANSES, 2011), BPA can be used for the synthesis of other polymers such as:

- Polyester carbonate: used in automotive and transport; consumer products (hair dryer parts, laundry iron parts, microwave doors, etc.); electrical/electronics (fuses, switches, light reflectors and diffusers, etc.); industry (protective masks,

<sup>&</sup>lt;sup>29</sup> Information obtained during the public consultation of the SVHC dossier regarding the identification of Bisphenol A for its reproductive toxicity (57(c)) in autumn 2016.

<sup>&</sup>lt;sup>30</sup> ANSES has interviewed SNITEM (The National Association of Medical Technology Industry) on 8 June 2016 to collect data on uses, alternatives and exposure of bisphenol A in medical devices.

<sup>&</sup>lt;sup>31</sup> ANSES has interviewed SNITEM (The National Association of Medical Technology Industry) on 8 June 2016 to collect data on uses, alternatives and exposure of bisphenol A in medical devices.

 $<sup>^{32}</sup>$  EU Commission Regulation n°321/2011 amending Regulation (EU) N°10/2011 as regards the restriction of use of bisphenol A in plastic infant feeding bottles.

indicator lights, etc.); medical (autoclaves, surgical lighting systems, medical packaging, etc.)<sup>27</sup>. At the time being, ANSES considers that BPA is an intermediate in the synthesis of certain polyester carbonate and that this use is therefore not subject to authorisation under the REACH Regulation.

- Polyarylates: used in medical equipment<sup>27</sup>, water treatment, automotive and aeronautics. At the time being, ANSES considers that BPA is an intermediate in the synthesis of polyarylates and that this use is therefore not subject to authorisation under the REACH Regulation.
- Polysulfones: used in medical equipment<sup>33</sup> (catheter, canula, cable interconnects<sup>34</sup>...), in domestic appliances (microwave utensils, dispensers, parts for coffee-makers, cookers, hair-dryers, etc.), as a material for filtration membranes (hemodialysis, drinking water, gas separation, food industry, etc.), in plumbing. BPA is a monomer in the synthesis of polysulfones, that is considered as an intermediate and this use is not subject to authorisation under the REACH Regulation.
- Polyetherimides: used in medical, electronic and electrical equipment (satellite antenna parts, components for methanol fuel cells), automotive, aeronautical (aviation parts, cockpit components, radomes), food equipment (pump components in beverage vending machines). At the time being, ANSES considers that BPA is an intermediate in the synthesis of polyetherimides and that this use is therefore not subject to authorisation under the REACH Regulation.
- Polyols: used in the production of polyurethane. At the time being, ANSES considers that BPA is an intermediate and that this use is therefore not subject to authorisation under the REACH Regulation.
- Polyamides: BPA has been reported to be introduced as an additive in a polyamide to give stability. It is however noted that according to the Bisphenol A-REACh-Consortium, this use is no longer part of the Joint-CSR<sup>35</sup>.
- Polybenzoxazines: BPA can be a precursor in the synthesis of benzoxazine monomers. These polymers have various applications: composites industry, in coatings, adhesives and encapsulants. At the time being, ANSES considers that BPA is an intermediate and that this use is therefore not subject to authorisation under the REACH Regulation.

#### Resins

#### Epoxy resins

Bisphenol A diglycidyl ether (BADGE) is the most widely used epoxy resin (95% of world tonnage for epoxy resins). Some epoxy resins are not BPA-based and are not in the scope of the dossier. Therefore, in the dossier, epoxy resin should be interpreted as BPA based.

<sup>&</sup>lt;sup>33</sup> ANSES has interviewed SNITEM (The National Association of Medical Technology Industry) on 8 June 2016 to collect data on uses, alternatives and exposure of bisphenol A in medical devices.

 $<sup>^{34}</sup>$  Information obtained during the public consultation of the SVHC dossier regarding the identification of Bisphenol A for its reproductive toxicity (57(c)) in autumn 2016.

 $<sup>^{35}</sup>$  Information obtained during the public consultation of the SVHC dossier regarding the identification of Bisphenol A for its reproductive toxicity (57(c)) in autumn 2016.

It is synthesised from BPA and epichlorhydrin in the presence of soda according to the reaction schema below.

$$(n+1) \quad HO \longrightarrow C \longrightarrow OH \quad + \quad (n+2) \quad CI - CH_2 - HC \longrightarrow CH_2 \quad + \quad (n+2) \quad NaOH$$

$$CH_3 \qquad \text{Epichlorhydrine} \qquad Soude$$

$$Bisphénol \quad A$$

$$CH_2 \longrightarrow CH - CH_2 - O \longrightarrow C \longrightarrow O \longrightarrow CH_2 \longrightarrow CH - CH_2 \longrightarrow O \longrightarrow CH_2 \longrightarrow O \longrightarrow CH_2 \longrightarrow CH \longrightarrow CH_2 \longrightarrow O \longrightarrow CH_2 \longrightarrow CH \longrightarrow CH_2 \longrightarrow OH \longrightarrow CH_2 \longrightarrow CH_2 \longrightarrow OH \longrightarrow CH_2 \longrightarrow CH_2$$

Figure 21: Synthesis of resins based on bisphenol A (ANSES, 2011)

Epoxy resins are used for their ability to protect against corrosion and their thermal stability. These resins are used as protective coatings, composites and laminates, in electrical and electronic applications, in the construction industry and in the manufacture of adhesives, among others (ANSES, 2011).

Applications of epoxy resins are summarised in Table 33 below.

Table 33: Applications of epoxy resins (ANSES, 2011; INERIS, 2010)

Sectors	Applications
Composites	Rackets, surfboards, helmets, pipe, wind turbine blades
	Composites in the hulls of ships, aircraft and coach structures
Electrical and electronic goods	Printed circuits and boards
Insulation materials	Flooring (industrial and public buildings)
	Garden tools and equipment
	Cutting tools for food
	Coatings in cookware
Coatings for cans and reels	Coatings for canned food and drinks
	Domestic appliances (refrigerators, dishwashers, vacuum cleaners)
	Heating, ventilation and air conditioning

	equipment		
Coatings in the automotive sector	Vehicle parts and coatings		
	Consumer and multi-purpose adhesives		
	Adhesives and sealants for civil engineering		
Adhesives and paintings	Mortar for tiling and flooring		
	Adhesives for floor coverings (linoleum, tiles)		
	Varnish wood mastic		
	Acid resistant floor paintings <sup>36</sup>		
Semiconductor industry	Die attach <sup>32</sup>		
	Printing ink		
Draducts for protection against light	Wood coatings		
Products for protection against light	Varnish on paper, cardboard, packaging		
	Coatings for plastics and metals		
	Products for reinforcing/repairing concrete surfaces, various materials pre-impregnated with resins: movable partitions, decorative panels, bonded gravel		
	Coatings for aluminium formwork, concrete or plaster moulds, in civil engineering		
	Coatings for metal containers (shipping containers, tanks, cooling, towers, industrial storage tanks)		
Civil engineering	Coatings for steel bridges		
	Coatings for gas and drinking water pipes		
	Construction of metal panels (roofs) and chipboard		
	Fibreglass in construction		
	Powder coating on concrete reinforcement bars, on shelving grilles and primer coatings		
Marine and Protective coatings	Water ballast tank, underwater ship hulls, cargo tank linings, offshore oil drilling platforms, supporting steel structures, sea containers, steel bridges, storage tanks, power plant scrubbers, electric motors, engines, machinery, drinking water distribution pipes, gas pipes,		

 $^{36}$  Information obtained during the public consultation of the SVHC dossier regarding the identification of Bisphenol A for its reproductive toxicity (57(c)) in autumn 2016.

	waterproofing		
Medical devices <sup>37</sup>	External carbon orthoses, implantable urogynaecology medical devices, in adhesives for the vast majority hypodermic needles, catheter lines and wing sets <sup>38</sup> etc.  Additional uses are referenced in the SCENIHR opinion on the safety of the use of bisphenol A in medical devices.		
Aerospace and aeronautics <sup>34</sup>	Composite prepeg and potting compound/edge filler for honeycomb structures, composite for aircraft structures		

For food contact applications, epoxy resins with BPA have been removed from the internal coatings of all containers and substituted by other resins in France since the national law n°2012-1442<sup>39</sup>. According to the French Association for Food Industry (ANIA) and the National Association of Manufacturers of Packaging Boxes and Metal Capping (SNFBM)<sup>40</sup>, epoxy resins are still used in the external coating of food-containers and in equipment for the food chain. Epoxy resins are also still utilised for internal coating in the rest of Europe.

Ethoxylated bisphenol A can be used in the manufacture of certain types of epoxy resins employed to increase adhesion and resistance against friction and abrasion in UV-curable formulations, such as wood varnish and inks.

At the time being, ANSES considers that BPA is an intermediate for manufacturing epoxy resin and that this use is therefore not subject to authorisation under the REACH Regulation.

#### Other resins

BPA can be used for the synthesis of other resins such as (INERIS, 2010; ANSES, 2011):

- Vinyl ester resins: these resins are obtained by adding a carboxylic acid to an epoxy resin. They are used in automotive and marine construction, swimming pools, mortars and concretes, coatings (floors, etc.), fibre optic media, aeronautics, gas cylinders.
- Phenolic plastic resins: certain phenoplast and phenolic resins could be made from BPA and formaldehyde (EC, 2010). These resins have several applications: timber industry (plywood, particle board), paper industry (laminates, separators for collector tank, filters), insulation (agglomeration of fibres, foams), abrasives (for grinders, abrasive papers), friction materials (for brakes and clutches), rubber and adhesives industry, coatings (paints, protective coatings, enamel

of bisphenol A in food contact materials.

<sup>&</sup>lt;sup>37</sup> ANSES has interviewed SNITEM (The National Association of Medical Technology Industry) on 8 June 2016 to collect data on uses, alternatives and exposure of bisphenol A in medical devices.

 $<sup>^{38}</sup>$  Information obtained during the public consultation of the SVHC dossier regarding the identification of Bisphenol A for its reproductive toxicity (57(c)) in autumn 2016.

<sup>&</sup>lt;sup>39</sup> The French law n°2012-1442 for the Suspension of Manufacture, Import, Export and Marketing of All-Purpose Food Packaging Containing Bisphenol A. The Constitutional Council Decision No 2015-480 QPC stated the suspension of the manufacture in France and the export from France do not affect the marketing of these products in other countries. Hence the "manufacturing" and "export" in the title were considered invalid.

<sup>40</sup> ANSES has interviewed ANIA and SNFBM on 1 June 2016 to collect data on uses, alternatives and exposure

insulated wires, printing inks), new high-tech applications (obtaining carbon and graphite, composites)

- Unsaturated polyester resins: they are divided into two groups:
  - Bisphenol fumarates
  - o BPA epoxy dimethacrylates (EC, 2010)

Unsaturated polyester resins are used in the following applications: furniture varnish (old application), gel coats (used as outer layers on boats, aircraft, swimming pools, etc.), buttons, inclusions, mastics (waterproofing, seals, vehicle repair), construction (mortars and concretes), reconstituted marble, fake wood, fake ivory, etc.

At the time being, ANSES considers that BPA is an intermediate for manufacturing others resins and that this use is therefore not subject to authorisation under the REACH Regulation.

#### Flame retardants

#### Tetrabromobisphenol A

BPA is used in the production of tetrabromobisphenol A (TBBPA) and derivatives (polymer TBBPA-based [68928-70-1], tetrabromobisphenol A bis (2,3-dibromopropylether) [21850-44-2], O,O-bis(2-hydroxyethyl)TBBPA [4162-45-2], O,O-bis(prop-2-en-1-yl)TBBPA [25327-89-3], TBBPA Diglycidyl ether [3072-84-2]). They can be used as reactive flame retardants in the manufacture of printed circuit boards (generally in BPA epoxy resins) and, as additive flame retardant for plastic casings of televisions or computer screens (generally in ABS plastic). TBBPA can be found in consumer products such as computers, televisions and dishwashers (ANSES 2011, INERIS 2010, INERIS 2015, BSEF Industry or trade association<sup>41</sup>). TBBPA has been reported to be also used in hydraulic fluids (ANSES, 2011) but according to BSEF Industry TBBA is not used for this application<sup>18</sup>.

TBBPA may be partially transformed into BPA in sewage treatment plants or the environment. (INERIS, 2010, ANSES, 2011)

At the time being, ANSES considers that BPA is an intermediate in the production of TBBPA and that this use is therefore not subject to authorisation under the REACH Regulation.

#### Tetrachlorobisphenol A

BPA is also used in the production of a second flame retardant, tetrachlorobisphenol A, employed as an additive in polymers, epoxy resins, phenolic resins and adhesives. (ANSES, 2011)

At the time being, ANSES considers that BPA is an intermediate in the production of

<sup>&</sup>lt;sup>41</sup> Information obtained during the public consultation of the SVHC dossier regarding the identification of Bisphenol A for its reproductive toxicity (57(c)) in autumn 2016.

tetrachlorobisphenol A and that this use is therefore not subject to authorisation under the REACH Regulation.

#### Bisphenol A-bis (diphenyl phosphate)

Bisphenol A bis (diphenyl phosphate) (BDP) is another flame retardant derived from BPA, but much less frequently used than TBBPA. It is sometimes proposed as an alternative to halogenated flame retardants (ANSES, 2011).

At the time being, ANSES considers that BPA is an intermediate for the production of BDP and that this use is therefore not subject to authorisation under the REACH Regulation.

## Thermal paper

BPA is an additive in the coating applied to thermal paper and is therefore used as a substance for this application. It is used as a developing agent, which causes a chemical reaction when the paper is heated, resulting in colour being produced when using the paper (ANSES, 2011; ANSES's restriction dossier, 2014).

There are two types of thermal paper manufactured for different uses:

- Top coating or high-quality paper (which provides high quality images) is used for identification tags (parcels, self-service weighing of fruit and vegetables, identification of pre-packaged fresh foods, etc.), tickets (cinema, concerts, etc.), identification badges, self-adhesive labels, lottery tickets and receipts.
- Eco-paper or unprotected thermal paper is used in receipts, cash register receipts and credit card slips.

BPA has not been used in top coating thermal paper since 2000, as it is not a rapid enough developer for this type of paper. BPA is mainly used in eco-paper, although it is increasingly being substituted (ANSES, 2011). Here BPA is used as an additive and this use is in the scope of Authorisation but the use of BPA in thermal paper is already the focus of a restriction. Indeed, further to the restriction dossier proposed by France for bisphenol A in thermal paper, RAC and SEAC released a joint opinion supporting restriction of use of BPA in thermal papers and the following restriction (entry 66) was introduced in Annex XVII of REACH by Regulation 2016/2235: BPA "Shall not be placed on the market in thermal paper in concentration equal to or greater than 0.02% by weight after 2 January 2020".

#### Special uses in the automotive industry

## Brake fluids

BPA itself is sometimes added in small proportions to brake fluids and hydraulic fluids as an antioxidant. In November 2010, the French Union of Petroleum Industries confirmed to Anses that BPA is only used in some brake fluid formulations (at concentrations less than 1%).

BPA is used as an antioxidant and this use is in the scope of the authorisation under REACH.

#### <u>Tyres</u>

According to the 2011 ANSES report, BPA can also be used in tyre manufacturing. BPA is incorporated into the polymer matrix for its antioxidant properties (ANSES, 2011).

BPA is used as an antioxidant and this use is in the scope of the authorisation under REACH.

During the public consultation of the SVHC dossier regarding the identification of Bisphenol A for its reproductive toxicity (57(c)) in autumn 2016, the European Automobile Manufacturers Association indicated that BPA can also be present in the lubricants and greases used in the Automotive Industry Processes. No information about the function of BPA has been provided.

#### Other uses

- Paint industry: bisphenols may be used in paint hardeners (ANSES, 2011).\_For this use the specific function of BPA and its relevance to the scope of authorisation is still under discussion.
- Heat transfer fluids and lubricants: BPA can be used in the composition of heat transfer fluids and lubricants, and as a treatment agent for resurfacing concrete (ANSES, 2011). BPA is used as a treatment agent and this use is in the scope of authorisation under REACH.
- Dental products: dental resins containing BPA-based monomers are reported to be used in resin-based composites for restoration and sealing. BPA can be present as an impurity in the cement (in the case of bisphenol A glycidyl methacrylate or bis-GMA) or as a by-product of degradation (in the case of bisphenol A dimethacrylate or bis-DMA) (ANSES, 2011; INERIS 2010). COMIDENT<sup>42</sup> has however indicated that they have no report of current use of bis-DMA (personal communication, 22 July 2016).\_For this use the specific function of BPA and its relevance to the scope of authorisation is still under discussion.
- Esters: BPA is used as a stabiliser in the manufacture of esters. It acts as an antioxidant to prevent product aging. These BPA-stabilised esters are used as PVC plasticisers primarily in the automotive industry, in pipes, dashboards, etc. BPA is used as an additive and this use is subject to authorisation under the REACH Regulation
- PVC: BPA is used in the manufacture of PVC and acts as (ANSES, 2011, aggregated dossier):
  - An antioxidant in the treatment of PVC and in the production of plasticisers used in PVC processing.
  - o A component of the package of additives used in treatment of PVC.

During the public consultation of the SVHC dossier regarding the identification of Bisphenol A for its reproductive toxicity (57(c)) in autumn 2016, the European Stabiliser Producers Association indicated that BPA was used as a PVC additive, generally in pre-

185 (239)

<sup>&</sup>lt;sup>42</sup> France Coordinating Committee of Dental Activities

mixes with other additives, including stabilisers. The manufacturers of stabilisers have replaced BPA in their new formulations; only a few electrical cable applications seem still to rely on this chemical and a reformulation process is on-going.

BPA is used as an antioxidant and additive and these uses are in the scope of authorisation under REACH.

BPA can be used in formulations used in parts marking ink and several types of adhesives (i.e rubber-based, for thread locking and structural bonding) (The Boeing Compagny: information obtained during the previous consultation phase of the SVHC dossier regarding the identification of Bisphenol A for its reproductive toxicity). For this use the specific function of BPA and its relevance to the scope of authorisation is not known.

The Table 34 below summarises the uses from ANSES's compiled knowledge. The conclusion on whether the use is in the scope of Authorisation or not is based on the Guidance for monomers and polymers (ECHA, 2012) and the general information available. Specific conditions and circumstances of use are not known and may impact the status of the substance as an intermediate or not as reflected in the ongoing acrylamide court case. Further analyses will be provided on the status of each use at later stages of the risk management process of the SVHC dossier.

Table 34: Uses from ANSES's compiled knowledge

Use of BPA	Function of BPA	Use in the scope of authorisation	
Synthesis of polycarbonate	Monomer	No	
Synthesis of certain polyester carbonate	Intermediate	No	
Synthesis of polyarylates	Intermediate	No	
Synthesis of polysulfones	Monomer	No	
Synthesis of polyetherimides	Intermediate	No	
Synthesis of polyols	Intermediate	No	
Synthesis of polyamides	of polyamides  Additive  Theoretically Yes according to the Bisphenol A-REAC Consortium, this use is more part of the Joint-CS		
Synthesis of polybenzoxazines	Intermediate	No	
Synthesis of epoxy resins	Intermediate	No	
Synthesis of vinyl ester resins	Intermediate	No	
Synthesis of phenolic plastic resins	Intermediate	No	
Synthesis of unsaturated polyester resin	Intermediate	No	
Synthesis of tetrabromobisphenol A and derivatives	Intermediate	No	
Synthesis of tetrachlorobisphenol A	Intermediate	No	
Synthesis of bisphenol A-bis (diphenyl phosphate)	Intermediate	No	
Coating to thermal paper	Additive	Yes but a restriction for bisphenol A in thermal paper comes into effect	
Used in brake fluids and hydraulic fluids	Antioxidant	Yes	
Used in tyres	Antioxidant	Yes	
Used in paint hardeners	To be determined at later stages		

 $<sup>^{43}</sup>$  Information obtained during the public consultation of the SVHC dossier regarding the identification of Bisphenol A for its reproductive toxicity (57(c)) in autumn 2016.

Used in heat transfer fluids and lubricants	Treatment agent	Yes	
Used in dental products	To be determined at later stages		
Synthesis of esters	Additive	Yes	
Used in the manufacture of PVC	Anti-oxidant	Yes	
	Additive	Yes	
Used in formulations, used in parts	For this use the specific function of BPA and its relevance		
marking ink and several types of	for the scope of authorisation is not known.		
adhesives			

## 10. Information on structure of the supply chain

No specific information is available on the structure of the supply chain.

## 11. Additional information

# **11.1** Substances with similar hazard and use profiles on the Candidate List

No other substances that have a bisphenol structure are currently on the Candidate List except phenolphthalein (EC 201-004-7) but that is not known to have similar uses as bisphenol A.

France is not aware of substances in the Candidate List that have similar uses. In particular, none of the identified potential alternatives of BPA as listed in Section 11.2 are on the Candidate list.

#### 11.2 Alternatives

Alternatives have been described for all uses of BPA without consideration whether the use is in the scope of authorisation in the aim to provide an overview of all available alternatives. Indeed, an alternative for one use may also be relevant for other uses. It needs to be noted that the relevance of these alternatives to reduce risks has not been assessed. Availability and technical feasibility of these alternatives have also not been assessed in this dossier.

The review of alternatives has been made on the basis of a) a bibliographical search, b) the restriction dossier for BPA in thermal paper proposed by France and c) a report published by ANSES compiling an inventory of potential alternatives of BPA according to the different uses (ANSES, 2013a). ANSES issued a call for contributions in September 2011, in order to collect any scientific data concerning available substitution products. The work of identifying alternatives to BPA, in the ANSES report, was performed on a review of literature and contacts with industry. 73 alternatives to BPA were identified in the report.

Alternatives to BPA may be identified in several ways:

- Direct substitution of BPA by another substance

- Substitution by another plastic material or another polymer having similar properties to the starting polymer
- Substitution by another material, other type of packaging
- Substitution by a mixture
- Substitution by a process

Alternatives are discussed for each of the main uses of BPA.

## Potential alternatives to polycarbonate BPA-based (PC)

Polycarbonate can be replaced by other types of plastics polymers or by other materials.

