Douglas Connect Working communities

Integration into risk assessment of open source human omics data from *in vitro* studies

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Introduction

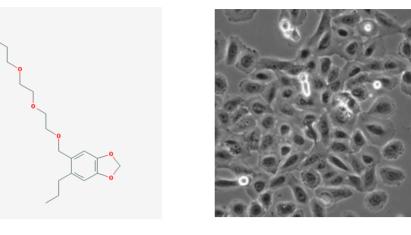
A recent milestone in toxicity research was the emergence of the field of toxicogenomics, resulting from the application of knowledge gained from genomics science into conventional toxicology. This field specifically tackles the complex interactions between toxic effects and the structure and activity of the genome [1, 2]. The question now is if the detection of chronic and systemic toxicity is possible via these alternative testing strategies to reduce and replace animal testing.

As part of the ToxBank project (www.toxbank.net) [3] in the SEURAT-1 program, *ab initio* case studies have been proposed in which the toxicity of four compounds are assessed using available human *in vitro* data only. These compounds are the reference compounds doxorubicin, valproic acid and methotrexate with well-defined modes of actions as well as the test compound piperonyl butoxide (PBO). The latter was chosen as a compound relevant for the cosmetics industry.

Here, an approach using the combination of omics data with information extracted from adverse outcome pathways (AOPs) to identify areas of concern and support an evidence-driven risk assessment