Table 35: Potential alternatives to Polycarbonate: other polymers (ANSES 2013a, CETIM 2012<sup>44</sup>)

Other plastics polymers			
Alternatives	Uses		
Polyphenylsulfone	Baby bottles and accessories		
Polyethersulfone	Baby bottles, children tableware		
Polyamides	PA-6: baby bottles, bottles, food packaging		
	PA-11: bottles, food containers		
	PA-12: baby bottles		
Polyethylene	PE-hd: milk bottles, reusable juice bottles,		
	packaging of milk and dairy products		
	PE-bd : food containers, bottles		
Polypropylene	Baby bottles, children tableware, childcare		
	articles, reusable water bottles, meal trays		
Copolyester	Baby bottles, reusable water bottles, kitchen		
	gears, childcare products, water bottles,		
	household appliances, medical devices		
Polyethylene terephtalate	Food bottles and containers		
Isosorbide material	Isosorbide polyester: containers used in the		
	microwave, reusable water bottles		
	Isosorbide polycarbonate: sport bottles, food		
	containers, parts of mixers and blenders		
Poly lactic acid (PLA)	Food packaging, bottles		
Topas® IT X1	Medical devices, food applications		
Melamine material	Tableware, kitchen utensils, meal trays		
Acrylonitrile-butadiene-styrene (ABS)	Tableware, kitchen utensils, electric kettles		

<sup>&</sup>lt;sup>44</sup> After SNITEM (The National Association of Medical Technology Industry)'s interview in June 2016, a report performed by CETIM (Technical Centre for Mechanical Industries) on alternatives to polycarbonate, has been sent to ANSES. This report is not public but some new alternatives have been identified.

Methyl methacrylate polystyrene	Perfect for thick parts
Styrene Butadiene Copolymer	Blister packs, Tests on Baby bottles
Styrene Butadiene Methacrylate	Cosmetic and food contact packaging
Methyl polymethacrylate	Cribs in maternities, dialysis cassette
Glycol polyethylene terephthalate	Blister packs

Table 36: Potential alternatives to Polycarbonate: other materials

Other materials		
Alternatives	Uses	
Glass	Baby bottles, reusable water bottles, water	
	bottles, food containers	
Ceramic	Tableware, kitchen utensils, meal trays	
Stainless steel or inox	Bottles and food containers	
Silicon	Baby bottles	

According to Plastics Europe<sup>45</sup>, substitution of BPA for the production of BPA-based polycarbonate is impossible because polycarbonate from another monomer doesn't have the same physicochemical and mechanical properties as BPA-based polycarbonate. Few studies have been conducted on the substitution of polycarbonate because alternatives are already in use for the major applications: baby bottles made from polypropylene were already manufactured before the ban of BPA, and containers, packaging and utensils used for food contact, are already manufactured with a copolyester to replace polycarbonate. Tritan copolyesters are produced from three monomers: dimethyl terephthalate (DMT, CAS # 120-61-6), 1,4 cyclohexanedimethanol (CHDM, CAS # 105-08-8), and 2,2,4,4-tetramethyl-1,3-cyclobutanediol (TMCD, CAS # 3010-96-6). These monomers are utilised in various ratios depending on the desired performance characteristics. For reusable water bottles, according to Plastics Europe, no alternative is needed now to replace BPA because reusable water bottles stocks are sufficient to meet the demand. Lifetime of reusable water bottles is estimated between 5 and 7 years. Reusable water bottles produced before 2014 are still on the market. As this use is minor compared to other uses, it did not impact the volume of PC produced. Producers are waiting for stabilisation of BPA's regulation in France and in Europe before working further on alternatives.

According to SNITEM's interview (The National Association of Medical Technology Industry), polycarbonate presents some properties such as transparency, radio sterilisability and heat resistance that are very important for medical devices. Some alternatives have been tested by industry but tests were not conclusive for all products. Certain polymers can replace polycarbonate in some products. In other cases, the product has to be redesigned with a new bonding process. In addition, in medical devices, prices are fixed for each product and substitution is hampered by constraints of cost limitation.

During the public consultation of the SVHC dossier regarding the identification of Bisphenol A for its reproductive toxicity (57(c)) in autumn 2016:

- European Diagnostic Manufacturers Association mentioned that alternatives to

 $<sup>^{45}</sup>$  ANSES has interviewed Plastics Europe on May 2016 to collect data on uses, alternatives and exposure of Polycarbonate.

BPA in polycarbonates for uses in medical devices are in very early stages of being assessed. However, overall the vast majority of known alternatives have a similar hazard profile to BPA. Due to the number of types of components involved across products (with each having its own set of functional requirements), it is likely that multiple alternative materials would have to be identified, to meet the appropriate safety, technical and sterilisation requirements of each product. Polycarbonate replacement, if possible, takes several years (for one product) to reach the regulatory approval stage from alternative material assessment and feasibility work and subsequent product development work.

- The industry Federchimica indicated that there are no verified alternatives to BPA.

## Potential alternatives to epoxy resins

Epoxy resins can be replaced by other resins or other materials or another process (ANSES 2013a, SNFBM 2016, Fache *et al* 2015; Janvier *et* al 2017).

Table 37: Potential alternatives to epoxy resins: other resins

Other resins			
Alternatives	Uses		
Polyester	Coating of food metal packaging :internal		
	coating of cans		
Carbonate polypropylene	Food packaging, internal coating of beer and		
	soda cans, food cans, food containers		
Oleoresin	Internal coating of cans		
Tannin resin	Cans, aeronautic, defense, and insulation		
	applications,		
Biolignine™	Adhesives		
Polyurethan	Fixed facilities for production, treatment and		
	distribution of water intended for human		
	consumption: internal coating of pipes, drinking		
	water tank; hot-water tank;		
	Wastewater, industrial water, sea water pipes;		
	heating circuits; Waterproofing for equipment		
	rooms		
Verdanol	Coating of flexible packaging, external coating		
	of packaging (cans for example)		
Resin UVL-Eco resin	Sailing applications		
Resin SPR	Electrical cables encapsulation, industrial floor		
	coating, drinking water pipe coating (upon		
	certification)		
Isosorbid resin	Internal coating of cans, top and metallic lids of		
	bottles and glass jars		
Polyacrylate	Floor coatings, electrical insulation		
Acrylic	Cans		

Vinyl resins :	Cans
- Vinyl coatings (vinyl chloride and vinyl	
acetate copolymer)	
- Organosol PVC	
PET	No specific information on uses
Vanilin diepoxy monomer	No specific information on uses
Syringaresinol-based resin	No specific information on uses

Table 38: Potential alternatives to epoxy resins: other materials

Other materials		
Alternatives	Uses	
Glass	Bottle, jar, food packaging	
Tetrapack®	Carton packaging	
Doypack®	Flexible bag	

Table 39: Potential alternatives to epoxy resins: other process

Other process		
Alternatives	Uses	
Decrease BPA migration in food containers <sup>46</sup>	Cans	

According to SNFBM (the National Association of Manufacturers of Packaging Boxes and Metal Capping), in France epoxy resins have been replaced by three types of resin in internal coatings of containers:

- Polyester
- Acrylic resin
- Vinyl resin

No single alternative exists to substitute BPA in epoxy resin for all food contact applications. Indeed, it is difficult to find an alternative that is adequate for both acid and non-acid food. SNFBM notes that the work on substitution is ongoing to improve the quality of alternatives (cost and use by date). More information about each alternative are detailed in the Anses Report (2013a).

During the public consultation of the SVHC dossier regarding the identification of Bisphenol A for its reproductive toxicity (57(c)) in autumn 2016:

- Aerospace Industries Association indicated that, To date, as with the primers, there are no viable alternatives to these materials that are based on epoxy chemistry. Lack of such materials would result in poorer performing primers, adhesives and composite structures, which would result in increased maintenance times and cost, decreased safety of flight, and increased risk of system failure.
- The industry Federchimica indicated that there are no effective substitutes exist of bisphenol A in epoxy resins and this would lead to difficulties to ensure the anti-corrosive properties to products both in the industrial field and in the food

<sup>&</sup>lt;sup>46</sup> It consists of adding a PET Film on cans to inactivate the internal surface of cans or using an Epoxy Resin paint to limit the migration of BPA in food.

sector.

- European Diagnostic Manufacturers Association mentioned that potential alternatives to BPA-based epoxy (including acrylic-based epoxy) for uses in medical devices is currently being investigated, require different manufacturing process technologies, and have a significantly higher price than BPA-based epoxy.

## Potential alternatives to bisphenol A in thermal paper

France has recently submitted a dossier about the restriction of BPA in thermal paper receipts. A detailed review of the alternatives of BPA for this use has been performed in this context (ANSES restriction dossier, 2014). Potential alternative substances to bisphenol A in thermal paper can either be another substance or another process (ANSES, 2011). Only direct substitution of BPA by another substance in thermal paper has been noted in this dossier to provide an overview of existing alternatives which can be maybe useful for other uses of BPA.

Substances that have been identified as potential alternatives to BPA in thermal paper are summarised in Table 40 below with their regulatory status (harmonised classification and SVHC status).

Table 40: Potential alternative substances to BPA in thermal paper

Substance	CAS N°	Harmonised classification according CLP regulation	Candidate List
4,4'-methylenediphenol (Bisphenol F - para)	620-92-8	No harmonised classification	No
2,2'methylenediphenol (Bisphenol F – ortho)	2467-02-9	No harmonised classification	No
4,4'-sulphonyldiphenol (Bisphenol S)	80-09-1	No harmonised classification	No
1,1-bis(4-hydroxyphenyl)-1-phenylethane (Bisphenol AP)	1571-75-1	H400: Aquatic Chronic 1 H410: Aquatic acute 1	No
2,2'-diallyl-4,4'-sulfonyldiphenol (TGSA)	41481-66-7	H317: Skin Sens 1 H411: Aquatic Chronic 2	No
4-(4-isopropoxyphenylsulfonyl)phenol (D8)	95235-30-6	H411: Aquatic Chronic 2	No
4-[[4-(2-Propenyloxy)phenyl]sulfonyl]phenol (BPS-MAE)	97042-18-7	No harmonised classification	No
4-4'-methylenebis(oxyethylenethio)diphenol	93589-69-6	H411: Aquatic Chronic 2	No
Phenol, 4,4'-sulfonylbis-,polymer with 1,1'- oxybis[2-chloroethane] (D90)	191680-83-8	No harmonised classification *	No
biphenyl-4-ol	92-69-3	No harmonised classification	No
4,4'-thiobisphenol	2664-63-3	No harmonised classification	No
4-tert-butylphenol	98-54-4	H315: Skin irrit 2 H318: Eye Dam 1	No

		H361f: Repr 2	
benzyl 4-hydroxybenzoate	94-18-8	No harmonised classification	No
dimethyl 4-hydroxyphthalate	120-47-8	No harmonised classification	No
dimethyl 4-hydroxyphthalate (DMP-OH)	22479-95-4	No harmonised classification	No
3,5-bis-tert-butylsalicylic acid	19715-19-6	No harmonised classification	No
zinc 3,5-bis(a-methylbenzyl)salicylate	53770-52-8	No harmonised classification	No
Benzenesulfonamide, 4-methyl-N-[[[3-[[(4-			
methylphenyl)sulfonyl]oxy]phenyl]amino]carbo nyl]-	232938-43-1	H411: Aquatic Chronic 2	No
p-[[p-benzyloxyphenyl]sulfonyl]phenol	63134-33-8	No harmonised classification	No
Urea-urethane Compound	321860-75-7	No harmonised classification*	No
4,4'-bis(N-carbamoyl-4- methylbenzenesulfonamide)diphenylmethane	151882-81-4	H351: Carc 2	No
o-[(4-hydroxyphenyl) sulfony]phenol	5397-34-2	No harmonised classification	No
4,4'-isopropylidenedi-o-cresol	79-97-0	No harmonised classification	No
methyl bis(4-hydroxyphenyl)acetate (MBHA)	5129-00-0	No harmonised classification	No
4,4'-isopropyllidenebis (2-phenylphenol)	24038-68-4	No harmonised classification	No
6,6'-di-tert-butyl-4,4'-butylidenedi-m-cresol	85-60-9	No harmonised classification	No
2,6-di-tert-butyl-p-cresol	128-37-0	No harmonised classification	No
octadecyl 3-(3,5-di-tert-butyl-4- hydroxyphenyl)propionate	2082-79-3	No harmonised classification	No
pentaerythritol tetrakis(3-(3,5-di-tert- butyl-4-hydroxyphenyl)propionate)	6683-19-8	No harmonised classification	No
4,4',4"-(1-methylpropanyl-3-ylidene) tris [6- tert-butyl-m-cresol]	1843-03-4	No harmonised classification	No
1,2-diphenoxyethane	104-66-5	No harmonised classification	No
Phenolic compound  *Assumed not to be on the Ell market (not referenced)	Confidential	/	/

<sup>\*</sup>Assumed not to be on the EU market (not referenced in ECHA database)

## **Potential alternatives to TBBPA**

Alternatives to TBBPA in flame retardants that have been identified, are summarised in table below with their regulatory status (harmonised classification and SVHC status) (ANSES, 2013; INERIS, 2015).

Table 41: Potential alternative substances to TBBPA in flame retardants

Substitution by a substance						
Alternatives	N°CAS	Harmonised classification according CLP regulation	Candidate List			
1,3,5-triazine, 2,4,6-tris(2,4,6- tribromophenoxy)-	25713-60-4	No harmonised classification	No			
1,2-Bis(pentabromophenyl) ethane	84852-53-9	No harmonised classification	No			
Ethylenebis(tetrabromophthalimide)	32588-76-4	No harmonised classification	No			
9,10-dihydro-9-oxy-10- phosphaphenanthrene-10-oxide (DOPO)	35948-25-5	No harmonised classification	No			
1,4-Benzenediol, 2-(6-oxido-6H- dibenz[c,e][1,2]oxaphosphorin-6-yl) (DOPO-HQ)	99208-50-1	No harmonised classification	No			
Poly (m-phenylene methyl phosphonate)	63747-58-0	No harmonised classification	No			
Hexaphenoxy-phosphazene	/	/	/			
Decabromodiphenylether (decaBDE)	1163-19-5	No harmonised classification	Yes			
Decabromodiphenylethane (DBDPE)	84852-53-9	No harmonised classification	No			
Triphenyl phosphate	115-86-6	No harmonised classification	No			
Isobutyl-phenyl phenyl phosphate	68937-40-6	No harmonised classification	No			
Resorcinol bis (2,6-xylyl phosphate)	139189-30-3	H317: Skin Sens 1	No			
Red phosphorus	7723-14-0	H228: Flam Sol H412: Aquatic Chronic 3	No			
Resorcinol-bis-diphényl-phosphate (DNP)	57583-54-7	No harmonised classification	No			
	Other alternati	ves				
Epoxy oligomers / Brominated polymer	rs hama					
Compounds with aluminium and phosp	HOFUS					
Aromatic phosphate oligomers Firoflex ® Sol-DP <sup>47</sup>						
Cyclic phenoxyphosphazenes						
<u>Cyclic phenoxyphosphazenes</u> PH-73FF™						

#### Potential alternatives to other uses

No data have been identified in literature on potential alternatives to other uses. Few information have been obtained on alternatives to other uses during communication with COMIDENT and during the public consultation of the SVHC dossier regarding the identification of Bisphenol A for its reproductive toxicity (57(c)) in autumn 2016.

COMIDENT has indicated (personal communication, 22 July 2016) that companies using bisGMA in light-curing dental composite material are looking for alternative components. One of them is a urethane dimethacrylate that has now begun to be found in the

<sup>&</sup>lt;sup>47</sup> No data are available on chemical name.

composition of some products on the market. However, during the public consultation of the SVHC dossier regarding the identification of Bisphenol A for its reproductive toxicity (57(c)) in autumn 2016, Bundesverb and der Arzneimittel-Hersteller e.V, the German Medicines Manufacturers' Association, commented on urethane dimethacrylate (UDMA) that can only be used to a limited extent as an alternative to Bisphenol A, because:

- the optical properties of radio-opaque composites would be different and would have to be revised
- UDMA can lead to a deterioration of mechanical properties.

During the public consultation of the SVHC dossier regarding the identification of Bisphenol A for its reproductive toxicity (57(c)) in autumn 2016, the European Automobile Manufacturers Association indicated that according to downstream users of Bisphenol A, most of the uses cannot be replaced with the same level of performance and cost efficiency, based on their experiences from assessing alternative substances for these products. Moreover, whenever the substitution is possible, the alternative could have the same toxicity level.

## 11.3 Existing EU legislation

BPA is subjected to a restriction in Annex XVII of REACH related to its use in thermal paper. Further to the restriction dossier proposed by France, RAC and SEAC released their joint Opinion in December 2015. For cashiers, the RAC concluded that the risks of exposure via thermal paper receipts are not adequately controlled and supported the proposed restriction. These risks include potentially severe effects on the unborn children of pregnant female workers. The SEAC concluded that comparing the socio-economic benefits to the socio-economic costs, the proposed restriction is considered unlikely to be proportionate. However, there may be favourable distributional and affordability considerations.

The following restriction (entry 66) was introduced on 12 December 2016 in Annex XVII of REACH by Regulation 2016/2235: BPA "Shall not be placed on the market in thermal paper in concentration equal to or greater than 0.02% by weight after 2 January 2020".

Furthermore, this substance has a harmonised classification under CLP as presented in section II of part 1.

BPA is not in the list of substances subject to authorisation (Annex XIV of REACH) but BPA is included in the Candidate List as an SVHC under Article 57 (c) by decision of ECHA ED/01/2017 of the 4 January 2017.

BPA is already addressed under several EU Directives and Regulations:

- Commission Directive 2009/161/EU of 17 December 2009 establishing an indicative occupational exposure limit of 10 mg/m³ (8h-TWA) for inhalable dust of BPA. Furthermore, SCOEL recommended on 11 June 2014 the value of 2 mg/m³ with respiratory tract irritation as critical effect (SCOEL 2014).
- EU Directive 94/33/EC on young people at work as BPA is currently classified as reprotoxic 1B (H361F), irritant (STOT SE 3) and sensitiser (Skin Sens.1) and thus fulfils the criteria specified in Annex I.3 (a-c) of this Directive: young people must not be in contact with this type of substance.
- EU Regulation 1980/2000 on products not eligible for a positive Eco-Label.
- Regulation (EC) No 1223/2009 on cosmetic products, Annex II list of

substances prohibited in cosmetic products (entry 1176). However, according to Article 17, the non-intended presence of a small quantity of a prohibited substance, stemming from impurities of natural or synthetic ingredients, the manufacturing process, storage, migration from packaging, which is technically unavoidable in good manufacturing practice, shall be permitted provided that such presence is in conformity with Article 3 (with regard to the safety for human health when cosmetic products are used under normal or reasonably foreseeable conditions of use).

- EU Toy Safety:
- 1) Annex II.III.3 of Directive 2009/48/EC on the safety of toys: Substances that are classified as [...] toxic for reproduction of category [...] 2 under Regulation (EC) No 1272/2008 shall not be used in toys, in components of toys or in micro-structurally distinct parts of toys. There are however derogations in particular for materials that comply with the specific limit values set out in Appendix C (annex II.III.7).
- 2) Commission Directive 2014/81/EU of 23 June 2014 amending Appendix C of Annex II to Directive 2009/48/EC of the European Parliament and of the Council on the safety of toys, as regards bisphenol A. The specific limit value for BPA used in toys intended for use by children under 36 months or in other toys intended to be placed in the mouth adopted in accordance with Article 46(2) is 0,1 mg/l (migration limit) in accordance with the methods laid down in EN 71-10:2005 and EN 71-11:2005.
  - EU Commission Regulation No 321/2011 amending Regulation (EU) No 10/2011 as regards the restriction of use of bisphenol A in plastic infant feeding bottles. BPA is authorised as additive or monomer in the manufacture of other plastic materials and articles in contact with food and water, except for the manufacture of polycarbonate infant feeding bottles.
  - Regulation 10/2011/EU: plastic materials in contact with food a specific migration limit value of 0.6 mg/kg food has been set.
  - Commission Directive 2011/8/EU of 28 January 2011 amending Directive 2002/72/EC as regards the restriction of use of bisphenol A in plastic infant feeding bottles established a specific migration limit SML (T) of 0.6 mg/kg (not to be used for the manufacture of polycarbonate infant feeding bottles).
  - Water Framework Directive (2000/60/EC):

In the frame of the Review of the Priority Substances, BPA is subject to a review for identification as a possible 'priority substance' or 'priority hazardous substance' (Annex 3 of the Position of the European Parliament adopted at second reading on 17 June 2008 with a view to the adoption of Directive 2008/105/EC of the European Parliament and of the Council on environmental quality standards in the field of water policy).

## 11.4 Previous assessments by other authorities

#### 11.4.1 Previous assessments in the EU regulatory context

A risk assessment for BPA was carried out in accordance with Council Regulation (EEC) 793/931 on the evaluation and control of the risks of "existing" substances (rapporteur

member state: UK). The final Risk Assessment Report was published in February 2010.<sup>48</sup> This risk assessment report was also the basis for a transitional Annex XV report submitted by the United Kingdom on 30 November 2008. No restriction on the manufacture or use of bisphenol A was proposed but some risk management measures for occupational uses were proposed:

- Establish an Indicative Occupational Exposure Limit (IOELV).
- Implementation of Risk Management Measures (RMMs) following registration of BPA under REACH.
- Industry to voluntarily update the 'Safety and Handling Guide'.

EFSA performed a full risk assessment of BPA in 2006 and updated its scientific advice on hazard identification several times since 2006 (EFSA, 2008; EFSA 2010).

EFSA published its latest comprehensive re-evaluation of BPA exposure and toxicity in January 2015. New data and refined methodologies have led EFSA's experts to reduce the safe level of BPA to 4  $\mu$ g/kg of bw/day to recognise potential uncertainties about potential effects of BPA on the mammary gland, reproductive, neurological, immune and metabolic systems. The highest estimates for dietary exposure and for exposure from a combination of sources (called "aggregated exposure" in EFSA's opinion) are three to five times lower than the new TDI and EFSA concluded on absence of consumer health risk from BPA exposure.

In April 2016, EFSA set up a working group of international experts to evaluate new scientific evidence on the potential effects of BPA on the immune system. EFSA is conducting the review following publication of a report by the Dutch National Institute for Public Health and the Environment, which raises concerns about the effects of BPA on the immune system of foetuses and young children on the basis of two new studies (Menard *et al.*, 2014) suggesting effects on the immune system in the same dose range as the current t-TDI. In its final opinion in October 2016, EFSA concluded that the new studies "are too limited to draw any conclusions for human health".

Regarding the use of BPA in medical devices, the Scientific Committee on Emerging and Newly Identified Health Risks (SCENIHR) concluded in 2015 on the safety for medical applications based on BPA that some scenarios still remain of concern. Risk for adverse effects of BPA may exist when the BPA is directly available for systemic exposure after non-oral exposure routes, especially for neonates in intensive care units, infants undergoing prolonged medical procedures and for dialysis patients. Although the benefit of medical devices must also be considered, the SCENIHR recommends that where practicable, medical devices that do not leach BPA should be used. The possibility of replacing BPA in these products should be considered against their efficiency in the treatment, as well as the toxicological profile of the alternative materials.

BPA was included in the Community Rolling Action Plan for Substance Evaluation 2012-2014 (evaluation year 2012, evaluating member state: Germany) and is still ongoing: a substance evaluation decision pursuant to Article 51(6) of the REACH regulation was taken by ECHA in November 2013<sup>49</sup> that required additional data with regard to skin absorption and exposure (industrial, professional and consumer) to be included in an update of Registration dossiers by 20 December 2015.

In 2014 RAC evaluated the effects of BPA on fertility and reproductive systems and concluded that BPA meets the criteria Repr. 1B (H360F) by exerting, in animal studies,

<sup>48</sup> http://echa.europa.eu/documents/10162/c6a8dcfc-1823-4d31-8a24-2c71168f0d217

<sup>49</sup> http://echa.europa.eu/de/information-on-chemicals/evaluation/community-rolling-action-plan/corap-table/-/dislist/details/0b0236e1807e375d

adverse effects on female reproductive capacity, female reproductive organs and male reproductive organs that are not secondary to other toxic effects and are considered relevant to humans (ECHA 2014).

Also in 2014 SCOEL adopted a recommendation for an OEL (SCOEL 2014). Respiratory tract irritation was taken as the critical effect and an inhalation NOAEC of 10 mg/m<sup>3</sup> as the starting point for recommending the OEL.

In the context of a restriction proposal by France, RAC evaluated the risks posed by BPA contained in thermal paper and a DNEL was derived (ECHA 2015). In accordance with EFSA's assessment, the DNEL was established based on kidney effects. RAC concluded that effects on brain and behaviour, effects on the female reproductive system and proliferative effects on the mammary gland were observed at and below the range where kidney effects occur. RAC considers it prudent to take into account these effects, as well as effects on metabolism and obesity and effects on the immune system, both in hazard and risk assessment and in health impact assessment. RAC however acknowledges that the available information does not allow a quantification of the dose-response relationship for these effects. Therefore, this was taken into account in the setting of Assessment Factors. On the basis of its evaluation, RAC considered that the risk for consumers is adequately controlled but the risk for workers is not adequately controlled.

#### 11.4.2 Previous French assessments

On 24 October 2008, the French Food Safety Agency (AFSSA) published an opinion regarding BPA in polycarbonate baby bottles likely to be heated in microwave ovens (AFSSA, 2008a). AFSSA concluded that "given the current knowledge and after analysing the latest publications and reports, it [deemed] that when the contents of polycarbonate baby bottles [were] heated in a microwave oven in realistic conditions (heating for less than 10 minutes), the quantities of BPA that [migrated] into food [were] much less than the maximum value of 50  $\mu$ g BPA per liter as used in EFSA as a conservative estimate of exposure."

On 21 November 2008, AFSSA published an opinion regarding the exposure assessment of BPA in water intended for human consumption and possible resulting health risks (AFSSA, 2008b). AFSSA concluded that "the daily intake of BPA, including that from migration from water contact materials, [did] not lead to a risk for consumers under normal conditions" and recommended nevertheless "when BPA is present in a compound, that migration of BPA in water be specifically screened for during public health certification procedures of water contact materials with a target quantification limit of 1  $\mu q/L$ ."

On 7 July 2009, AFSSA published a note regarding the publication by Stahlhut *et al.* (2009) on urinary elimination of BPA in humans (AFSSA, 2009). It concluded that the result of this study [did] not require AFSSA to reconsider either the information already acquired on BPA or the risk assessment previously issued by EFSA, on which AFSSA's risk assessments were based."

On 29 January 2010, AFSSA published an opinion on the critical analysis of the results of a developmental neurotoxicity study of BPA together with other recently-published data on its toxic effects (AFSSA, 2010a). The conclusions were that "toxicity studies performed in compliance with international standards [had] not so far demonstrated any risk to health at current levels of exposure. [...] However, some recent publications whose methodology [did] not authorise any formal conclusions, [mentioned] warning signals. The Agency also [recommended] investigating sources of exposure to bisphenol A other than food contact materials."

In the clarification of this last opinion published on 2 March 2010 (AFSSA, 2010b), AFSSA concluded that "in order to reduce, through the use of substitute products, human exposure to bisphenol A, particularly for pregnant women and newborns, AFSSA stresses the importance of a rigorous risk assessment process for any products being considered as substitutes for bisphenol A."

On 7 June 2010, AFSSA published an opinion regarding exposure to BPA in the French population and maximum levels of BPA in foods (AFSSA, 2010c). The conclusions were that BPA levels in foods, analysed mainly in France, were "appreciably lower than the specific migration limit that [had] been established in Europe". Dietary exposure levels in the French population, including infants and children under 3 were "significantly lower than the TDI set by EFSA" and were "comparable to those observed in other international studies". AFSSA considered that "consumer exposure to BPA should be kept as low as possible, especially for the most sensitive consumers. It therefore [recommended] reassessing BPA's specific migration limit by using the best technologies currently available [...] and the systematic labelling of household utensils that are in contact with foods and that contain BPA".

In 2010, the French Institute for Environment and Industrial Risks (INERIS) published a report compiling the technico-economic data about BPA (INERIS, 2010). In this report, the various uses of BPA were presented and a focus on emissions was made. The conclusion was that, at the EU level, BPA was mainly emitted in water. The main source was industrial recycling of thermal paper (representing 70% of total emissions in water). Substitution of BPA in thermal paper was thus considered as a priority but, according to this report, the technical difficulties encountered with substitution were important. Recent papers show that water is not a major route of human exposure to BPA<sup>50</sup>.

In 2011, ANSES published the results of an extensive work in order to identify the uses of reprotoxic substances and/or potential endocrine disruptors, and BPA in particular. It examined nearly sixty industry sectors that potentially use BPA, which were identified either through a literature search or through a questionnaire sent to French manufacturers that potentially use this substance. This work confirmed previous studies showing the great variety of uses, and therefore, products and articles, containing BPA. A second conclusion was that substitution was an issue because of the wide variety of uses that led to numerous possible alternatives (ANSES, 2011).

In another recent report, ANSES<sup>51</sup> (2011) reviewed recent scientific data with the aim to classify the observed effects according to a dedicated decision tree. The conclusions were based on the results of available human and animal data, which have most often been obtained at doses lower than the NOAEL of 5 mg/kg/day that was used to calculate the TDI currently used by EFSA. This report highlighted the possible existence of a non-monotonic dose-response relationship, which adds extra complexity to the interpretation of the results. The need to discuss the relevance of using toxicity reference values or Tolerable Daily Intakes for substances with non-monotonic dose-response curves was also highlighted. Some adverse effects in animals were considered as recognised and must be considered in a future risk assessment of this substance. These effects observed in animals are the following:

- Increase of occurrence of ovarian cysts after pre and postnatal exposures.
- Hyperplastic modification of the endometrium after pre and postnatal exposure.
- Early onset of puberty after pre- and postnatal exposures.
- Altered sperm production after adult exposure.
- Histological changes in neurogenesis following pre- or perinatal exposure.
- Effects on lipogenesis following prenatal or perinatal exposures or during

<sup>&</sup>lt;sup>50</sup> Arch Environ Contam Toxicol (2014) 66:86–99 Is Drinking Water a Major Route of Human Exposure to Alkylphenol and Bisphenol Contaminants in France? A Colin, C Bach, Ce Rosin, JF Munoz, X Dauchy

<sup>&</sup>lt;sup>51</sup> Available at: https://www.anses.fr/fr/system/files/CHIM-Ra-BisphenolA.pdf [in French]

adulthood.

- Effects on the mammary gland: acceleration of the mammary gland's structural maturation in adulthood and development of intra-ductal hyperplastic lesions after pre- or perinatal exposure to BPA.

Some effects are considered as suspected in humans and should be taken into account for the risk assessment:

- Effects on oocyte maturation in females in infertile couples undergoing ART.
- Effects on cardiovascular pathologies (coronary diseases) and diabetes.

In 2013, ANSES (2013b) published its assessment of the health risks of bisphenol  $A^{52}$ : The risk assessment, which took into account all exposure media (but excluded specific exposure situations), shows that under certain circumstances, the exposure of pregnant women to bisphenol A could pose a potential risk to the health of the unborn child. The identified effects relate to a change in the structure of the mammary gland in the unborn child that could promote subsequent tumour development. The risk potentially affects children of both sexes.

The confidence level associated with these results was described as "moderate" by the majority of the experts, given the many uncertainties in the current state of scientific knowledge. Concerning the other three types of effects examined for the risk assessment (effects on the brain and behaviour, effect on metabolism and obesity, effect on the female reproductive system), the risk appears to be "negligible", depending on the assumptions made. The calculation of exposure via refillable polycarbonate water containers shows that water bottled in such containers is a significant source of exposure to bisphenol A. Its consumption can contribute to an increase in exposure to bisphenol A and could therefore, when combined with other sources of exposure, lead to an "additional" risk to the unborn child of an exposed pregnant woman.

The specific assessment of risks associated with the handling or use of products and/or articles intended for the general public and containing bisphenol A shows that handling thermal paper receipts leads to potential risk situations for the four types of effects considered in the risk assessment, but with a confidence level considered "limited" by the experts, due to the many uncertainties.

Following this risk assessment, restriction of BPA in thermal papers has been proposed by France. RAC and SEAC released their joint Opinion in December 2015. The proposed restriction was approved on the 6th July 2016 by the REACH Committee and its formal adoption and publication is expected shortly.

## 11.4.3 Previous assessments in other contexts

In 2008, Environment Canada and Health Canada published a joint screening assessment of BPA (Health Canada, 2008a and b). Dietary intake was considered to be the major source of BPA. Other sources of exposure were examined: dental materials' contribution was minimal while the other uses of BPA were insignificant.

Neuro-behavioural effects of BPA in newborns and infants were the subject of concern regarding the results of studies in rodents. The most sensitive populations are pregnant women/fetus and infants. It was "considered appropriate to apply a precautionary

<sup>&</sup>lt;sup>52</sup> Available at: <a href="https://www.anses.fr/fr/system/files/CHIM2009sa0331Ra-0.pdf">https://www.anses.fr/fr/system/files/CHIM2009sa0331Ra-0.pdf</a> [in French]. See also ANSES' Opinion on the assessment of the risks associated with bisphenol A for human health, and on toxicological data and data on the use of bisphenols S, F, M, B, AP, AF and BADGE: <a href="https://www.anses.fr/en/system/files/CHIM2009sa0331Ra-0EN.PDF">https://www.anses.fr/en/system/files/CHIM2009sa0331Ra-0EN.PDF</a>

approach when characterising risk to human health. Therefore, it was concluded that bisphenol A [...] may constitute a danger in Canada to human life or health". These conclusions led to the addition of BPA to the list of Toxic Substances in Schedule 1 of CEPA 1999 (Canadian Environmental Protection Act). This addition enables the Canadian authorities to use regulatory instruments available in CEPA 1999 to manage the human health and environmental risks posed by BPA. The prohibition of the importation, sale and advertising of polycarbonate baby bottles that contain BPA came into force in March 2010.

In November 2009, WHO and FAO published an information note about the current state of knowledge and their future actions regarding the BPA issue. They expressed concern because of potential toxic and hormonal properties of BPA (INFOSAN, 2009).

In January 2010, the American Food and Drugs Administration (FDA) reversed its position that BPA is safe and expressed "some concern about the potential effects of BPA on the brain, behavior and prostate gland in fetuses, infants and young children" based on the NTP-CERHR<sup>53</sup> report (NTP-CERHR, 2008). At the same time, the FDA announced that it "will support changes in food can linings and manufacturing to replace BPA or minimise BPA levels where the changes can be accomplished while still protecting food safety and quality". In its latest updated report (FDA, 2014), FDA considers that BPA is safe at the current levels occurring in foods.

In a BPA study published on March 9, 2010, researchers from the Swiss Federal Institute of Technology, based in Zurich (von Goetz *et al.*, 2010) have examined nine different consumer groups who were exposed to 17 different potential sources of BPA (various food products, beverages, house dust...). The researchers concluded that PC baby bottles were the most relevant BPA exposure source for infants and children, and that for teenagers and adults, it was the consumption of canned food. The Swiss federal health authority (FOPH) concluded in 2016 that there is no risk for consumers from BPA as exposure is low (FOPH, 2016).

In 2010, a joint FAO/WHO expert meeting examined the Toxicological and Health Aspects of BPA. The Expert Meeting considered BPA concentrations in food, BPA migration from food contact and dental materials and BPA concentrations in air, dust and water. The Expert Meeting concluded that the highest estimated exposure occurs for infants (0-6 months) who are fed with liquid formula out of PC bottles. For children older than 3, the highest exposure estimates are well below the infants' level and the main source of exposure is canned food (94%). For adults the highest exposure is twice that of children older than 3. Food is the major source of exposure (limited data available). Some additional potential sources of exposure (unpackaged food and thermal paper) have been identified.

In 2011, The Japanese Research Institute of Science for Safety and Sustainability (RISS) and the National Institute of Advanced Industrial Science and Technology (AIST) published an updated hazard assessment of BPA (AIST & RISS, 2011). AIST had previously published a risk assessment document in November 2007 where it was concluded that the risks posed by BPA were below the levels of concern, so it was unnecessary to prohibit or restrict the use of BPA at this time (AIST, 2007). In their 2011 update, the BPA exposure estimate in Japanese people is found to be highest for the 1 to 6 year old children. For this population, the calculated Margin Of Exposure (MOE) was 730 to 770 while it was 40 000 to 81 000 for adults. The risk represented by BPA was qualified as "very small" but it should be noted that, since approximately ten years, the Japanese can manufacturers use a method for inactivating the surface of drink cans that reduce the BPA intake to 0,1-0,2  $\mu g/kg/day$  for average-exposure individuals (AIST, 2007).

<sup>&</sup>lt;sup>53</sup> National Toxicology Program - Center for the Evaluation of Risks to Human Reproduction

In a report published in 2011, the National Toxicology Program and Center for the Evaluation of Risks to Human Reproduction (NTP CERHR, 2011) concluded that the highest potential for human exposure to BPA was through "products that directly contact food such as food and beverage containers with internal epoxy resin coatings and through the use of polycarbonate tableware such as those used to feed infants". The exposure via dental sealant was concluded to be an "acute and infrequent event with little relevance to estimating general population exposure".

In 2010, the United States Environmental Protection Agency (US EPA, 2010a; US EPA, 2010b) launched a program to identify safer substitutes for BPA in the manufacture of thermal paper. Liao and Kannan (2011) confirmed the fact that, among all kinds of papers, the highest concentrations of BPA were found in thermal papers but it also underlined that non-negligible amounts of BPA were found in some recycled napkins and toilet papers due to contamination during the paper recycling process. The study also showed that the value of the general population's exposure due to this route was minor compared with exposure through diet. But the authors also concluded that this exposure had to be studied and with particular attention to occupationally exposed individuals whose median daily intake is 74 times higher than the general population.

In 2011, the Danish Environmental Protection Agency (Danish EPA) published a report about the migration of BPA from cash register receipts and baby dummies. This report has taken into account the actual recognised NOAEL of 5 mg/kg/day and estimated consequently a DNEL of 0.029 mg/kg bw/d. It concludes that there are no risks either for consumers (RCR=0.19) or for cashiers (RCR=0.79) with the cash register receipts. It also concludes that there is no risk with baby dummies (RCR=0.281).

The Swedish Chemicals Agency has initiated a survey on bisphenol A in toys and childrens' articles (KEMI 2012). The contracted laboratory identified that toys made of polycarbonate, PVC, polyurethane and epoxy may contain BPA. Of 80 analysed articles only 20 of them were made of any of these plastic materials. In toys made of polycarbonate up to 600 ppm could be detected. However migration studies showed that there was no leakage of BPA.

In 2016, the Dutch National Institute for Public Health and the Environment (RIVM) published a report on recommendations for risk management of BPA. This report advises the Dutch government to reduce BPA exposure in the short term wherever possible. According to RIVM, special attention needs to be devoted to protecting small children, pregnant women and women who breastfeed.

The RIVM further concludes that the RCRs for consumers, patients and workers may require revision in the light of the new insights into the immune system effects of BPA.

Different recommendations (general, for reduction of environmental risks and for reduction of human health risks) are proposed. In particular, Bureau REACH (RIVM) intends to inform other Member State competent authorities (MSCAs) under REACH and the ECHA on the effects of BPA exposure on the immune system. With this information, the ECHA could take action to initiate a Compliance Check requesting the DNELs for workers and consumers to be updated. This information will also be disseminated by Bureau REACH in the Risk Management Expert Meeting (RiME), via commenting on ongoing risk management analyses.

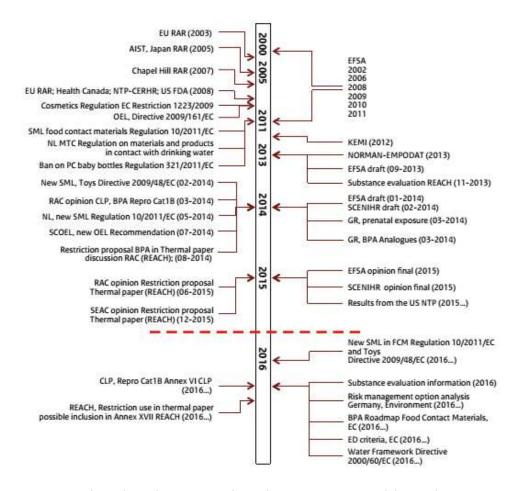


Figure 22 : Chronological overview of regulatory measures and key risk assessments on BPA, implemented and under development (RIVM 2016)

## REFERENCES

## References for Part I

- Abad MC *et al.* (2008) Structural determination of estrogen-related receptor gamma in the presence of phenol derivative compounds. J Steroid Biochem Mol Biol 108: 44-54
- Acconcia F et al. (2015) Molecular Mechanisms of Action of BPA. Dose Response. Oct 7;13(4)
- Acevedo N, Davis B, Schaeberle CM, Sonnenschein C, Soto AM. (2013) Perinatally administered bisphenol a as a potential mammary gland carcinogen in rats. Environ Health Perspect, 121:1040-6
- Adewale HB *et al.* (2009) Neonatal Bisphenol-A Exposure Alters Rat Reproductive Development and Ovarian Morphology Without Impairing Activation of Gonadotropin-Releasing Hormone Neurons. Biology of Reproduction, 81, 690-699.
- Aekplakorn W, Chailurkit LO, Ongphiphadhanakul B. (2014) Relationship of serum bisphenol A with diabetes in the Thai population, National Health Examination Survey IV, 2009. J Diabetes. 7(2):240-9.
- Agay-Shay K, Martinez D, Valvi D, Garcia-Esteban R, Basagana X, Robinson O, Casas M, Sunyer J, Vrijheid M. (2015) Exposure to Endocrine-Disrupting Chemicals during Pregnancy and Weight at 7 Years of Age: A Multi-pollutant Approach. Environ Health Perspect., 123(10):1030-7
- Ahmadkhaniha R, Mansouri M, Yunesian M, Omidfar K, Jeddi MZ, Larijani B, Mesdaghinia A, and Rastkari N. (2014). Association of urinary bisphenol a concentration with type-2 diabetes mellitus. J Environ Health Sci Eng, 12 (1):64.
- Ahmed S, Atlas E. (2016) Bisphenol S- and bisphenol A-induced adipogenesis of murine preadipocytes occurs through direct peroxisome proliferator-activated receptor gamma activation. Int J Obes (Lond), 40(10):1566-1573
- Ahn SW *et al.* (2008) Bisphenol A bis(2,3-dihydroxypropyl) ether (BADGE.2H2O) induces orphan nuclear receptor Nur77 gene expression and increases steroidogenesis in mouse testicular Leydig cells. Mol Cells, 31;26(1):74-80
- Aikawa H *et al.* (2004) Relief effect of vitamin A on the decreased motility of sperm and the increased incidence of malformed sperm in mice exposed neonatally to bisphenol A. Cell and Tissue research 315, 119-124.
- Akingbemi B *et al.* (2004a) Inhibition of testicular steroidogenesis by the xenoestrogen Bisphenol A is associated with reduced pituitary luteinizing hormone secretion and decreased steroidogenic enzyme gene expression in rat Leydig cells. Endocrinology 145, 592-603.
- Akingbemi BT *et al.* (2004b) Estrogen receptor-alpha gene deficiency enhances androgen biosynthesis in the mouse Leydig cell. Endocrinology, 144:84–93
- Al-Hiyasat AS *et al.* (2004). Leached components from dental composites and their effects on fertility of female mice. Eur J Oral Sci. 112(3), 267-72.

- Allred DC, Clark GM, Molina R, Tandon AK, Schnitt SJ, Gilchrist KW, Osborne CK, Tormey DC, McGuire WL (1992). Overexpression of HER-2/neu and its relationship with other prognostic factors change during the progression of in situ to invasive breast cancer. Human Pathology, 23, 974-979.
- Alonso-Magdalena P, Laribi O, Ropero AB, Fuentes E, Ripoll C, Soria B, and Nadal A. (2005). Low doses of bisphenol A and diethylstilbestrol impair Ca2+ signals in pancreatic alpha-cells through a nonclassical membrane estrogen receptor within intact islets of Langerhans. Environ Health Perspect, 113 (8):969-77.
- Alonso-Magdalena P, Morimoto S, Ripoll C, Fuentes E and Nadal A. (2006). The estrogenic effect of bisphenol A disrupts pancreatic beta-cell function in vivo and induces insulin resistance. Environ Health Perspect, 114 (1):106-12.
- Alonso-Magdalena P, Ropero AB, Carrera MP, Cederroth CR, Baquie M, Gauthier BR, Nef S, Stefani E and Nadal A. (2008). Pancreatic insulin content regulation by the estrogen receptor ER alpha. PLoS One, 3 (4):e2069.
- Alonso-Magdalena P, Vieira E, Soriano S, Menes L, Burks D, Quesada I, Nadal A. (2010) Bisphenol exposure during pregnancy disrupts glucose homeostasis in mothers and adult male offspring. Environmental Health Perspectives, 2010 118, 1243-1250.
- Alonso-Magdalena P, García-Arévalo M, Quesada I, Nadal Á. (2015) Bisphenol-A treatment during pregnancy in mice: a new window of susceptibility for the development of diabetes in mothers later in life. Endocrinology, 156(5):1659-70.
- Angle BM, Do RP, Ponzi D, Stahlhut RW, Drury BE, Nagel SC, Welshons WV, Besch-Williford CL, Palanza P, Parmigiani S, vom Saal FS, Taylor JA. (2013) Metabolic disruption in male mice due to fetal exposure to low but not high doses of bisphenol A (BPA): evidence for effects on body weight, food intake, adipocytes, leptin, adiponectin, insulin and glucose regulation. Reprod Toxicol, 42: 256-68.
- Ansari KI, Hussain I, Shrestha B, Kasiri S, Mandal SS. (2011a) HOXC6 Is transcriptionally regulated via coordination of MLL histone methylase and estrogen receptor in an estrogen environment. J Mol Biol, 411:334-49
- Ansari KI, Shrestha B, Hussain I, Kasiri S, Mandal SS. (2011b) Histone methylases MLL1 and MLL3 coordinate with estrogen receptors in estrogen-mediated HOXB9 expression. Biochemistry, 50:3517-27
- ANSES (2013a). CLH report. Proposal for harmonised classification and labelling. Bisphenol A. 17/07/2013. Available at <a href="https://echa.europa.eu/documents/10162/36b05a93-3e3c-44b1-bc8d-bff66b4b37ae">https://echa.europa.eu/documents/10162/36b05a93-3e3c-44b1-bc8d-bff66b4b37ae</a> (accessed on 4 November 2016)
- ANSES (2013b). Risk assessment of Bisphenol A (BPA) on human health. March 2013.
- ANSES (2014). Annex XV Restriction Report Proposal for a restriction 4.4′- isopropylidenediphenol (bisphenol A; BPA). May 2014.
- Antoine M, Teilhac MF, Poulet B, Cros J. (2010) De la cellule mammaire normale à la cellule cancéreuse. Médecine Nucléaire, 34, 14-22.
- Ariemma F, D'Esposito V, Liguoro D, Oriente F, Cabaro S, Liotti A, Cimmino I, Longo M, Beguinot F, Formisano P, Valentino R. (2016) Low-Dose Bisphenol-A Impairs Adipogenesis and Generates Dysfunctional 3T3-L1 Adipocytes. PLoS One, 4;11(3):e0150762.

- Arnsten AF et al. (2017) Novel Dopamine Therapeutics for Cognitive Deficits in Schizophrenia. Biol Psychiatry, 1;81(1):67-77.
- Aschengrau A, Coogan PF, Quinn M, Cashins LJ. (1998) Occupational exposure to estrogenic chemicals and the occurrence of breast cancer: an exploratory analysis. Am J Ind Med, 34:6-14
- Atlas E, Pope L, Wade MG, Kawata A, Boudreau A, Boucher JG. (2014) Bisphenol A increases aP2 expression in 3T3L1 by enhancing the transcriptional activity of nuclear receptors at the promoter. Adipocyte, 1; 3(3):170-9.
- Ayyanan A, Laribi O, Schuepbach-Mallepell S, Schrick C, Gutierrez M, Tanos T, Lefebvre G, Rougemont J, Yalcin-Ozuysal O, Brisken C. (2011) Perinatal exposure to bisphenol a increases adult mammary gland progesterone response and cell number. Mol Endocrinol, 25:1915-23
- Bäckström T. (1976) Epileptic seizures in women related to plasma estrogen and progesterone during the menstrual cycle. Acta Neurol Scand, 54(4):321-47.
- Baylin SB, Ohm JE. (2006) Epigenetic gene silencing in cancer a mechanism for early oncogenic pathway addiction? Nat Rev Cancer, 6:107-16
- Bellanger M, Demeneix B, Grandjean P, Zoeller RT, Trasande L (2015). Neurobehavioral deficits, diseases, and associated costs of exposure to endocrine-disrupting chemicals in the European Union. The Journal of Clinical Endocrinology and Metabolism, 100(4), 1256-1266.
- Benachour N *et al.* (2007) Cytotoxic effects and aromatase inhibition by xenobiotic endocrine disrupters alone and in combination. Toxicol Appl Pharmacol, 222(2):129-40.
- Berger RG *et al.* (2007) Influence of oral and subcutaneous bisphenol-A on intrauterine implantation of fertilized ova in inseminated female mice. Reproductive Toxicology, 23, 138-144.
- Berger RG *et al.* (2008). Impact of acute bisphenol-A exposure upon intrauterine implantation of fertilized ova and urinary levels of progesterone and 17beta-estradiol. Reprod Toxicol, 26(2), 94-9.
- Berger RG *et al.* (2010) Bisphenol-A exposure during the period of blastocyst implantation alters uterine morphology and perturbs measures of estrogen and progesterone receptor expression in mice. Reproductive Toxicology, 30, 393-400.
- Berger A, Ziv-Gal A, Cudiamat J, Wang W, Zhou C, Flaws JA. (2016) The effects of *in utero* bisphenol A exposure on the ovaries in multiple generations of mice. Reprod Toxicol, 60:39-52.
- Betancourt AM, Eltoum IA, Desmond RA, Russo J, Lamartiniere CA. (2010) *In utero* exposure to bisphenol A shifts the window of susceptibility for mammary carcinogenesis in the rat. Environ Health Perspect, 118:1614-9
- Betancourt AM, Wang J, Jenkins S, Mobley J, Russo J, Lamartiniere CA. (2012) Altered carcinogenesis and proteome in mammary glands of rats after prepubertal exposures to the hormonally active chemicals bisphenol a and genistein. J Nutr, 142:1382S-8S
- Beydoun HA, Khanal S, Zonderman AB, Beydoun MA. (2014) Sex differences in the association of urinary bisphenol-A concentration with selected indices of glucose homeostasis among U.S. adults. Ann Epidemiol, 24(2):90-7.

- Bhan A, Hussain I, Ansari KI, Kasiri S, Bashyal A, Mandal SS. (2013) Antisense transcript long noncoding RNA (IncRNA) HOTAIR is transcriptionally induced by estradiol. J Mol Biol, 425:3707-22
- Bhan A, Hussain I, Ansari KI, Bobzean SA, Perrotti LI, Mandal SS. (2014a) Histone methyltransferase EZH2 is transcriptionally induced by estradiol as well as estrogenic endocrine disruptors bisphenol-A and diethylstilbestrol. J Mol Biol, 426:3426-41
- Bhan A, Hussain I, Ansari KI, Bobzean SA, Perrotti LI, Mandal SS. (2014b) Bisphenol-A and diethylstilbestrol exposure induces the expression of breast cancer associated long noncoding RNA HOTAIR in vitro and in vivo. J Steroid Biochem Mol Biol, 141:160-70
- Biasiotto G, Zanella I, Masserdotti A, Pedrazzani R, Papa M, Caimi L, Di Lorenzo D. (2016) Municipal wastewater affects adipose deposition in male mice and increases 3T3-L1 cell differentiation. Toxicol Appl Pharmacol, 297:32-40.
- Biemann R, Navarrete Santos A, Navarrete Santos A, Riemann D, Knelangen J, Blüher M, Koch H, Fischer B. (2012) Endocrine disrupting chemicals affect the adipogenic differentiation of mesenchymal stem cells in distinct ontogenetic windows. Biochem Biophys Res Commun, 417(2):747-52.
- Bloom *et al.* (2011) Bisphenol A exposure reduces the estradiol response to gonadotropin stimulation during in vitro fertilization. Fertil Steril, 96(3):672-677.
- Boivin J, Bunting L, Collins JA, Nygren KG. (2007) International estimates of infertility prevalence and treatment-seeking: potential need and demand for infertility medical care. Hum Reprod, 22(6):1506-12.
- Bonefeld-Jorgensen EC *et al.* (2007) Endocrine-disrupting potential of bisphenol A, bisphenol A dimethacrylate, 4-n-nonylphenol, and 4-n-octylphenol in vitro: new data and a brief review. Environ Health Perspect, 115 (Suppl 1):69–76
- Boucher JG, Husain M, Rowan-Carroll A, Williams A, Yauk CL, Atlas E. (2014) Identification of mechanisms of action of bisphenol a-induced human preadipocyte differentiation by transcriptional profiling. Obesity (Silver Spring), 22(11):2333-43.
- Bowman RE *et al.* (2015) Bisphenol-A exposure during adolescence leads to enduring alterations in cognition and dendritic spine density in adult male and female rats. Horm Behav, 69:89-97
- Brisken, C., S. Kaur, T. E. Chavarria, N. Binart, R. L. Sutherland, R. A. Weinberg, P. A. Kelly, and C. J. Ormandy. (1999) Prolactin controls mammary gland development via direct and indirect mechanisms. Dev Biol, 210 (1):96-106.
- Brisken C and Rajaram RD. (2006) Alveolar and lactogenic differentiation. J Mammary Gland Biol Neoplasia, 11 (3-4):239-48.
- Brisken C. and Ataca D. (2015) Endocrine hormones and local signals during the development of the mouse mammary gland. WIREs Dev Biol, 4:181–195.
- Bromer JG, Wu J, Zhou Y, Taylor HS. (2009) Hypermethylation of homeobox A10 by in utero diethylstilbestrol exposure: an epigenetic mechanism for altered developmental programming. Endocrinology, 150:3376–3382.
- Bromer J, Zhou Y, Taylor M, Doherty L, Taylor H. (2010) Bisphenol-A exposure *in utero* leads to epigenetic alterations in the developmental programming of uterine estrogen response. FASEB J, 24(7):2273-80

- Cabaton NJ et al. (2010) Perinatal Exposure to Environmentally Relevant Levels of Bisphenol-A Decreases Fertility and Fecundity in CD-1 Mice. Environmental Health Perspectives, 119, 547-552.
- Campos C *et al.* (2016) Treatment of Cognitive Deficits in Alzheimer's disease: A psychopharmacological review. Psychiatr Danub, 28(1):2-12.
- Cantonwine D *et al.* (2010) Bisphenol a exposure in Mexico City and risk of prematurity: a pilot nested case control study. Environmental Health, 9, 62.
- Cao J et al. (2012) Neonatal Bisphenol A exposure alters sexually dimorphic gene expression in the postnatal rat hypothalamus. Neurotoxicology, 33(1):23-36.
- Cao J *et al.* (2014). Sex-specific Esr2 mRNA expression in the rat hypothalamus and amygdala is altered by neonatal bisphenol A exposure. Reproduction, 147(4):537-54.
- Carchia E, Porreca I, Almeida PJ, D'Angelo F, Cuomo D, Ceccarelli M, De Felice M, Mallardo M, Ambrosino C. (2015) Evaluation of low doses BPA-induced perturbation of glycaemia by toxicogenomics points to a primary role of pancreatic islets and to the mechanism of toxicity. Cell Death Dis, 29; 6:e1959.
- Cardiff RD, Anver MR, Gusterson BA, Hennighausen L, Jensen RA, Merino MJ, Rehm S, Russo J, Tavassoli FA, Wakefield LM, Ward JM, Green JE. (2000) The mammary pathology of genetically engineered mice: the consensus report and recommendations from the Annapolis meeting. Oncogene, 19, 968-988.
- Casas M *et al.* (2015) Exposure to bisphenol A during pregnancy and child neuropsychological development in the INMA-Sabadell cohort. Environ Res, 142:671-9.
- Castro B *et al.* (2013) Effects of adult exposure to bisphenol a on genes involved in the physiopathology of rat prefrontal cortex. PLoS One, 16;8(9):e73584.
- Castro B *et al.* (2015) Identification of dopamine- and serotonin-related genes modulated by bisphenol A in the prefrontal cortex of male rats. Chemosphere, 139:235-9.
- Chakraborti CK. (2015). Role of adiponectin and some other factors linking type 2 diabetes mellitus and obesity. World J Diab, 6 (15): 1296–1308.
- Chamorro-García R, Kirchner S, Li X, Janesick A, Casey SC, Chow C, Blumberg B. (2012) Bisphenol A diglycidyl ether induces adipogenic differentiation of multipotent stromal stem cells through a peroxisome proliferator-activated receptor gamma-independent mechanism. Environ Health Perspect, 120(7):984-9.
- Chen F, Capecchi MR. (1999) Paralogous mouse Hox genes, Hoxa9, Hoxb9, and Hoxd9, function together to control development of the mammary gland in response to pregnancy. Proc Natl Acad Sci U S A, 96:541-6
- Chen H, Sukumar S. (2003) Role of homeobox genes in normal mammary gland development and breast tumorigenesis. J Mammary Gland Biol Neoplasia, 8:159-75
- Cherrier MM *et al.* (2005) The role of aromatization in testosterone supplementation: effects on cognition in older men. Neurology, 64(2):290-6.
- Cherrier MM *et al.* (2009) Cognitive and mood changes in men undergoing intermittent combined androgen blockade for non-metastatic prostate cancer. Psychooncology, 18(3):237-47.

- Chitra KC *et al.* (2003) Induction of oxidative stress by bisphenol A in the epididymal sperm of rats. Toxicology, 185, 119-127.
- Cho J, Yu N-K, Choi J-H, Sim S-E, Kang SJ, Kwak C *et al.* (2015) Multiple repressive mechanisms in the hippocampus during memory formation. Science 350: 82–87.
- Choi J, Eom J, Kim J, Lee S, Kim Y. (2014) Association between some endocrine-disrupting chemicals and childhood obesity in biological samples of young girls: a cross-sectional study. Environ Toxicol Pharmacol, 38(1):51-7.
- Chrysant SG. (2015) Association of Exposure to Bisphenol A and Incidence of Cardiovascular Disease and Hypertension. J Clin Hypertens (Greenwich), 17(9):737-9
- Cobellis L *et al.* (2009) Measurement of bisphenol A and bisphenol B levels in human blood sera from endometriotic women. Biomedical Chromatography, 23, 1186-1190.
- Collett K, Eide GE, Arnes J, Stefansson IM, Eide J, Braaten A, Aas T, Otte AP, Akslen LA. (2006) Expression of enhancer of zeste homologue 2 is significantly associated with increased tumor cell proliferation and is a marker of aggressive breast cancer. Clin Cancer Res, 12:1168-74
- Commission Regulation (EU) 2016/1179 of 19 July 2016 amending, for the purposes of its adaptation to technical and scientific progress, Regulation (EC) No 1272/2008 of the European Parliament and of the Council on classification, labelling and packaging of substances and mixtures. Available at <a href="http://eurlex.europa.eu/legal-content/EN/TXT/?uri=uriserv%3AOJ.L..2016.195.01.0011.01.ENG">http://eurlex.europa.eu/legal-content/EN/TXT/?uri=uriserv%3AOJ.L..2016.195.01.0011.01.ENG</a> (accessed on 25 August 2016).
- Cox KH *et al.* (2010) Gestational exposure to bisphenol A and cross-fostering affect behaviors in juvenile mice. Hormones and Behavior, 58 (5): 754-761.
- Dai YE, Chen W, Qi H, Liu QQ. (2016) Effect of bisphenol A on SOCS-3 and insulin signaling transduction in 3T3-L1 adipocytes Mol Med Rep, 14(1):331-6.
- Daniel CW, Silberstein GB, Strickland P. (1987) Direct action of 17 beta-estradiol on mouse mammary ducts analyzed by sustained release implants and steroid autoradiography. Cancer Res, 47(22):6052-7
- Davis B, Fenton S. (2013) Mammary Gland. Tufts University, Boston, MA, USA, National Institute of Environmental Health Sciences, Research Triangle Park, NC, USA
- de Roux N *et al.* (2003) Hypogonadotropic hypogonadism due to loss of function of the KiSS1-derived peptide receptor GPR54. Proc Natl Acad Sci U S A, 100(19):10972-6.
- Deb P, Bhan A, Hussain I, Ansari KI, Bobzean SA, Pandita TK, Perrotti LI, Mandal SS. (2016) Endocrine disrupting chemical, bisphenol-A, induces breast cancer associated gene HOXB9 expression *in vitro* and *in vivo*. Gene, 590:234-43
- Delclos KB *et al.* (2014) Toxicity evaluation of bisphenol A administered by gavage to Sprague Dawley rats from gestation day 6 through postnatal day 90. Toxicol Sci, 139(1):174-97
- Della Seta D *et al.* (2006) Pubertal exposure to estrogenic chemicals affects behavior in juvenile and adult male rats. Hormones and Behavior, 50, 301-307.

- Demaegdt A, H, Vandermeiren K, Hectors TL, Jorens PG, Blust R, Vanparys C. (2013) Evaluation of a screening system for obesogenic compounds: screening of endocrine disrupting compounds and evaluation of the PPAR dependency of the effect. PLoS One, 8(10):e77481.
- Dhimolea E, Wadia PR, Murray TJ, Settles ML, Treitman JD, Sonnenschein C, Shioda T, Soto AM. (2014) Prenatal exposure to BPA alters the epigenome of the rat mammary gland and increases the propensity to neoplastic development. PLoS One, 9:e99800
- Diaz Weinstein S *et al.* (2013) Adolescent exposure to Bisphenol-A increases anxiety and sucrose preference but impairs spatial memory in rats independent of sex. Brain Res, 1529:56-65.
- Diel P, Olff S, Schmidt S and Michna H. (2002) Effects of the environmental estrogens bisphenol A, o,p'-DDT, p-tert-octylphenol and coumestrol on apoptosis induction, cell proliferation and the expression of estrogen sensitive molecular parameters in the human breast cancer cell line MCF-7. J Steroid Biochem Mol Biol, 80 (1):61-70.
- Ding L, Erdmann C, Chinnaiyan AM, Merajver SD, Kleer CG. (2006a) Identification of EZH2 as a molecular marker for a precancerous state in morphologically normal breast tissues. Cancer Res, 66:4095-9
- Ding L, Kleer CG. (2006b) Enhancer of Zeste 2 as a marker of preneoplastic progression in the breast. Cancer Res, 66:9352-5
- Dodds EC *et al.* (1938) Oestrogenic Activity of Certain Synthetic Compounds. Nature, 141, 247-248
- Doherty LF, Bromer JG, Zhou Y, Aldad TS, Taylor HS. (2010) *In utero* exposure to diethylstilbestrol (DES) or bisphenol-A (BPA) increases EZH2 expression in the mammary gland: an epigenetic mechanism linking endocrine disruptors to breast cancer. Horm Cancer, 1:146-55
- Durando M, Kass L, Piva J, Sonnenschein C, Soto AM, Luque EH, Munoz-de-Toro M. (2007) Prenatal bisphenol A exposure induces preneoplastic lesions in the mammary gland in Wistar rats. Environ Health Perspect, 115:80-6
- ECHA (2014). Committee for Risk Assessment (RAC). Opinion proposing harmonised classification and labelling at EU level of Bisphenol A; 4,4'-isopropylidenediphenol. Adopted 14 march 2014. ECHA. Available at <a href="http://echa.europa.eu/documents/10162/777918ff-33b5-46ff-be89-2bdc406d34fa">http://echa.europa.eu/documents/10162/777918ff-33b5-46ff-be89-2bdc406d34fa</a> (accessed on 6 June 2016).
- ECHA (2014b). Prioritisation of substances of very high concern (SVHCs) for inclusion in the Authorisation List (Annex XIV). 10 February 2014 <a href="https://echa.europa.eu/documents/10162/13640/gen approach svhc prior in recommendations en.pdf/e18a6592-11a2-4092-bf95-97e77b2f9cc8">https://echa.europa.eu/documents/10162/13640/gen approach svhc prior in recommendations en.pdf/e18a6592-11a2-4092-bf95-97e77b2f9cc8</a>
- ECHA (2015). Opinion on an Annex XV dossier proposing restrictions on Bisphenol A. Compiled version prepared by the ECHA Secretariat of RAC's opinion (adopted 5 June 2015) and SEAC's opinion (adopted 4 December 2015).
- ECHA (2016). Practical Guide: How to use alternatives to animal testing to fulfil your information requirements for REACH registration. Version 2.0 July 2016 <a href="https://echa.europa.eu/documents/10162/13655/practical guide how to use alternatives">https://echa.europa.eu/documents/10162/13655/practical guide how to use alternatives</a> en.pdf/148b30c7-c186-463c-a898-522a888a4404

- EFSA (2015). Scientific Opinion on the risks to public health related to the presence of bisphenol A (BPA) in foodstuffs, EFSA journal 2015;13(1):3978.
- Ehrlich S *et al.* (2012) Urinary Bisphenol A Concentrations and Implantation Failure among Women Undergoing in Vitro Fertilization. Environmental Health Perspectives, 120 (7), 978-983.
- Eilam-Stock T *et al.* (2012) Bisphenol-A impairs memory and reduces dendritic spine density in adult male rats. Behav Neurosci, 126(1):175-85. Erratum in: Behav Neurosci. 2012 Feb;126(1):195.
- Ekbom A, Trichopoulos D, Adami HO, Hsieh CC, Lan SJ. (1992) Evidence of prenatal influences on breast cancer risk. Lancet, 340:1015-8
- Elsworth JD *et al.* (2013) Prenatal exposure to bisphenol A impacts midbrain dopamine neurons and hippocampal spine synapses in non-human primates. Neurotoxicology, 35:113-20.
- Elsworth JD *et al.* (2015) Low circulating levels of bisphenol-A induce cognitive deficits and loss of asymmetric spine synapses in dorsolateral prefrontal cortex and hippocampus of adult male monkeys. J Comp Neurol, 523(8):1248-57.
- Ema M *et al.* (2001) Rat two-generation reproductive toxicity study of bisphenol A. Reprod Toxicol, 15(5):505-23.
- Ernster VL, Ballard-Barbash R, Barlow WE, Zheng Y, Weaver DL, Cutter G, Yankaskas BC, Rosenberg R, Carney PA, Kerlikowske K, Taplin SH, Urban N, Geller BM. (2002) Detection of ductal carcinoma in situ in women undergoing screening mammography. Journal of the National Cancer Institute, 94, 1546-1554.
- Esteller M. (2007) Cancer epigenomics: DNA methylomes and histone-modification maps. Nat Rev Genet, 8:286-98
- European Commission. (2016) Report from the Commission to the European Parliament, the Council and the European Economic and Social Committee in accordance with Article 138(7) of REACH to review if the scope of Article 60(3) should be extended to substances identified under Article 57(f) as having endocrine disrupting properties with an equivalent level of concern to other substances listed as substances of very high concern COM (2016) 814 final <a href="https://ec.europa.eu/transparency/regdoc/rep/1/2016/EN/COM-2016-814-F1-EN-MAIN-PART-1.PDF">https://ec.europa.eu/transparency/regdoc/rep/1/2016/EN/COM-2016-814-F1-EN-MAIN-PART-1.PDF</a>
- Fan Y et al. (2013) Does preconception paternal exposure to a physiologically relevant level of bisphenol A alter spatial memory in an adult rat? Horm Behav, 64(4):598-604.
- Faryna M, Konermann C, Aulmann S, Bermejo JL, Brugger M, Diederichs S, Rom J, Weichenhan D, Claus R, Rehli M, Schirmacher P, Sinn HP, Plass C, Gerhauser C. (2012) Genome-wide methylation screen in low-grade breast cancer identifies novel epigenetically altered genes as potential biomarkers for tumor diagnosis. FASEB J, 26:4937-50
- Feinberg AP, Ohlsson R, Henikoff S. (2006) The epigenetic progenitor origin of human cancer. Nat Rev Genet, 7:21-33
- Fenton SE. (2006) Endocrine-disrupting compounds and mammary gland development: early exposure and later life consequences. Endocrinology, 147(6 Suppl):S18-24.

- Fenton SE, Reed C, Newbold RR. (2012) Perinatal environmental exposures affect mammary development, function, and cancer risk in adulthood. Annu Rev Pharmacol Toxicol, 52:455-79
- Ferguson SA *et al.* (2014) Developmental treatment with ethinyl estradiol, but not bisphenol A, causes alterations in sexually dimorphic behaviors in male and female Sprague Dawley rats. Toxicol Sci, 140(2):374-92.
- Ferlay J, Steliarova-Foucher E, Lortet-Tieulent J, Rosso S, Coebergh JW, Comber H, Forman D, Bray F. (2013) Cancer incidence and mortality patterns in Europe: estimates for 40 countries in 2012. Eur J Cancer, 49(6):1374-403.
- Fernández M et al. (2009) Neonatal exposure to bisphenol A alters reproductive parameters and gonadotropin releasing hormone signaling in female rats. Environ Health Perspect, 117(5):757-62
- Fernandez SV, Russo J. (2010) Estrogen and xenoestrogens in breast cancer. Toxicol Pathol, 38:110-22
- Fernandez M, Bourguignon N, Lux-Lantos V, Libertun C. (2010a) Neonatal Exposure to Bisphenol A and Reproductive and Endocrine Alterations Resembling the Polycystic Ovarian Syndrome in Adult Rats. Environmental Health Perspectives, 118, 1217-1222.
- Fernandez SV, Snider KE, Wu YZ, Russo IH, Plass C, Russo J. (2010b) DNA methylation changes in a human cell model of breast cancer progression. Mutat Res, 688:28-35
- Fernandez SV, Huang Y, Snider KE, Zhou Y, Pogash TJ, Russo J. (2012) Expression and DNA methylation changes in human breast epithelial cells after bisphenol A exposure. Int J Oncol, 41:369-77
- Filardo EJ, Quinn JA, Bland KI, Frackelton AR, Jr. (2000) Estrogen-induced activation of Erk-1 and Erk-2 requires the G protein-coupled receptor homolog, GPR30, and occurs via trans-activation of the epidermal growth factor receptor through release of HB-EGF. Mol Endocrinol, 14:1649-60
- Filardo EJ, Thomas P. (2005) GPR30: a seven-transmembrane-spanning estrogen receptor that triggers EGF release. Trends Endocrinol Metab, 16:362–367.
- Filardo EJ, Graeber CT, Quinn JA, Resnick MB, Giri D, DeLellis RA, Steinhoff MM, Sabo E. (2006) Distribution of GPR30, a seven membrane-spanning estrogen receptor, in primary breast cancer and its association with clinicopathologic determinants of tumor progression. Clin Cancer Res, 12:6359-66
- Fischer C, Mamillapalli R, Goetz LG, Jorgenson E, Ilagan Y, Taylor HS. (2016) Bisphenol A (BPA) Exposure *In Utero* Leads to Immunoregulatory Cytokine Dysregulation in the Mouse Mammary Gland: A Potential Mechanism Programming Breast Cancer Risk. Horm Cancer, 7(4):241-51
- Fish EN. (2008) The X-files in immunity: sex-based differences predispose immune responses. Nat. Rev. Immunol, 8:737–744
- Foryst-Ludwig A, Clemenz M, Hohmann S, Hartge M, Sprang C, Frost N, Krikov M, Bhanot S, Barros R, Morani A, Gustafsson JA, Unger T and Kintscher U. (2008) Metabolic actions of estrogen receptor beta (ERbeta) are mediated by a negative cross-talk with PPARgamma. PLoS Genet, 4 (6):e1000108.

- Franssen *et al.* (2016) Delayed Neuroendocrine Sexual Maturation in Female Rats After a Very Low Dose of Bisphenol A Through Altered GABAergic Neurotransmission and Opposing Effects of a High Dose. Endocrinology, 157(5):1740-50
- Frick KM (2015). Molecular mechanisms underlying the memory-enhancing effects of estradiol. Hormones and Behavior, 74, 4–18.
- Friedmann Y, Daniel CA, Strickland P, Daniel CW. (1994) Hox genes in normal and neoplastic mouse mammary gland. Cancer Res, 54:5981-5
- Fujimoto VY *et al.* (2011) Serum unconjugated bisphenol A concentrations in women may adversely influence oocyte quality during in vitro fertilization. Fertility and Sterility, 95, 1816-1819.
- Funabashi T *et al.* (2004) Exposure to bisphenol A during gestation and lactation causes loss of sex difference in corticotropin-releasing hormone-immunoreactive neurons in the bed nucleus of the stria terminalis of rats. Psychoneuroendocrinology, 29 (4): 475-485.
- Gad SC (2007). The mouse. In 'Animal models in toxicology'. (Ed. SC Gad) pp. 19-146. (CRC: Sound Parkway, NW)
- Galea LA *et al.* (2013) Sex, hormones and neurogenesis in the hippocampus: hormonal modulation of neurogenesis and potential functional implications. J Neuroendocrinol, 25(11):1039-61.
- Galloway T *et al.* (2010) Daily Bisphenol A Excretion and Associations with Sex Hormone Concentrations: Results from the InCHIANTI Adult Population Study. Environmental Health Perspectives, 118, 1603-1608.
- García-Arevalo M, Alonso-Magdalena P, Rebelo Dos Santos J, Quesada I, Carneiro EM, Nadal A. (2014) Exposure to bisphenol-A during pregnancy partially mimics the effects of a high-fat diet altering glucose homeostasis and gene expression in adult male mice. PLoS One, 9(6):e100214.
- García-Arévalo M, Alonso-Magdalena P, Servitja JM, Boronat-Belda T, Merino B, Villar-Pazos S, Medina-Gómez G, Novials A, Quesada I, Nadal A. (2016) Maternal Exposure to Bisphenol-A During Pregnancy Increases Pancreatic β-Cell Growth During Early Life in Male Mice Offspring. Endocrinology, 157(11):4158-4171.
- Garcia-Gasca A, Spyropoulos DD. (2000) Differential mammary morphogenesis along the anteroposterior axis in Hoxc6 gene targeted mice. Dev Dyn,219:261-76
- Gilmour G et al. (2012) NMDA receptors, cognition and schizophrenia--testing the validity of the NMDA receptor hypofunction hypothesis. Neuropharmacology, 62(3):1401-12.
- Goldberg AD, Allis CD, Bernstein E. (2007) Epigenetics: a landscape takes shape. Cell, 128:635-8
- Gonçalves CR et al. (2010) Effects of prenatal and postnatal exposure to a low dose of bisphenol A on behavior and memory in rats. Environ Toxicol Pharmacol, 30(2):195-201.
- Gong H, Zhang X, Cheng B, Sun Y, Li C, Li T, Zheng L, Huang K. (2013) Bisphenol A accelerates toxic amyloid formation of human islet amyloid polypeptide: a possible link between bisphenol A exposure and type 2 diabetes. PLoS One, 8(1):e54198.

- Gould MN (1995). Rodent models for the study of etiology, prevention and treatment of breast cancer. Seminars in Cancer Biology, 6, 147-152.
- Grimm SL, Seagroves TN, Kabotyanski EB, Hovey RC, Vonderhaar BK, Lydon JP, Miyoshi K, Hennighausen L, Ormandy CJ, Lee AV, Stull MA, Wood TL and Rosen JM. (2002) Disruption of steroid and prolactin receptor patterning in the mammary gland correlates with a block in lobuloalveolar development. Mol Endocrinol, 16 (12):2675-91.
- Hampson E. (1990) Variations in sex-related cognitive abilities across the menstrual cycle. Brain Cogn, 14(1):26-43.
- Hamson DK *et al.* (2016) Sex Hormones and Cognition: Neuroendocrine Influences on Memory and Learning. Compr Physiol, 6(3):1295-337.
- Hasegawa Y *et al.* (2013) Bisphenol A significantly modulates long-term depression in the hippocampus as observed by multi-electrode system. Neuro Endocrinol Lett, 34(2):129-34.
- Haslam SZ, Shyamala G. (1979) Progesterone receptors in normal mammary glands of mice: characterization and relationship to development. Endocrinology, 105(3):786-95.
- Hass U *et al.* (2016) Low-dose effect of developmental bisphenol A exposure on sperm count and behaviour in rats. Andrology, 4(4):594-607
- Hayashida T, Takahashi F, Chiba N, Brachtel E, Takahashi M, Godin-Heymann N, Gross KW, Vivanco M, Wijendran V, Shioda T, Sgroi D, Donahoe PK, Maheswaran S. (2010) HOXB9, a gene overexpressed in breast cancer, promotes tumorigenicity and lung metastasis. Proc Natl Acad Sci U S A, 107:1100-5
- HEAL. (2014). Health costs in the European Union how much is related to EDCs? Health and Environment Alliance (HEAL). <a href="http://env-health.org/IMG/pdf/18062014">http://env-health.org/IMG/pdf/18062014</a> final health costs in the european union how much is realted to edcs-2.pdf
- Héliès-Toussaint C, Peyre L, Costanzo C, Chagnon MC, Rahmani R. (2014) Is bisphenol S a safe substitute for bisphenol A in terms of metabolic function? An *in vitro* study. Toxicol Appl Pharmacol, 280(2):224-35.
- Heindel JJ, Newbold RR, Bucher JR, Camacho L, Delclos KB, Lewis SM, Vanlandingham M, Churchwell MI, Twaddle NC, McLellen M, Chidambaram M, Bryant M, Woodling K, Gamboa da Costa G, Ferguson SA, Flaws J, Howard PC, Walker NJ, Zoeller RT, Fostel J, Favaro C, Schug TT. (2015) NIEHS/FDA CLARITY-BPA research program update. Reprod Toxicol., 58:33-44.
- Herath CB *et al.* (2004) Adverse effects of environmental toxicants, octylphenol and bisphenol A, on male reproductive functions in pubertal rats. Endocrine, 25, 163-172.
- Hiroi H *et al.* (2004) Differences in serum bisphenol a concentrations in premenopausal normal women and women with endometrial hyperplasia. Endocrine Journal, 51, 595-600.
- Hiyama M et al. (2011) Bisphenol-A (BPA) affects reproductive formation across generations in mice. J Vet Med Sci, 73(9), 1211-5.
- Hong SB *et al.* (2013) Bisphenol A in relation to behavior and learning of school-age children. J Child Psychol Psychiatry, 54(8):890-9.

- Honma S *et al.* (2002) Low dose effect of in utero exposure to bisphenol A and diethylstilbestrol on female mouse reproduction. Reproductive Toxicology, 16, 117-122.
- Hoover RN, Hyer M, Pfeiffer RM, Adam E, Bond B, Cheville AL, Colton T, Hartge P, Hatch EE, Herbst AL, Karlan BY, Kaufman R, Noller KL, Palmer JR, Robboy SJ, Saal RC, Strohsnitter W, Titus-Ernstoff L, Troisi R. (2011) Adverse health outcomes in women exposed in utero to diethylstilbestrol. N Engl J Med, 365:1304-14
- Howdeshell KL, Hotchkiss AK, Thayer KA, Vandenbergh JG, vom Saal FS (1999) Environmental toxins: Exposure to bisphenol A advances puberty. Nature, 401, 763-764.
- Howdeshell KL *et al.* (2008) Gestational and lactational exposure to ethinyl estradiol, but not bisphenol A, decreases androgen-dependent reproductive organ weights and epididymal sperm abundance in the male long evans hooded rat. Toxicological Sciences, 102, 371-382.
- Hrabovszky E *et al.* (2010) The kisspeptin system of the human hypothalamus: sexual dimorphism and relationship with gonadotropin-releasing hormone and neurokinin B neurons. Eur J Neurosci, 31(11):1984-98.
- Hu J, Yang S, Wang Y, Goswami R, Peng C, Gao R, Zhou H, Zhang Y, Cheng Q, Zhen Q, Li Q. (2015) Serum bisphenol A and progression of type 2 diabetic nephropathy: a 6-year prospective study. Acta Diabetol, 52(6):1135-41
- Huang H and Leung LK. (2009) Bisphenol A downregulates CYP19 transcription in JEG-3 cells. Toxicol Lett, 28;189(3):248-52.
- Humphreys RC, Krajewska M, Krnacik S, Jaeger R, Weiher H, Krajewski S, Reed JC and Rosen JM. (1996) Apoptosis in the terminal endbud of the murine mammary gland: a mechanism of ductal morphogenesis. Development, 122(12):4013-22.
- Hunt PA *et al.* (2003). Bisphenol A exposure causes meiotic aneuploidy in the female mouse. Current Biology, 13: 546-553.
- Hussain I, Bhan A, Ansari KI, Deb P, Bobzean SA, Perrotti LI, Mandal SS. (2015) Bisphenol-A induces expression of HOXC6, an estrogen-regulated homeobox-containing gene associated with breast cancer. Biochim Biophys Acta, 1849:697-708
- Ibrahim MA, Elbakry RH and Bayomy NA. (2016) Effect of bisphenol A on morphology, apoptosis and proliferation in the resting mammary gland of the adult albino rat. Int J Exp Pathol, 97 (1):27-36.
- Ida H *et al.* (2002) Disturbed spermatogenesis in mice prenatally exposed to an endocrine disruptor, Bisphenol A. Mammal Study, 27(1), 73-82.
- Inagaki T *et al.* (2012) Estrogen-induced memory enhancements are blocked by acute bisphenol A in adult female rats: role of dendritic spines. Endocrinology, 153(7):3357-67.
- INSERM (Institut national de la santé et de la recherche médicale). 2011. Reproduction et environnement, Expertise collective. Inserm, Paris. <a href="https://www.inserm.fr/content/download/38030/244999/.../reproduction">www.inserm.fr/content/download/38030/244999/.../reproduction</a> et environnem ent.pdf
- JRC (2013). Key Scientific issues relevant to the identification and characterisation of endocrine disrupting substances Report of the Endocrine Disruptors Expert Advisory Group (ED EAG). Eds. Munn S. and Gourmenou M. Pp 32. Available at:

- https://ec.europa.eu/jrc/en/publication/eur-scientific-and-technical-research-reports/key-scientific-issues-relevant-identification-and-characterisation-endocrine-disrupting
- JRC (2015). Identification of Substances of Very High Concern (SVHC) under the 'equivalent level of concern' route (REACH Article 57(f)) neurotoxicants and immunotoxicants as examples. JRC science for policy report. Authors: Laia Quiros Pesudo and Karin Aschberger. Available at <a href="http://publications.jrc.ec.europa.eu/repository/bitstream/JRC96572/jrc96572-identification%20svhc%20reach%20article%2057f.pdf">http://publications.jrc.ec.europa.eu/repository/bitstream/JRC96572/jrc96572-identification%20svhc%20reach%20article%2057f.pdf</a>
- Itoh H *et al.* (2007) Urinary bisphenol-A concentration in infertile Japanese women and its association with endometriosis: A cross-sectional study. Environmental Health and Preventive Medicine, 12, 258-264.
- Jain S *et al.* (2011) Protective effect of N-acetylcysteine on bisphenol A-induced cognitive dysfunction and oxidative stress in rats. Food Chem Toxicol, 49(6):1404-9.
- Jang YJ *et al.* (2012) High dose bisphenol A impairs hippocampal neurogenesis in female mice across generations. Toxicology, 296(1-3):73-82.
- Jašarević E *et al.* (2011) Disruption of adult expression of sexually selected traits by developmental exposure to bisphenol A. Proc Natl Acad Sci U S A, 12;108(28):11715-20.
- Jašarević E *et al.* (2013) Sex and dose-dependent effects of developmental exposure to bisphenol A on anxiety and spatial learning in deer mice (Peromyscus maniculatus bairdii) offspring. Horm Behav, 63(1):180-9.
- Jayashree S, Indumathi D, Akilavalli N, Sathish S, Selvaraj J, Balasubramanian K. (2013) Effect of Bisphenol-A on insulin signal transduction and glucose oxidation in liver of adult male albino rat. Environ Toxicol Pharmacol, 35(2):300-10.
- Jenkins S, Raghuraman N, Eltoum I, Carpenter M, Russo J, Lamartiniere CA. (2009) Oral exposure to bisphenol a increases dimethylbenzanthracene-induced mammary cancer in rats. Environ Health Perspect, 117:910-5
- Jenkins S, Betancourt AM, Wang J, Lamartiniere CA. (2012) Endocrine-active chemicals in mammary cancer causation and prevention. J Steroid Biochem Mol Biol, 129:191-200
- Johnson SA et al. (2016) Effects of developmental exposure to bisphenol A on spatial navigational learning and memory in rats: A CLARITY-BPA study. Horm Behav, 80:139-48.
- Jones PA, Baylin SB. (2007) The epigenomics of cancer. Cell, 128:683-92
- Jones BA and Watson NV. (2012) Perinatal BPA exposure demasculinizes males in measures of affect but has no effect on water maze learning in adulthood. Horm Behav, 61(4):605-10.
- Jukic AM *et al.* (2015) Urinary Concentrations of Phthalate Metabolites and Bisphenol A and Associations with Follicular-Phase Length, Luteal-Phase Length, Fecundability, and Early Pregnancy Loss. Environ Health Perspect,124(3):321-8.
- Kabuto H et al. (2004) Exposure to bisphenol A during embryonic/fetal life and infancy increases oxidative injury and causes underdevelopment of the brain and testis in mice. Life Sciences, 74, 2931-2940.

- Kandaraki E *et al.* (2010). Endocrine Disruptors and Polycystic Ovary Syndrome (PCOS): Elevated Serum Levels of Bisphenol A in Women with PCOS. J Clin Endocrinol Metab, 96: E480-4
- Kashiwada, S. *et al.* (2002). Fish test for endocrine-disruption and estimation of water quality of Japanese rivers. Water research, 36, 2161-2166
- Kato *et al.* (2003) Changes in reproductive organs of female rats treated with BPA during the neonatal period. Reprod Toxicol, 17(3):283-8.
- Kato H *et al.* (2006) Effects of bisphenol A given neonatally on reproductive functions of male rats. Reproductive Toxicology, 22, 20-29.
- Kauffman AS *et al.* (2007) Sexual differentiation of Kiss1 gene expression in the brain of the rat. Endocrinology, 148(4):1774-83.
- Kauffman AS. (2009) Sexual differentiation and the Kiss1 system: hormonal and developmental considerations. Peptides, 30(1):83-93.
- Khalil N, Ebert JR, Wang L, Belcher S, Lee M, Czerwinski SA, Kannan K. (2014) Bisphenol A and cardiometabolic risk factors in obese children. Sci Total Environ, 470-471:726-32.
- Kim ME *et al.* (2011) Exposure to bisphenol A appears to impair hippocampal neurogenesis and spatial learning and memory. Food Chem Toxicol, 49(12):3383-9.
- Kim K and Park H. (2013) Association between urinary concentrations of bisphenol A and type 2 diabetes in Korean adults: A population-based cross-sectional study. International Journal of Hygiene and Environmental Health, 216, 467-471
- Kitamura S *et al.* (2002) Thyroid hormonal activity of the flame retardants tetrabromobisphenol A and tetrachlorobisphenol A. Biochem Biophys Res Commun, 293: 554-559
- Kleer CG, Cao Q, Varambally S, Shen R, Ota I, Tomlins SA, Ghosh D, Sewalt RG, Otte AP, Hayes DF, Sabel MS, Livant D, Weiss SJ, Rubin MA, Chinnaiyan AM. (2003) EZH2 is a marker of aggressive breast cancer and promotes neoplastic transformation of breast epithelial cells. Proc Natl Acad Sci U S A, 100:11606-11
- Klenke U *et al.* (2016) BPA Directly Decreases GnRH Neuronal Activity via Noncanonical Pathway. Endocrinology, 157(5):1980-90.
- Knower KC, To SQ, Leung YK, Ho SM, Clyne CD. (2014) Endocrine disruption of the epigenome: a breast cancer link. Endocr Relat Cancer, 21:T33-55
- Ko A, Hwang MS, Park JH, Kang HS, Lee HS, Hong JH. (2014) Association between Urinary Bisphenol A and Waist Circumference in Korean Adults. Toxicol Res, 30(1):39-44.
- Kobayashi K *et al.* (2012) Lack of effects for dietary exposure of bisphenol A during *in utero* and lactational periods on reproductive development in rat offspring. J Toxicol Sci, 37(3):565-73.
- Koolschijn PC *et al.* (2014) The influence of sex steroids on structural brain maturation in adolescence. PLoS One, 8;9(1):e83929.
- Kortenkamp A. *et al.* (2012) State of the art assessment of endocrine disrupters. Final report.

  Available at <a href="http://ec.europa.eu/environment/chemicals/endocrine/pdf/sota">http://ec.europa.eu/environment/chemicals/endocrine/pdf/sota</a> edc final report.

  pdf

- Kortenkamp A *et al.* (2016). EU regulation of endocrine disruptors: a missed opportunity.Lancet Diabetes Endocrinol, 4(8):649-50
- Kramár EA, Chen LY, Brandon NJ, Rex CS, Liu F, Gall CM, et al. (2009) Cytoskeletal changes underlie estrogen's acute effects on synaptic transmission and plasticity. J Neurosci. 29: 12982–12993
- Kubo K *et al.* (2001) Exposure to bisphenol A during the fetal and suckling periods disrupts sexual differentiation of the locus coeruleus and of behavior in the rat. Neuroscience Letters, 304: 73-76.
- Kubo K *et al.* (2003) Low dose effects of bisphenol A on sexual differentiation of the brain and behavior in rats. Neuroscience Research, 45 (3): 345-356.
- Kuiper G et al. (1998) Interaction of estrogenic chemicals and phytoestrogens with estrogen receptor beta. Endocrinology, 139: 4252-4263
- Kumamoto T and Oshio S. (2013) Effect of fetal exposure to bisphenol A on brain mediated by X-chromosome inactivation. J Toxicol Sci, 38(3):485-94.
- Kumar D and Thakur MK. (2014) Perinatal exposure to bisphenol-A impairs spatial memory through upregulation of neurexin1 and neuroligin3 expression in male mouse brain. PLoS One, 9(10):e110482.
- Kundakovic M *et al.* (2013) Sex-specific epigenetic disruption and behavioral changes following low-dose in utero bisphenol A exposure. Proc Natl Acad Sci U S A, 110(24):9956-61.
- Kurian JR *et al.* (2015) Acute Influences of Bisphenol A Exposure on Hypothalamic Release of Gonadotropin-Releasing Hormone and Kisspeptin in Female Rhesus Monkeys. Endocrinology, 156(7):2563-70.
- Kuwahara R *et al.* (2013) Perinatal exposure to low-dose bisphenol A impairs spatial learning and memory in male rats. J Pharmacol Sci, 123(2):132-9.
- Kuwahara R et al. (2014) Bisphenol A does not affect memory performance in adult male rats. Cell Mol Neurobiol, 34(3):333-42.
- Kwintkiewicz J. *et al* (2010) Peroxisome proliferator-activated receptor gamma mediates the endocrine disrupter bisphenol a inhibition of fsh stimulated igf-i, aromatase and estradiol in human granulosa cells. Environ Health Perspect, 118: 400-406
- Kwon S *et al.* (2000) Pubertal development and reproductive functions of CrI:CD BR Sprague-Dawley rats exposed to bisphenol A during prenatal and postnatal development. Toxicological Sciences, 55, 399-406.
- Lacreuse A. (2006) Effects of ovarian hormones on cognitive function in nonhuman primates. Neuroscience, 138(3):859-67.
- Lacreuse A. and Herndon J.(2009) Nonhuman primate models of cognitive aging. In: Bizon J & Woods A., editors. Animal Models of Cognitive Aging. Humana Press, New York: p. 29-58
- LaKind JS, Goodman M and Naiman DQ. (2012) "Use of NHANES data to link chemical exposures to chronic diseases: a cautionary tale." PLoS One 7 (12):e51086.
- Lakind J. S., Goodman M and Mattison DR. (2014) Bisphenol A and indicators of obesity, glucose metabolism/type 2 diabetes and cardiovascular disease: a systematic review of epidemiologic research. Crit Rev Toxicol, 44 (2):121-50.

- Lang IA, Galloway TS, Scarlett A, Henley WE, Depledge M, Wallace RB and Melzer D. (2008) "Association of urinary bisphenol A concentration with medical disorders and laboratory abnormalities in adults." JAMA, 300 (11):1303-10.
- LaRocca J *et al.* (2011) Effects of in utero exposure to bisphenol A or diethylstilbestrol on the adult male reproductive system. Birth Defects Research (part B), 92, 526-533.
- Laws SC *et al.* (2000) Estrogenic activity of octylphenol, nonylphenol, bisphenol A and methoxychlor in rats. Toxicol Sci, 54:154–167
- Leclerc F, Dubois MF, Aris A. (2014) Maternal, placental and fetal exposure to bisphenol A in women with and without preeclampsia. Hypertens Pregnancy, 33(3):341-8
- Lee HR, Hwang KA, Park MA, Yi BR, Jeung EB, Choi KC. (2012) Treatment with bisphenol A and methoxychlor results in the growth of human breast cancer cells and alteration of the expression of cell cycle-related genes, cyclin D1 and p21, via an estrogen receptor-dependent signaling pathway. Int J Mol Med, 29:883-90
- Lee MR, Kim JH, Choi YH, Bae S, Park C, Hong YC. (2015) Association of bisphenol A exposure with overweight in the elderly: a panel study. Environ Sci Pollut Res Int, 22(12):9370-7
- Lee SG *et al.* (2013) Bisphenol A exposure during adulthood causes augmentation of follicular atresia and luteal regression by decreasing  $17\beta$ -estradiol synthesis via downregulation of aromatase in rat ovary. Environ Health Perspect, 121(6):663-9
- Leranth C *et al.* (2008) Bisphenol A prevents the synaptogenic response to estradiol in hippocampus and prefrontal cortex of ovariectomized nonhuman primates. Proc Natl Acad Sci U S A, 105(37):14187-91.
- Levy G *et al.* (2004) Bisphenol A induces feminization in Xenopus laevis tadpoles. Environmental Research, 94(1):102-111.
- Li DK *et al.* (2010a) Occupational exposure to bisphenol-A (BPA) and the risk of self-reported male sexual dysfunction. Human Reproduction, 25, 519-527.
- Li DK *et al.* (2010b) Relationship between urine bisphenol-A level and declining male sexual function. Journal of Andrology, 31, 500-506.
- Li DK *et al.* (2011) Urine bisphenol-A (BPA) level in relation to semen quality. Fertility and Sterility, 95, 625-630.
- Li J et al. (2008) A two-hybrid yeast assay to quantify the effects of xenobiotics on retinoid X receptor-mediated gene expression. Toxicol Lett, 176: 198-206
- Li J *et al.* (2010) In vitro profiling of endocrine disrupting effects of phenols. Toxicol In Vitro, 24: 201-207
- Lin CY, Shen FY, Lian GW, Chien KL, Sung FC, Chen PC and Su TC. (2015) Association between levels of serum bisphenol A, a potentially harmful chemical in plastic containers, and carotid artery intima-media thickness in adolescents and young adults. Atherosclerosis, 241 (2):657-63.
- Liu J, Yu P, Qian W, Li Y, Zhao J, Huan F, Wang J, Xiao H. (2013) Perinatal bisphenol A exposure and adult glucose homeostasis: identifying critical windows of exposure. PLoS One, 8(5):e64143.
- Liu Y, Peterson KE. (2015) Maternal Exposure to Synthetic Chemicals and Obesity in the Offspring: Recent Findings. Curr Environ Health Rep, 2(4):339-47

- Longacre TA and Bartow SA. (1986) A correlative morphologic study of human breast and endometrium in the menstrual cycle. Am J Surg Pathol, 10 (6):382-93.
- Losa-Ward SM *et al.* (2012) Disrupted organization of RFamide pathways in the hypothalamus is associated with advanced puberty in female rats neonatally expodes to BPA. Biol Reprod, 87(2):28
- Macon MB, Fenton SE. (2013) Endocrine disruptors and the breast: early life effects and later life disease. J Mammary Gland Biol Neoplasia, 18:43-61
- Maffini MV, Soto AM, Calabro JM, Ucci AA and Sonnenschein C. (2004) The stroma as a crucial target in rat mammary gland carcinogenesis. J Cell Sci, 117 (Pt 8):1495-502.
- Magruder HT, Quinn JA, Schwartzbauer JE, Reichner J, Huang A and Filardo EJ. (2014) The G protein-coupled estrogen receptor-1, GPER-1, promotes fibrillogenesis via a Shc-dependent pathway resulting in anchorage-independent growth. Horm Cancer, 5 (6):390-404.
- Mahoney MM and Padmanabhan V. (2010) Developmental programming: impact of fetal exposure to endocrine-disrupting chemicals on gonadotropin-releasing hormone and estrogen receptor mRNA in sheep hypothalamus. Toxicol Appl Pharmacol, 247: 98-104.
- Makaji E, Raha S, Wade MG and Holloway AC. (2011) Effect of environmental contaminants on Beta cell function. Int J Toxicol, 30 (4):410-8.
- Mallepell S, Krust A, Chambon P and Brisken C. (2006) Paracrine signaling through the epithelial estrogen receptor alpha is required for proliferation and morphogenesis in the mammary gland. Proc Natl Acad Sci U S A, 103 (7):2196-201.
- Mallo M, Alonso CR. (2013) The regulation of Hox gene expression during animal development. Development, 140:3951-63
- Mansur A *et al.* (2016) Does BPA alter steroid hormone synthesis in human granulosa cells in vitro? Hum Reprod, 31(7):1562-9.
- Mao Z, Xia W, Chang H, Huo W, Li Y, Xu S. (2015) Paternal BPA exposure in early life alters Igf2 epigenetic status in sperm and induces pancreatic impairment in rat offspring. Toxicol Lett, 238(3):30-8.
- Marchese S, Silva E. (2012) Disruption of 3D MCF-12A breast cell cultures by estrogensan in vitro model for ER-mediated changes indicative of hormonal carcinogenesis. PLoS One, 7(10):e45767
- Margueron R, Reinberg D. (2011) The Polycomb complex PRC2 and its mark in life. Nature, 469:343-9
- Markey CM, Wadia PR, Rubin BS, Sonnenschein C, Soto AM. (2005) Long-term effects of fetal exposure to low doses of the xenoestrogen bisphenol-A in the female mouse genital tract. Biol Reprod, 72:1344-51
- Marques-Pinto A and Carvalho D. (2013) Human infertility: are endocrine disruptors to blame? Endocr Connect, 2(3): R15–R29.
- Masters JR, Drife JO and Scarisbrick JJ. (1977) Cyclic Variation of DNA synthesis in human breast epithelium. J Natl Cancer Inst, 58 (5):1263-5.
- Matsuda S *et al.* (2010) Changes in brain monoamine levels in neonatal rats exposed to bisphenol A at low doses. Chemosphere, 78: 894-906.

- Matsuda S *et al.* (2013) Perinatal exposure to bisphenol A enhances contextual fear memory and affects the serotoninergic system in juvenile female mice. Horm Behav, 63(5):709-16.
- Mauvais-Jarvis F *et al.* (2013). The role of estrogens in control of energy balance and glucose homeostasis. Endocr Rev 34(3): 309-338.
- Meeker JD *et al.* (2010a) Urinary Bisphenol A Concentrations in Relation to Serum Thyroid and Reproductive Hormone Levels in Men from an Infertility Clinic. Environmental Science & Technology, 44, 1458-1463.
- Meeker JD *et al.* (2010b) Semen quality and sperm DNA damage in relation to urinary bisphenol A among men from an infertility clinic. Reproductive Toxicology, 30, 532-539.
- Melzer D, Rice NE, Lewis C, Henley WE and Galloway TS. (2010) Association of urinary bisphenol a concentration with heart disease: evidence from NHANES 2003/06. PLoS One, 5 (1):e8673.
- Menale C, Piccolo MT, Cirillo G, Calogero RA, Papparella A, Mita L, Del Giudice EM, Diano N, Crispi S, Mita DG. (2015) Bisphenol A effects on gene expression in adipocytes from children: association with metabolic disorders. J Mol Endocrinol, 54(3):289-303.
- Menale C, Grandone A, Nicolucci C, Cirillo G, Crispi S, Di Sessa A, Marzuillo P, Rossi S, Mita DG, Perrone L, Diano N, Miraglia Del Giudice E. (2016) Bisphenol A is associated with insulin resistance and modulates adiponectin and resistin gene expression in obese children. Pediatr Obes, Epub ahead of print
- Menard S et al. (2014a) Food intolerance at adulthood after perinatal exposure to the endocrine disruptor bisphenol A. FASEB J, 28(11):4893-900.
- Menard S *et al.* (2014b). Perinatal exposure to a low dose of bisphenol A impaired systemic cellular immune response and predisposes young rats to intestinal parasitic infection. PLoS One, 9(11):e112752.
- Mendiola J *et al.* (2010) Are environmental levels of bisphenol a associated with reproductive function in fertile men? Environmental Health Perspectives, 118, 1286-1291.
- Mendoza-Rodríguez CA *et al.* (2011) Administration of bisphenol A to dams during perinatal period modifies molecular and morphological reproductive parameters of the offspring. Reproductive Toxicology, 31, 177-183.
- Meyers R. (1983) DES, the bitter pill. Seaview/Putnam; New York.
- Miao M et al. (2011) In Utero Exposure to Bisphenol-A and Anogenital Distance of Male Offspring. Birth Defects Research (Part A), 91, 867-872.
- Miyawaki J et al. (2007) Perinatal and postnatal exposure to bisphenol a increases adipose tissue mass and serum cholesterol level in mice. Journal of Atherosclerosis and Thrombosis, 14 (5): 245-252.
- Mizuno K and Giese KP. (2010). Towards a molecular understanding of sex differences in memory formation. Trends Neurosci, 33(6):285-91.
- Mlynarcikova A *et al.* (2005) Alterations in steroid hormone production by porcine ovarian granulosa cells caused by bisphenol A and bisphenol A dimethacrylate. Mol. Cell. Endocrinol, 244:57–62.

- Moghaddam HS, Samarghandian S, Farkhondeh T. (2015) Effect of bisphenol A on blood glucose, lipid profile and oxidative stress indices in adult male mice. Toxicol Mech Methods, 25(7):507-13.
- Mok-Lin E *et al.* (2010) Urinary bisphenol A concentrations and ovarian response among women undergoing IVF. International Journal of Andrology, 33, 385-393.
- Monje L *et al.* (2009) Neonatal exposure to bisphenol A alters estrogen-dependent mechanisms governing sexual behavior in the adult female rat. Reproductive Toxicology, 28 (4): 435-442.
- Monje L et al. (2010) Exposure of neonatal female rats to bisphenol A disrupts hypothalamic LHRH pre-mRNA processing and estrogen receptor alpha expression in nuclei controlling estrous cyclicity. Reprod Toxicol, 30(4):625-34.
- Moon MK, Jeong IK, Jung Oh T, Ahn HY, Kim HH, Park YJ, Jang HC, Park KS. (2015) Long-term oral exposure to bisphenol A induces glucose intolerance and insulin resistance. J Endocrinol, 226(1):35-42.
- Moore-Ambriz TR *et al* (2015) Exposure to bisphenol A in young adult mice does not alter ovulation but does alter the fertilization ability of oocytes. Toxicol Appl Pharmacol, 289(3):507-14.
- Moral R, Wang R, Russo IH, Lamartiniere CA, Pereira J, Russo J. (2008) Effect of prenatal exposure to the endocrine disruptor bisphenol A on mammary gland morphology and gene expression signature. J Endocrinol, 196:101-12
- Munoz-de-Toro M, Markey CM, Wadia PR, Luque EH, Rubin BS, Sonnenschein C, Soto AM. (2005) Perinatal exposure to bisphenol-A alters peripubertal mammary gland development in mice. Endocrinology, 146:4138-47
- Murray TJ, Maffini MV, Ucci AA, Sonnenschein C, Soto AM. (2007) Induction of mammary gland ductal hyperplasias and carcinoma in situ following fetal bisphenol A exposure. Reprod Toxicol, 23:383-90
- Nagao T *et al.* (2002) Low-dose bisphenol A does not affect reproductive organs in estrogensensitive C57BL/6N mice exposed at the sexually mature, juvenile, or embryonic stage. Reproductive Toxicology, 16, 123-130.
- Nah WH *et al.* (2011). Effects of early prepubertal exposure to bisphenol A on the onset of puberty, ovarian weights, and estrous cycle in female mice. Clin Exp Reprod Med, 38(2), 75-81.
- Nakamura D *et al.* (2010) Bisphenol A may cause testosterone reduction by adversely affecting both testis and pituitary systems similar to estradiol. Toxicology Letters, 194, 16–25.
- Nakamura K et al. (2012) Prenatal and lactational exposure to low-doses of bisphenol A alters adult mice behavior. Brain Dev, 34(1):57-63.
- National Toxicology Program (NTP). (2010) NTP Historical Controls for NTP-2000 Diet [online database]. <a href="http://ntp.niehs.nih.gov/?objectid=92E705C7-F1F6-975E-72D23026B1645EB9">http://ntp.niehs.nih.gov/?objectid=92E705C7-F1F6-975E-72D23026B1645EB9</a> [accessed on March 2011].
- Nativelle-Serpentini C *et al.* (2003) Aromatase activity modulation by lindane and bisphenol-A in human placental JEG-3 and transfected kidney E293 cells. Toxicol In Vitro, 17(4):413-22.
- Naulé L et al. (2014) Neuroendocrine and behavioral effects of maternal exposure to oral bisphenol A in female mice. J Endocrinol, 10;220(3):375-88.

- Naulé L and Mhaouty-Kodja S (2016). [The neural oestrogen receptor beta, a new player in female pubertal maturation]. Med Sci (Paris), 32(5): 452-455. French
- Navarro VM *et al.* (2009). Persistent impairment of hypothalamic KiSS-1 system after exposures to estrogenic compounds at critical periods of brain sex differentiation. Endocrinology, 150, 2359–2367
- Newbold RR, Jefferson WN, Padilla-Banks E (2007) Long-term adverse effects of neonatal exposure to bisphenol A on the murine female reproductive tract. Reproductive Toxicology, 24, 253-258.
- Newbold RR, Jefferson WN, Padilla-Banks E (2009) Prenatal Exposure to Bisphenol A at Environmentally Relevant Doses Adversely Affects the Murine Female Reproductive Tract Later in Life. Environmental Health Perspectives, 117, 879-885.
- Nikaido Y *et al.* (2004) Effects of maternal xenoestrogen exposure on development of the reproductive tract and mammary gland in female CD-1 mouse offspring. Reproductive Toxicolog,y 18, 803-811.
- Nikaido Y *et al.* (2005) Effects of prepubertal exposure to xenoestrogen on development of estrogen target organs in female CD-1 mice. In Vivo, 19, 487-494.
- Ning G, Bi Y, Wang T, Xu M, Xu Y, Huang Y, Li M, Li X, Wang W, Chen Y, Wu Y, Hou J, Song A, Liu Y and Lai S. (2011) Relationship of urinary bisphenol A concentration to risk for prevalent type 2 diabetes in Chinese adults: a cross-sectional analysis. Ann Intern Med, 155 (6):368-74.
- Nishizawa H et al. (2005) Effects of exposure in utero to bisphenol A on the expression of aryl hydrocarbon receptor, related factors, and xenobiotic metabolizing enzymes in murine embryos. J Reprod Dev, 51(5):593–605
- Norden (2014) The Cost of Inaction A socioeconomic analysis of costs linked to effects of endocrine disrupting substances on male reproductive health. Nordic Council of Ministers.

  TemaNord :557

  http://www.miliodirektoratet.no/Documents/publikasioner/M289/M289.pdf
- NTP (1985) Bisphenol A: reproduction and fertility assessment in CD-1 mice when administered in the feed. NTP-85-192. Research Triangle Park, NC.
- OECD (2013). Guidance document on developing and assessing adverse outcome pathways. Series on Testing and Assessment No. 184. Available at <a href="http://www.oecd.org/officialdocuments/publicdisplaydocumentpdf/?cote=env/jm/mono(2013)6&doclanguage=en">http://www.oecd.org/officialdocuments/publicdisplaydocumentpdf/?cote=env/jm/mono(2013)6&doclanguage=en</a> Appendix I. Collection of working definitions. Available at <a href="http://www.oecd.org/chemicalsafety/testing/49963576.pdf">http://www.oecd.org/chemicalsafety/testing/49963576.pdf</a>
- Oehlmann J *et al.* (2006) Bisphenol A induces superfeminization in the ramshorn snail Marisa cornuarietis(Gastropoda: Prosobranchia) at environmentally relevant concentrations. Environ Health Perspect, 114 Suppl 1:127-133.
- Oehlmann J *et al.* (2009). A critical analysis of the biological impacts of plasticizers on wildlife. Philosophical Transactions of the Royal Society B: Biological Sciences, 364(1526):2047-2062.
- Ohlstein JF, Strong AL, McLachlan JA, Gimble JM, Burow ME, Bunnell BA. (2014) Bisphenol A enhances adipogenic differentiation of human adipose stromal/stem cells. J Mol Endocrinol, 53(3):345-53.

- Okada A and Kai O (2008a) Effects of estradiol-17beta and bisphenol A administered chronically to mice throughout pregnancy and lactation on the male pups reproductive system. Asian Journal of Andrology, 10, 271-276.
- Okada H, Tokunaga T, Liu X, Takayanagi S, Matsushima A, Shimohigashi Y. (2008b) Direct evidence revealing structural elements essential for the high binding ability of bisphenol A to human estrogen-related receptor-gamma. Environ Health Perspect, 116:32-8
- Paap E, Broeders MJ, van Schoor G, Otten JD, Verbeek AL. (2008) Large increase in a Dutch woman's lifetime risk of developing breast cancer. Eur J Cancer, 44(11):1485-7.
- Packard MG, Kohlmaier JR, Alexander GM. (1996) Posttraining intrahippocampal estradiol injections enhance spatial memory in male rats: interaction with cholinergic systems. Behav Neurosci. 110: 626–632.
- Page DL, Dupont WD, Rogers LW, Jensen RA, Schuyler PA (1995) Continued local recurrence of carcinoma 15-25 years after a diagnosis of low grade ductal carcinoma in situ of the breast treated only by biopsy. Cancer, 76, 1197-1200.
- Palanza PL, Howdeshell KL, Parmigiani S, vom Saal FS. (2002) Exposure to a low dose of bisphenol A during fetal life or in adulthood alters maternal behavior in mice. Environ Health Perspect, 110 Suppl 3:415-22.
- Palanza P et al. (2008). Effects of developmental exposure to bisphenol A on brain and behavior in mice. Environmental Research, 108 (2): 150-157.
- Palmer JR, Hatch EE, Rosenberg CL, Hartge P, Kaufman RH, Titus-Ernstoff L, Noller KL, Herbst AL, Rao RS, Troisi R, Colton T, Hoover RN. (2002) Risk of breast cancer in women exposed to diethylstilbestrol in utero: prelimiinary results (United States) Cancer Causes Control, 13:753–758.
- Palmer JR, Wise LA, Hatch EE, Troisi R, Titus-Ernstoff L, Strohsnitter W, Kaufman R, Herbst AL, Noller KL, Hyer M, Hoover RN. (2006) Prenatal diethylstilbestrol exposure and risk of breast cancer. Cancer Epidemiol Biomarkers Prev, 15:1509–1514.
- Patisaul HB and Fortino AE PE. (2007) Differential disruption of nuclear volume and neuronal phenotype in the preoptic area by neonatal exposure to genistein and bisphenol-A. Neurotoxicology (Amsterdam), 28: 1-12.
- Patisaul HB *et al.* (2009). Impact of neonatal exposure to the ERalpha agonist PPT, bisphenol-A or phytoestrogens on hypothalamic kisspeptin fiber density in male and female rats. Neurotoxicology, 30(3), 350-7.
- Patisaul HB *et al.* (2014) Soy but not bisphenol A (BPA) induces hallmarks of polycystic ovary syndrome (PCOS) and related metabolic co-morbidities in rats. Reprod Toxicol, 49:209-18.
- Pawluski JL et al. (2009) Effects of steroid hormones on neurogenesis in the hippocampus of the adult female rodent during the estrous cycle, pregnancy, lactation and aging. Front Neuroendocrinol, 30(3):343-57.
- Peluso ME, Munnia A, Ceppi M. (2014) Bisphenol-A exposures and behavioural aberrations: median and linear spline and meta-regression analyses of 12 toxicity studies in rodents. Toxicology, 325:200-8.

- Pereira-Fernandes *et al.* (2013) Evaluation of a screening system for obesogenic compounds: screening of endocrine disrupting compounds and evaluation of the PPAR dependency of the effect. PLoS One, 8(10):e7748.
- Pereira-Fernandes A *et al.* (2014) Toxicogenomics in the 3T3-L1 cell line, a new approach for screening of obesogenic compounds. Toxicol Sci, 140(2):352-63..
- Peretz J *et al.* (2011) Bisphenol A impairs follicle growth, inhibits steroidogenesis, and downregulates rate-limiting enzymes in the estradiol biosynthesis pathway. Toxicol Sci, 119(1):209-17.
- Peretz J and Flaws JA. (2013) Bisphenol A down-regulates rate-limiting Cyp11a1 to acutely inhibit steroidogenesis in cultured mouse antral follicles. Toxicol Appl Pharmacol, 271(2):249-56.
- Picot M et al. (2014) Vulnerability of the neural circuitry underlying sexual behavior to chronic adult exposure to oral bisphenol a in male mice. Endocrinology, 155(2):502-12.
- Picot M *et al.* (2016) Neural Androgen Receptor Deletion Impairs the Temporal Processing of Objects and Hippocampal CA1-Dependent Mechanisms. PLoS One, 11(2):e0148328.
- Poimenova A *et al.* (2010) Corticosterone-regulated actions in the rat brain are affected by perinatal exposure to low dose of bisphenol A. Neuroscience, 167 (3): 741-749.
- Prossnitz ER, Maggiolini M. (2009) Mechanisms of estrogen signaling and gene expression via GPR30. Mol Cell Endocrinol, 308(1-2):32-8
- Pupo M, Pisano A, Lappano R, Santolla MF, De Francesco EM, Abonante S, Rosano C, Maggiolini M. (2012) Bisphenol A induces gene expression changes and proliferative effects through GPER in breast cancer cells and cancer-associated fibroblasts. Environ Health Perspect, 120:1177-82
- Qin XY, Fukuda T, Yang L, Zaha H, Akanuma H, Zeng Q, Yoshinaga J, Sone H. (2012) Effects of bisphenol A exposure on the proliferation and senescence of normal human mammary epithelial cells. Cancer Biol Ther, 13:296-306
- Quesada I, Tudurí E, Ripoll C, Nadal A. (2008) Physiology of the pancreatic alpha-cell and glucagon secretion: role in glucose homeostasis and diabetes. J Endocrinol, 199:5-19.
- Ranciere F, Lyons JG, Loh VH, Botton J, Galloway T, Wang T, Shaw JE and Magliano DJ. (2015) Bisphenol A and the risk of cardiometabolic disorders: a systematic review with meta-analysis of the epidemiological evidence. Environ Health, 14:46.
- Rebuli ME *et al.* (2014) Investigation of the effects of subchronic low dose oral exposure to bisphenol A (BPA) and ethinyl estradiol (EE) on estrogen receptor expression in the juvenile and adult female rat hypothalamus. Toxicol Sci, 140(1):190-203.
- Reed CE, Fenton SE. (2013) Exposure to diethylstilbestrol during sensitive life stages: a legacy of heritable health effects. Birth Defects Res C Embryo Today, 99:134-46
- Rijk I, van Duursen M, van den Berg M. (2016) Health cost that may be associated with Endocrine Disrupting Chemicals An inventory, evaluation and way forward to assess the potential socio-economic impact of EDC-associated health effects in the EU. IRAS, <a href="https://www.uu.nl/sites/default/files/rijk et al 2016">https://www.uu.nl/sites/default/files/rijk et al 2016</a> report iras health cost associated with edcs 3.pdf

- Rivera OE *et al.* (2011) Neonatal exposure to bisphenol A or diethylstilbestrol alters the ovarian follicular dynamics in the lamb. Reprod Toxicol, 32(3):304-12.
- Rivera OE *et al.* (2015) Neonatal exposure to xenoestrogens impairs the ovarian response to gonadotropin treatment in lambs. Reproduction, 149(6):645-55.
- Robinson GW, Karpf AB and Kratochwi, K. (1999) Regulation of mammary gland development by tissue interaction. J. Mam. Gland Biol.Neoplasia, 4:, 9-19.
- Rodríguez HA *et al.* (2010) Neonatal exposure to bisphenol A reduces the pool of primordial follicles in the rat ovary. Reprod Toxicol, 30(4):550-7.
- Roepke TA *et al.* 2005 Jan. Estradiol and endocrine disrupting compounds adversely affect development of sea urchin embryos at environmentally relevant concentrations. Aquatic Toxicol, 71(2):155-173.
- Ronn M, Lind L, Orberg J, Kullberg J, Soderberg S, Larsson A, Johansson L, Ahlstrom H and Lind PM. (2014) Bisphenol A is related to circulating levels of adiponectin, leptin and ghrelin, but not to fat mass or fat distribution in humans. Chemosphere, 112:42-8.
- Roof RL and Havens MD. (1992) Testosterone improves maze performance and induces development of a male hippocampus in females. Brain Res, 572(1-2):310-3.
- Rorsman P, Braun M. (2013) Regulation of insulin secretion in human pancreatic islets. Annu Rev Physiol, 75:155-79.
- Ruan H and Dong LQ. (2016) Adiponectin signaling and function in insulin target tissues. J Mol Cell Biol 8 (2):101-9.
- Rubin BS *et al.* (2001) Perinatal exposure to low doses of bisphenol A affects body weight, patterns of estrous cyclicity, and plasma LH levels. Environmental Health Perspectives, 109, 675-680.
- Rubin BS *et al.* (2006) Evidence of altered brain sexual differentiation in mice exposed perinatally to low, environmentally relevant levels of bisphenol A. Endocrinology, 147 (8): 3681-3691.
- Russo J and Russo IH. (1980) Susceptibility of the mammary gland to carcinogenesis. II. Pregnancy interruption as a risk factor in tumor incidence. Am J Pathol, 100 (2):497-512.
- Russo J, Gusterson BA, Rogers AE, Russo IH, Wellings SR, van Zwieten MJ. (1990) Comparative study of human and rat mammary tumorigenesis. Lab Invest 62:244-78
- Russo IH, Russo J. (1996) Mammary gland neoplasia in long-term rodent studies. Environ Health Perspect 104:938-67
- Ryan BC *et al.* (2010a) *In Utero* and Lactational Exposure to Bisphenol A, In Contrast to Ethinyl Estradiol, Does Not Alter Sexually Dimorphic Behavior, Puberty, Fertility, and Anatomy of Female Rats. Toxicological Sciences 114: 133-148.
- Ryan KK, Haller AM, Sorrell JE, Woods SC, Jandacek RJ, Seeley RJ. (2010b) Perinatal exposure t bisphenol-A and the development of metabolic syndrome in CD-1 mice. Endocrinology 151, 2603-2612.
- Sakaue M *et al.* (2001) Bisphenol-A affects spermatogenesis in the adult rat even at low dose. Journal of Occupational Health 43, 185-190.

- Salian S *et al.* (2009a) Impairment in protein expression profile of testicular steroid receptor coregulators in male rat offspring perinatally exposed to Bisphenol A. Life Sciences 85, 11-18.
- Salian S *et al.* (2009b) Neonatal exposure of male rats to Bisphenol A impairs fertility and expression of sertoli cell junctional proteins in the testis. Toxicology 265, 56-67.
- Salian S *et al.* (2009c) Perinatal exposure of rats to Bisphenol A affects the fertility of male offspring. Life Sciences 85, 742-752.
- Savabieasfahani M *et al.* (2006) Developmental programming: Differential effects of prenatal exposure to bisphenol-A or methoxychlor on reproductive function. Endocrinology 147, 5956-5966.
- Savastano S, Tarantino G, D'Esposito V, Passaretti F, Cabaro S, Liotti A, Liguoro D, Perruolo G, Ariemma F, Finelli C, Beguinot F, Formisano P, Valentino R. (2015) Bisphenol-A plasma levels are related to inflammatory markers, visceral obesity and insulin-resistance: a cross-sectional study on adult male population. J Transl Med 13:169
- Seachrist DD, Bonk KW, Ho SM, Prins GS, Soto AM, Keri RA. (2016) A review of the carcinogenic potential of bisphenol A. Reprod Toxicol 59:167-82
- Segal D, Makris SL, Kraft AD, Bale AS, Fox J, Gilbert M, Bergfelt DR, Raffaele KC, Blain RB, Fedak KM, Selgrade MK, Crofton KM. (2015). Evaluation of the ToxRTool's ability to rate the reliability of toxicological data for human health hazard assessments. Regul Toxicol Pharmacol. 72(1):94-101.
- Seki H, Hayashida T, Jinno H, Hirose S, Sakata M, Takahashi M, Maheswaran S, Mukai M, Kitagawa Y. (2012) HOXB9 expression promoting tumor cell proliferation and angiogenesis is associated with clinical outcomes in breast cancer patients. Ann Surg Oncol 19:1831-40
- Sengupta S, Obiorah I, Maximov PY, Curpan R, Jordan VC. (2013) Molecular mechanism of action of bisphenol and bisphenol A mediated by oestrogen receptor alpha in growth and apoptosis of breast cancer cells. Br J Pharmacol 169:167-78.
- Shankar A, Teppala S. (2011) Relationship between urinary bisphenol A levels and diabetes mellitus. J Clin Endocrinol Metab, 96(12):3822-6.
- Shankar A, Teppala S. (2012) Urinary bisphenol A and hypertension in a multi-ethnic sample of US adults. J Environ Public Health 481641.
- Shapiro GD, Dodds L, Arbuckle TE, Ashley-Martin J, Fraser W, Fisher M, Taback S, Keely E, Bouchard MF, Monnier P, Dallaire R, Morisset A, Ettinger AS. (2015) Exposure to phthalates, bisphenol A and metals in pregnancy and the association with impaired glucose tolerance and gestational diabetes mellitus: The MIREC study. Environ Int. 83:63-71
- Shrestha B, Ansari KI, Bhan A, Kasiri S, Hussain I, Mandal SS. (2012) Homeodomain-containing protein HOXB9 regulates expression of growth and angiogenic factors, facilitates tumor growth in vitro and is overexpressed in breast cancer tissue. FEBS J 279:3715-26
- Signorile PG *et al.* (2010) Pre-natal exposure of mice to bisphenol A elicits an endometriosis-like phenotype in female offspring. General and Comparative Endocrinology 168, 318-325.

- Silver MK, O'Neill MS, Sowers MR and Park SK. (2011) Urinary bisphenol A and type-2 diabetes in U.S. adults: data from NHANES 2003-2008. PLoS One 6: 10
- Simpson ER, Mahendroo MS, Means GD, Kilgore MW, Hinshelwood MM, Graham-Lorence S, Amarneh B, Ito Y, Fisher CR, Michael MD, *et al.* (1994) Aromatase cytochrome P450, the enzyme responsible for estrogen biosynthesis. Endocr Rev. 15(3):342-55
- Skorupskaite K *et al.* (2014) The kisspeptin-GnRH pathway in human reproductive health and disease. Hum Reprod Update. 20(4):485–500.
- Sleeckx N, de Rooster NH, Veldhuis Kroeze EJ, Van Ginneken C and Van Brantegem L. 2011. Canine mammary tumours, an overview. Reprod Domest Anim 46 (6):1112-31.
- Small CM, Manatunga AK, Klein M, Feigelson HS, Dominguez CE, McChesney R, Marcus M. (2006) Menstrual cycle characteristics: associations with fertility and spontaneous abortion. Epidemiology 17(1):52-60.
- Smith MS, Freeman ME, Neill JD. (1975). The control of progesterone secretion during the estrous cycle and early pseudopregnancy in the rat: prolactin, gonadotropin and steroid levels associated with rescue of the corpus luteum of pseudopregnancy. Endocrinology. 96(1):219–226
- Sohoni *et al.* (1998) Several environmental estrogens are also anti-androgens. J Endocrinol 158: 327-339.
- Somm E, Schwitzgebel VM, Toulotte A, Cederroth CR, Combescure C, Nef S, Aubert ML and Huppi PS. 2009. Perinatal exposure to bisphenol a alters early adipogenesis in the rat. *Environ Health Perspect* 117 (10):1549-55.
- Song KH *et al.* (2002) Endocrine disrupter bisphenol a induces orphan nuclear receptor Nur77 gene expression and steroidogenesis in mouse testicular Leydig cells. 316 Endocrinology 143: 2208-2215
- Song L, Xia W, Zhou Z, Li Y, Lin Y, Wei J, Wei Z, Xu B, Shen J, Li W, Xu S. (2012) Low-level phenolic estrogen pollutants impair islet morphology and  $\beta$ -cell function in isolated rat islets. J Endocrinol. 215(2):303-11.
- Song Y, Hauser R, Hu FB, Franke AA, Liu S, Sun Q. (2014) Urinary concentrations of bisphenol A and phthalate metabolites and weight change: a prospective investigation in US women.Int J Obes (Lond). 38(12):1532-7.
- Song H, Zhang T, Yang P, Li M, Yang Y, Wang Y, Du J, Pan K and Zhang K. (2015). Low doses of bisphenol A stimulate the proliferation of breast cancer cells via ERK1/2/ERRgamma signals. Toxicol In Vitro 30 (1 Pt B):521-8.
- Song Y, Chou EL, Baecker A, You NC, Song Y, Sun Q and Liu S. (2016) Endocrine-disrupting chemicals, risk of type 2 diabetes, and diabetes-related metabolic traits: A systematic review and meta-analysis. J Diabetes 8 (4):516-32.
- Soriano S, Alonso-Magdalena P, García-Arévalo M, Novials A, Muhammed SJ, Salehi A, Gustafsson JA, Quesada I, Nadal A. (2012) Rapid insulinotropic action of low doses of bisphenol-A on mouse and human islets of Langerhans: role of estrogen receptor β. PLoS One. 7(2):e31109.
- Soto AM, Vandenberg LN, Maffini MV, Sonnenschein C. (2008) Does breast cancer start in the womb? Basic Clin Pharmacol Toxicol 102:125-33

- Soto AM, Brisken C, Schaeberle C, Sonnenschein C. (2013) Does cancer start in the womb? altered mammary gland development and predisposition to breast cancer due to *in utero* exposure to endocrine disruptors. J Mammary Gland Biol Neoplasia 18:199-208
- Sparmann A, van Lohuizen M. (2006) Polycomb silencers control cell fate, development and cancer. Nat Rev Cancer 6:846-56
- Sprague BL, Trentham-Dietz A, Hedman CJ, Wang J, Hemming JD, Hampton JM, Buist DS, Aiello Bowles EJ, Sisney GS and Burnside ES. (2013) Circulating serum xenoestrogens and mammographic breast density. Breast Cancer Res 15 (3):R45.
- Stump DG *et al.* (2010) Developmental neurotoxicity study of dietary bisphenol A in Sprague-Dawley rats. Toxicol Sci. 115 (1): 167-182.
- Stumvoll M, Goldstein BJ, van Haeften TW. (2005) Type 2 diabetes: principles of pathogenesis and therapy. Lancet. 365 (9467): 1333-46
- Sugiura-Ogasawara M *et al.* (2005) Exposure to bisphenol A is associated with recurrent miscarriage. Human Reproduction 20, 2325-2329.
- Sui Y *et al.* (2012) Bisphenol A and its analogues activate human pregnane X receptor. Environ Health Perspect. 120(3):399-405.
- Sun Q, Cornelis MC, Townsend MK, Tobias DK, Eliassen AH, Franke AA, Hauser R, Hu FB. (2014) Association of urinary concentrations of bisphenol A and phthalate metabolites with risk of type 2 diabetes: a prospective investigation in the Nurses' Health Study (NHS) and NHSII cohorts. Environ Health Perspect. 122(6):616-23.
- Susiarjo M *et al.* (2007). Bisphenol A Exposure In Utero Disrupts Early Oogenesis in the Mouse. PLoS Genetics, 3(1): e5.
- Takagi H *et al.* (2004) Lack of maternal dietary exposure effects of bisphenol A and nonylphenol during the critical period for brain sexual differentiation on the reproductive/endocrine systems in later life. Arch Toxicol. 78:97–105.
- Takahashi O & Oishi S (2003) Testicular toxicity of dietarily or parenterally administered bisphenol A in rats and mice. Food and Chemical Toxicology 41 (7), 1035-44.
- Takayanagi S, Tokunaga T, Liu X, Okada H, Matsushima A, Shimohigashi Y. (2006) Endocrine disruptor bisphenol A strongly binds to human estrogen-related receptor gamma (ERRgamma) with high constitutive activity. Toxicol Lett 167:95-105
- Takeuchi T *et al.*(2004). Positive relationship between androgen and the endocrine disruptor, bisphenol A, in normal women and women with ovarian dysfunction. Endocrine Journal 51: 165-169.
- Tan BLL *et al.* (2003) Assessment of pubertal development in juvenile male rats after sub-acute exposure to bisphenol A and nonylphenol. Toxicology Letters 143, 261-270.
- Tanaka M, Sokabe M. (2013) Bidirectional modulatory effect of 17ß-estradiol on NMDA receptors via ERa and ERß in the dentate gyrus of juvenile male rats. Neuropharmacology 75: 262–273.
- Tanos T, Sflomos G, Echeverria PC, Ayyanan A, Gutierrez M, Delaloye JF, Raffoul W, Fiche M, Dougall W, Schneider P, Yalcin-Ozuysal O and Brisken C. (2013) Progesterone/RANKL is a major regulatory axis in the human breast. Sci Transl Med 5 (182):182ra55.

- Taylor JA, Vom Saal FS, Welshons WV, Drury B, Rottinghaus G, Hunt PA, Toutain PL, Laffont CM, and Vandevoort CA. (2011) Similarity of bisphenol A pharmacokinetics in rhesus monkeys and mice: relevance for human exposure. Environmental Health Perspectives 119 (4): 422-430.
- Tharp AP, Maffini MV, Hunt PA, VandeVoort CA, Sonnenschein C, Soto AM. (2012) Bisphenol A alters the development of the rhesus monkey mammary gland. Proc Natl Acad Sci U S A 109:8190-5
- Tian YH *et al.* (2010) Prenatal and postnatal exposure to bisphenol A induces anxiolytic behaviors and cognitive deficits in mice. Synapse 64 (6): 432-439.
- Tiano JP, Mauvais-Jarvis F. (2012) Importance of oestrogen receptors to preserve functional  $\beta$ -cell mass in diabetes. Nat Rev Endocrinol. 8(6): 342-51
- Timms BG *et al.* (2005) Estrogenic chemicals in plastic and oral contraceptives disrupt development of the fetal mouse prostate and urethra. Proceedings of the National Academy of Sciences of the United States of America 102, 7014-7019.
- Tinwell H et al. (2002) Normal sexual development of two strains of rat exposed in utero to low doses of bisphenol A. Toxicological Sciences 68, 339-348.
- Topper YJ and Freeman CS. (1980) Multiple hormone interactions in the developmental biology of the mammary gland. Physiol Rev. 60(4):1049-106.
- Toyama Y et al. (2004a) Adverse effects of bisphenol A to spermiogenesis in mice and rats. Archivesin Histology and Cytology 64(4), 373-381.
- Toyama Y & Yuasa S (2004b) Effects of neonatal administration of  $17\beta$ -estradiol,  $\beta$ -estradiol 3-benzoate, or bisphenol A on mouse and rat spermatogenesis. Reproductive Toxicology 19, 181-188.
- Trabert B, Falk RT, Figueroa JD, Graubard BI, Garcia-Closas M, Lissowska J, Peplonska B, Fox SD and Brinton LA. (2014) Urinary bisphenol A-glucuronide and postmenopausal breast cancer in Poland. Cancer Causes Control 25 (12):1587-93.
- Trasande L, Zoeller RT, Hass U, Kortenkamp A, Grandjean P, Myers JP, DiGangi J, Hunt PM, Rudel R, Sathyanarayana S, Bellanger M, Hauser R, Legler J, Skakkebaek NE, Heindel JJ. (2016) Burden of disease and costs of exposure to endocrine disrupting chemicals in the European Union: an updated analysis. Andrology. 4(4):565-72.
- Tyl RW *et al.* (2002) Three-generation reproductive toxicity study of dietary bisphenol A in CD Sprague-Dawley rats. Toxicological Sciences 68, 121-146.
- Tyl RW *et al.* (2008) Two-generation reproductive toxicity study of dietary bisphenol a in CD-1 (Swiss) mice. Toxicological Sciences 104, 362-384.
- Valentino R, D'Esposito V, Passaretti F, Liotti A, Cabaro S, Longo M, Perruolo G, Oriente F, Beguinot F, Formisano P. (2013) Bisphenol-A impairs insulin action and upregulates inflammatory pathways in human subcutaneous adipocytes and 3T3-L1 cells. PLoS One. 8(12):e82099.
- Van Esterik JC, Dollé ME, Lamoree MH, van Leeuwen SP, Hamers T, Legler J, van der Ven LT. (2014) Programming of metabolic effects in C57BL/6JxFVB mice by exposure to bisphenol A during gestation and lactation. Toxicology. 321:40-52.

- Vandenberg LN, Maffini MV, Schaeberle CM, Ucci AA, Sonnenschein C, Rubin BS, Soto AM. (2008) Perinatal exposure to the xenoestrogen bisphenol-A induces mammary intraductal hyperplasias in adult CD-1 mice. Reprod Toxicol 26:210-9
- Vandenberg LN, Maffini MV, Wadia PR, Sonnenschein C, Rubin BS, Soto AM. (2007) Exposure to environmentally relevant doses of the xenoestrogen bisphenol-A alters development of the fetal mouse mammary gland. Endocrinology 148:116-27
- Varayoud J et al. (2011). Neonatal exposure to bisphenol A alters rat uterine implantation-associated gene expression and reduces the number of implantation sites. Endocrinology. 152(3), 1101-11.
- Veiga-Lopez A *et al.* (2014) Developmental programming: prenatal BPA treatment disrupts timing of LH surge and ovarian follicular wave dynamics in adult sheep. Toxicol Appl Pharmacol. 279(2):119-28
- Veiga-Lopez A, Moeller J, Sreedharan R, Singer K, Lumeng C, Ye W, Pease A, Padmanabhan V. (2016) Developmental programming: interaction between prenatal BPA exposure and postnatal adiposity on metabolic variables in female sheep. Am J Physiol Endocrinol Metab. 310(3):E238-47.
- Viberg H *et al.* (2011) Dose-dependent behavioural disturbances after a single neonatal Bisphenol A dose. Toxicology. 290(2-3):187-94.
- Volberg V, Harley K, Calafat AM, Dave V, McFadden J, Eskenazi B and Holland N. (2013) Maternal bisphenol a exposure during pregnancy and its association with adipokines in Mexican-American children. Environmental and Molecular Mutagenesis, 54, 621-628
- Wadia PR, Vandenberg LN, Schaeberle CM, Rubin BS, Sonnenschein C, Soto AM. (2007) Perinatal bisphenol A exposure increases estrogen sensitivity of the mammary gland in diverse mouse strains. Environ Health Perspect 115:592-8
- Wadia PR, Cabaton NJ, Borrero MD, Rubin BS, Sonnenschein C, Shioda T, Soto AM. (2013) Low-dose BPA exposure alters the mesenchymal and epithelial transcriptomes of the mouse fetal mammary gland. PLoS One 8:e63902
- Wang T, Li M, Chen B, Xu M, Xu Y, Huang Y, Lu J, Chen Y, Wang W, Li X, Liu Y, Bi Y, Lai S and Ning G. (2012) Urinary bisphenol A (BPA) concentration associates with obesity and insulin resistance. *J Clin Endocrinol Metab* 97 (2):E223-7.
- Wang J, Gupta A, Hu H, Chatterton RT, Clevenger CV and Khan SA. (2013) Comment on "Progesterone/RANKL is a major regulatory axis in the human breast". Sci Transl Med 5 (215):215le4.
- Wang J, Sun B, Hou M, Pan X, Li X. (2013) The environmental obesogen bisphenol A promotes adipogenesis by increasing the amount of  $11\beta$ -hydroxysteroid dehydrogenase type 1 in the adipose tissue of children. Int J Obes (Lond). 37(7):999-1005.
- Wang W et al. (2014a) In utero bisphenol A exposure disrupts germ cell nest breakdown and reduces fertility with age in the mouse. Toxicol Appl Pharmacol. 276(2):157-64.
- Wang X et al. (2014b) Bisphenol A enhances kisspeptin neurons in anteroventral periventricular nucleus of female mice. J Endocrinol. 221(2):201-13.

- Wang C *et al.* (2016) Impairment of object recognition memory by maternal bisphenol A exposure is associated with inhibition of Akt and ERK/CREB/BDNF pathway in the male offspring hippocampus. Toxicology. 341-343:56-64.
- Watanabe S *et al.* (2003) Imbalance of testosterone level in male offspring of rats perinatally exposed to bisphenol A. Industrial Health 41, 338-341.
- Watanabe M, Ohno S, Nakajin S. (2012) Effects of bisphenol A on the expression of cytochrome P450 aromatase (CYP19) in human fetal osteoblastic and granulosa cell-like cell lines. Toxicol Lett. 210(1):95-9.
- Watson CS *et al.* (2007) Nongenomic actions of low concentration estrogens and xenestrogens on multiple tissues. Mol Cell Endocrinol 274: 1-7
- Watson CS et al. (2010) Nongenomic signaling pathways of estrogen toxicity. Toxicol Sci 115(1): 1-11
- Weng YI, Hsu PY, Liyanarachchi S, Liu J, Deatherage DE, Huang YW, Zuo T, Rodriguez B, Lin CH, Cheng AL, Huang TH. (2010) Epigenetic influences of low-dose bisphenol A in primary human breast epithelial cells. Toxicol Appl Pharmacol 248:111-21
- Whitehead R, Guan H, Arany E, Cernea M, Yang K. (2016) Prenatal exposure to bisphenol A alters mouse fetal pancreatic morphology and islet composition. Horm Mol Biol Clin Investig. 25(3):171-9.
- WHO/International Programme on Chemical Safety. (2002) Global assessment of the state of the science of endocrine disruptors (Damstra T, Barlow S, Bergman A, Kavlock R, Van Der Kraak G, eds.). <a href="http://www.who.int/ipcs/publications/new">http://www.who.int/ipcs/publications/new</a> issues/endocrine disruptors/en/
- WHO (2016) Global report on diabetes. World Health Organization, Geneva <a href="http://apps.who.int/iris/bitstream/10665/204871/1/9789241565257">http://apps.who.int/iris/bitstream/10665/204871/1/9789241565257</a> eng.pdf
- Wolff MS *et al.* (2008). Environmental exposures and puberty in inner-city girls. Environmental Research 107: 393-400.
- Wolff MS *et al.* (2010) Investigation of Relationships between Urinary Biomarkers of Phytoestrogens, Phthalates, and Phenols and Pubertal Stages in Girls. Environmental Health Perspectives 118(7):1039-46.
- Wozniak AL, Bulayeva NN, Watson CS. (2005) Xenoestrogens at picomolar to nanomolar concentrations trigger membrane estrogen receptor-alpha-mediated Ca2+ fluxes and prolactin release in GH3/B6 pituitary tumor cells. Environ Health Perspect 113:431-9
- Xi W, Lee CK, Yeung WS, Giesy JP, Wong MH, Zhang X, Hecker M, Wong CK. (2011) Effect of perinatal and postnatal bisphenol A exposure to the regulatory circuits at the hypothalamus-pituitary-gonadal axis of CD-1 mice. Reprod Toxicol. 31(4):409-17.
- Xiong Q, Liu X, Shen Y, Yu P, Chen S, Hu J, Yu J, Li J, Wang HS, Cheng X, Hong K. (2015) Elevated serum bisphenol a level in patients with dilated cardiomyopathy. Int J Environ Res Public Health. 12(5):5329-37.
- Xu XH et al. (2010a) Perinatal exposure to bisphenol-A changes N-methyl-D-aspartate receptor expression in the hippocampus of male rat offspring. Environ Toxicol Chem. 29(1):176-81.

- Xu XH et al. (2010b) Perinatal exposure to bisphenol-A impairs learning-memory by concomitant down-regulation of N-methyl-D-aspartate receptors of hippocampus in male offspring mice. Hormones and Behavior 58 (2): 326-333.
- Xu X, Ye Y, Li T, Chen L, Tian D, Luo Q, Lu M. (2010c) Bisphenol-A rapidly promotes dynamic changes in hippocampal dendritic morphology through estrogen receptor-mediated pathway by concomitant phosphorylation of NMDA receptor subunit NR2B. Toxicol Appl Pharmacol. 249(2):188-96.
- Xu X, Li T, Luo Q, Hong X, Xie L, Tian D. (2011a) Bisphenol-A rapidly enhanced passive avoidance memory and phosphorylation of NMDA receptor subunits in hippocampus of young rats. Toxicol Appl Pharmacol. 255(2):221-8.
- Xu X *et al.* (2011b) Sex-specific influence of exposure to bisphenol-A between adolescence and young adulthood on mouse behaviors. Neuropharmacology. 61(4):565-73.
- Xu X et al. (2013a) Perinatal exposure to bisphenol-A inhibits synaptogenesis and affects the synaptic morphological development in offspring male mice. Chemosphere. 91(8):1073-81.
- Xu X et al. (2013b) Sex-specific effects of bisphenol-A on memory and synaptic structural modification in hippocampus of adult mice. Horm Behav. 63(5):766-75.
- Xu X et al. (2014a) Bisphenol A promotes dendritic morphogenesis of hippocampal neurons through estrogen receptor-mediated ERK1/2 signal pathway. Chemosphere. 96:129-37.
- Xu XB *et al.* (2014b) Bisphenol A regulates the estrogen receptor alpha signaling in developing hippocampus of male rats through estrogen receptor. Hippocampus. 24(12):1570-80.
- Xu X et al. (2015a) Sex-specific effects of long-term exposure to bisphenol-A on anxiety-and depression-like behaviors in adult mice. Chemosphere. 120:258-66.
- Xu X, Gu T, Shen Q. (2015b) Different effects of bisphenol-A on memory behavior and synaptic modification in intact and estrogen-deprived female mice. J Neurochem.132(5):572-82.
- Xue J, Wu Q, Sakthivel S, Pavithran PV, Vasukutty JR, Kannan K. (2015) Urinary levels of endocrine-disrupting chemicals, including bisphenols, bisphenol A diglycidyl ethers, benzophenones, parabens, and triclosan in obese and non-obese Indian children. Environ Res. 137:120-8.
- Yang M, Ryu JH, Jeon R, Kang D, Yoo KY. (2009) Effects of bisphenol A on breast cancer and its risk factors. Arch Toxicol 83:281-5
- Yang L, Luo L, Ji W, Gong C, Wu D, Huang H, Liu Q, Xia B, Hu G, Zhang W, Zhang Q, Liu J, Zhuang Z. (2013) Effect of low dose bisphenol A on the early differentiation of human embryonic stem cells into mammary epithelial cells. Toxicol Lett 218:187-93
- Yang M, Chen M, Wang J, Xu M, Sun J, Ding L, Lv X, Ma Q, Bi Y, Liu R, Hong J, Ning G. (2016) Bisphenol A promotes adiposity and inflammation in a nonmonotonic dose-response way in five-week old male and female C57BL/6J mice fed a low-calorie diet. Endocrinology 157(6):2333-45
- Yeo M *et al.* (2013) Bisphenol A delays the perinatal chloride shift in cortical neurons by epigenetic effects on the Kcc2 promoter. Proc Natl Acad Sci USA. 110(11):4315-20.

- Yokota H *et al.* (2000) Effect of bisphenol A on the early life stage in Japanese medaka (Oryzias latipes). Environmental Toxicology and Chemistry19(7):1925-1930.
- Yoshida M *et al.* (2004) Maternal exposure to low doses of bisphenol a has no effects on development of female reproductive tract and uterine carcinogenesis in Donryu rats. Journal of Reproduction and Development 50, 349-360.
- Yu CJ *et al.* (2015) Pubertal BPA exposure changes central ERa levels in female mice. Environ Toxicol Pharmacol. 40(2):606-14.
- Zaid SS, Othman S, Kassim NM. (2014) Potential protective effect of Tualang honey on BPA-induced ovarian toxicity in prepubertal rat. BMC Complement Altern Med. 14:509.
- Zhang Q et al. (2013) Exposure to bisphenol-A affects fear memory and histone acetylation of the hippocampus in adult mice. Horm Behav. 65(2):106-13.
- Zhou W *et al.* (2008) Effect of bisphenol A on steroid hormone production in rat ovarian theca-interstitial and granulosa cells. Mol. Cell. Endocrinol. 283:12–18

## **References for Part II**

- AFSSA (2008a). Opinion of the French Food Safety Agency regarding bisphenol A in polycarbonate baby bottles likely to be heated in microwave ovens. Request no. 2008-SA-0141, 24 October 2008.
- AFSSA (2008b). Opinion of the French Food Safety Agency regarding the exposure assessment of bisphenol A in water intended for human consumption and possible resulting health risks. Request no. 2008-SA-0141A, 21 November 2008.
- AFSSA (2009). Note from the French Food Safety Agency regarding the publication by Stahlhut et al. (2009) on urinary elimination of bisphenol A in humans. Request no. 2009-SA-0177, 07 July 2009.
- AFSSA (2010a). Opinion of the French Food Safety Agency on the critical analysis of the results of a developmental neurotoxicity study of bisphenol A together with other recently-published data on its toxic effects. Request no. 2009-SA-0270, 29 January 2010.
- AFSSA (2010b). Opinion of the French Food Safety Agency regarding clarification of the AFSSA opinion on bisphenol A issued on 29 January 2010. Request no. 2010-SA-0041, 7 June 2010.
- AFSSA (2010c). Opinion of the French Food Safety Agency regarding exposure to bisphenol in the French population and maximum levels of bisphenol A in foods. Request no. 2010-SA-0040, 2 March 2010.
- AIST (2007). Bisphenol A risk assessment document summary AIST risk assessment document series n°4.
- AIST & RISS (2011). Updated hazard assessment of bisphenol A.
- ANSES (2011). Health effects of bisphenol A Knowledge about the uses of bisphenol A.
- ANSES (2012). Analysis of the most appropriate risk management option for bisphenol A. January 2012
- ANSES (2013a). Substitution of bisphenol A: an inventory of alternatives to BPA, identification of the hazards of potential substitutes for bisphenol A. March 2013
- ANSES (2013b). Risk assessment of Bisphenol A (BPA) on human health. March 2013.
- ANSES (2013c). CLH report. Proposal for harmonised classification and labelling. Bisphenol A. 17/07/2013 Available at <a href="https://echa.europa.eu/documents/10162/36b05a93-3e3c-44b1-bc8d-bff66b4b37ae">https://echa.europa.eu/documents/10162/36b05a93-3e3c-44b1-bc8d-bff66b4b37ae</a> (accessed on 4 November 2016)
- ANSES (2014). Annex XV Restriction Report Proposal for a restriction 4.4'- isopropylidenediphenol (bisphenol A; BPA). May 2014.
- ANSES (2016) Analysis of the most appropriate risk management option for bisphenol A. May 2016
- Canada Gazette (2010). Order amending Schedule I to the Hazardous Products Act (bisphenol A). P.C. 2010-256 March 11, 2010.
- CETIM (2012). Remplacement des polycarbonates. March 2012 (Unpublished)
- Danish EPA (2011). Migration of bisphenol A from cash register receipts and baby

- dummies. Survey of chemical substances in consumer products n°110 2011.
- EC, February 2010. European Union Risk Assessment Report 4,4'- isopropylidenediphenol (bisphenol A) complete risk assessment in one document.
- ECHA (2012). Guidance for monomers and polymers. April 2012
- ECHA (2014). Committee for Risk Assessment (RAC). Opinion proposing harmonised classification and labelling at EU level of Bisphenol A; 4,4'-isopropylidenediphenol. Adopted 14 march 2014. ECHA. Available at <a href="http://echa.europa.eu/documents/10162/777918ff-33b5-46ff-be89-2bdc406d34fa">http://echa.europa.eu/documents/10162/777918ff-33b5-46ff-be89-2bdc406d34fa</a> (accessed on 6 June 2016).
- ECHA (2015). Opinion on an Annex XV dossier proposing restrictions on Bisphenol A. Compiled version prepared by the ECHA Secretariat of RAC's opinion (adopted 5 June 2015) and SEAC's opinion (adopted 4 December 2015).
- EFSA (2016) A statement on the developmental immunotoxicity of bisphenol A (BPA): answer to the question from the Dutch Ministry of Health, Welfare and Sport. Panel on Food Contact Materials, Enzymes, Flavourings and Processing Aids (CEF). EFSA Journal?. Adopted on 14 September 2016;14 (10):4580 <a href="http://onlinelibrary.wiley.com/doi/10.2903/j.efsa.2016.4580/epdf">http://onlinelibrary.wiley.com/doi/10.2903/j.efsa.2016.4580/epdf</a>
- EFSA (2015). Scientific Opinion on the risks to public health related to the presence of bisphenol A (BPA) in foodstuffs. EFSA Journal 2015;13(1):3978.
- EFSA (2011). Agreed joint report of EFSA and ANSES according to Article 30 of the Regulation (EC) No 178/2002 on bisphenol A. Parma, 30 November 2011.
- EFSA (2010). Scientific opinion on bisphenol A: evaluation of a study investigating its neurodevelopmental toxicity, review of recent scientific literature on its toxicity and advice on the Danish risk assessment of bisphenol A. EFSA Journal 2010; 8(9): 1829 [116 pp.].
- EFSA (2008). Toxicokinetics of bisphenol A Scientific opinion of the panel on food additives, flavourings, processing aids and materials in contact with food. EFSA Journal 2008; 759.
- EFSA (2006). Opinion of the scientific panel on food additives, flavourings, processing aids and materials in contact with food on a request from the Commission related to 2,2-bis(4-hydroxyphenyl)propane (Bisphenol A). Question number EFSA-Q-2005-100 adopted on 29 November 2006. The EFSSA Journal (2006) 428, 75p.
- Fache M, Auvergne R, Boutevin B, Caillol S (2015). New vanilin-derived diepoxy monomers for the synthesis of biobased thermosets. European Polymer Journal 67:527-538
- FAO/WHO (2010). Toxicological and health aspects of bisphenol A. Report of Joint FAO/WHO Expert Meeting and report of Stakeholder Meeting on bisphenol A.
- FDA (2014a) Updated Review of Literature and Data on Bisphenol A (CAS RN 80-05-7). Memorandum. Bisphenol A (BPA) Joint Emerging Science Working Group. Food and Drug Administration (USA) June 6, 2014 <a href="http://www.fda.gov/downloads/Food/IngredientsPackagingLabeling/FoodAdditivesIngredients/UCM424071.pdf">http://www.fda.gov/downloads/Food/IngredientsPackagingLabeling/FoodAdditivesIngredients/UCM424071.pdf</a>
- FDA (2014b) Updated safety assessment of Bisphenol A (BPA) for use in food contact applications. Memorandum. Food and Drug Administration (USA) June 17, 2014 <a href="http://www.fda.gov/downloads/NewsEvents/PublicHealthFocus/UCM424266.pdf">http://www.fda.gov/downloads/NewsEvents/PublicHealthFocus/UCM424266.pdf</a>

- FOPH (Federal Office of Public Health) (2016). Bisphenol A.. Factsheet. February 2016. FOPH of the Swiss Confederation. <a href="https://www.bag.admin.ch/dam/bag/fr/dokumente/chem/themen-a-z/factsheet-bisphenol-a.pdf.download.pdf/2016-bpa-factsheet-bisphenol-a.pdf.downl
- Health Canada (2008a). Health risk assessment of bisphenol A from food packaging applications. Bureau of chemical safety, food Directorate, health products and food branch. <a href="http://www.hc-sc.gc.ca/fn-an/securit/packag-emball/bpa/bpa hraers-eng.php">http://www.hc-sc.gc.ca/fn-an/securit/packag-emball/bpa/bpa hraers-eng.php</a>
- Health Canada (2008b). Proposed risk management approach for Phenol, 4,4'-(1-methylethylidene)bis (bisphenol A). https://www.ec.gc.ca/ese-ees/default.asp?lang=En&n=6FA54372-1
- INERIS (2010). Données technico-économiques sur les substances chimiques en France : Le bisphénol A. Rapport d'étude. NO.INERIS-DRC-10-109972-01125C. – October 2010.
- INERIS (2015). Possibilité technique et faisabilité économique de la substitution du tétrabromo bisphenol A. Rapport d'étude. N°DRC-15-142535-00175A
- INFOSAN (2009). Bisphenol A (BPA) Current state of knowledge and future actions by WHO and FAO. INFOSAN information note No. 5/2009 Bisphenol A.
- Janvier M *et al* (2017). Syringaresinol: a renewable and safer alternative to Bisphenol A for epoxy-amine resins. ChemSusChem 10(4):738-746
- KemI (2012). Swedish Chemicals Agency. Bisphenol A in toys and children's products a need for exposure reduction? (in Swedish with English summary). Rapport Nr 6/12. September 2012. <a href="http://www.kemi.se/Documents/Publikationer/Trycksaker/Rapporter/Rapport-6-12-BPA-i-leksaker-och-barnartiklar.pdf">http://www.kemi.se/Documents/Publikationer/Trycksaker/Rapporter/Rapport-6-12-BPA-i-leksaker-och-barnartiklar.pdf</a>
- Liao C. and Kannan K. (2011). Widespread occurrence of bisphenol A in paper and paper products: implications for human exposure. Environmental Science and Technology 45, 9372-9379.
- Menard S *et al.* (2014a) Food intolerance at adulthood after perinatal exposure to the endocrine disruptor bisphenol A. FASEB J.28(11):4893-900.
- Menard S *et al.* (2014b). Perinatal exposure to a low dose of bisphenol A impaired systemic cellular immune response and predisposes young rats to intestinal parasitic infection. PLoS One9(11):e112752.
- NTP-CERHR (2008). Monograph of the potential human reproductive and developmental effects of bisphenol A. National Toxicology Program, Center for the Evaluation of Risks to Human Reproduction, Expert Panel evaluation of bisphenol A.
- Report of Joint FAO/WHO Expert Meeting, 2-5 November 2010 and Report of Stakeholder Meeting on Bisphenol A, 1 November 2010. Toxicological and health aspects of bisphenol A.
- RIVM (2016). Bakker J *et al.* Bisphenol A: Part 2. Recommendations for risk management. RIVM report 2015-0192, 84p. Published on RIVM website <a href="http://www.rivm.nl/Documenten en publicaties/Wetenschappelijk/Rapporten/2016/maart/Bisphenol A Part 2 Recommendations for risk management">http://www.rivm.nl/Documenten en publicaties/Wetenschappelijk/Rapporten/2016/maart/Bisphenol A Part 2 Recommendations for risk management (accessed on 7 June 2016)</a>
- SCENIHR (2015). Opinion on the safety of the use of bisphenol A in medical devices. Scientific Committee on Emerging and Newly Identified Health Risks. Adopted on

- 18 February 2015 http://ec.europa.eu/health/scientific committees/emerging/docs/scenihr o 040.pdf
- SCOEL (2014). Recommendation from the Scientific Committee on Occupational Exposure Limits for Bisphenol-A. SCOEL/SUM/113. June 2014. (accessed on 7 June 2016)
- US EPA (2010a). List of known or expected functional BPA alternatives for use in thermal paper to be evaluated in the DfE alternatives Assessment. US EPA Action plan on BPA.
- US EPA (2010b). Bisphenol A Action Plan. 29 March 2010.
- Von Goetz N, Wormuth M, Scheringer M and Hungerbühler K. (2010). Bisphenol A: how the most relevant exposure sources contribute to total consumer exposure. Risk Analysis 473-487.

## Annex I - *In vitro* data were previously evaluated and quoted in ANSES report 2013 and 2014

In vitro, BPA at concentrations ranging from 100 pM to 1  $\mu$ M promotes adipogenesis in mouse preadipocyte 3T3-L1 cells (Sargis, 2010). The activation of this lipogenesis is mediated by glucocorticoid receptors (GR). BPA increases lipogenesis in differentiating adipocytes and activates the expression of specific adipocytic proteins (adiponectin, transcription factor CCAAT enhancer binding protein  $\alpha$  (C/EBP- $\alpha$ ), a factor induced in the terminal phase of adipogenesis). However, the action of BPA on adiponectin induction shows a bell curve with a visible effect from 10 nM, peaking at 100 mN and disappearing at 1000 nM. An identical dose-response relationship was observed with DEX. It should be noted that in this study, the other compounds under consideration, dicyclohexyl phthalate, tolyfluanid, troglitazone and triphenyltin had lesser effects at the highest concentration of 1  $\mu$ M.

In the studies by Kidani *et al.*, 3T3-L1 cells were exposed to various forms of bisphenol (BP): BPA, BPB, BPE and BPF at concentrations of 0, 20, 40 and 80  $\mu$ M. In a dose-dependent manner, BPA decreased the concentration of cellular adiponectin and was secreted in the extracellular medium (Kidani, 2010). Forms of BPA can be classified as follows according to their ability to reduce adiponectin secretion: BPB > BPA > BPE > BPF. BPA negatively regulates the Phosphatidylinositol 3-Kinase (PI3K)-Akt signalling pathway by reducing Akt and p-Akt expression.

However, the inhibition of adiponectin expression by BPA should be compared with the results obtained by Sargis *et al.* showing a bell-shaped dose-response relationship between BPA and adiponectin (Sargis, 2010). The negative effects on adiponectin expression observed by Kidani *et al.* are therefore not surprising in that they were produced at concentrations greater than 1  $\mu$ M (Kidani, 2010). Thus, BPA may induce adiponectin expression at low doses and suppress it at high doses (which are already very low).

Asahi et al. undertook studies in cultured non-parenchymal hepatocytes, NCTC Clone 1469 cells (Asahi J, 2010). The cells were exposed to BPA at concentrations of 0, 1, 10, 50, 100 and 200  $\mu$ M for 48 hr or at a concentration of 100  $\mu$ M for a period of 120 hours, with an analysis of BPA's effects at various times. After having examined the cytotoxicity of BPA at various concentrations, the studies continued exposing the cells to BPA at the concentration of 100 µM. At this concentration, BPA induced apoptosis which was expressed by DNA fragmentation, phosphatidylserine externalisation on the outer plasma membrane leaflet, an increase in caspase-12, the GRP78/BiP protein (involved in endoplasmic reticulum homeostasis) and transcription factor CHOP (C/EBP homologous protein, a transcription factor involved in stress-induced apoptosis in the endoplasmic reticulum), and a slight decrease in the anti-apoptic protein Bcl-2. These results strongly suggest that the endoplasmic reticulum plays a role in the apoptosis induced by BPA. The effects of BPA are accompanied by oxidative stress, with an increase in reactive oxygen species (ROS) counteracted by antioxidant N-acetylcysteine (N-AC). At the concentration of 100 µM, the effects of BPA do not appear to be mediated by estrogen receptors; the estrogen receptor inhibitors 4-OHT and ICI do not prevent the cytotoxicity of BPA and 4-OHT enhances it (Note: 4-OHT has a partial agonist effect on estrogen receptors).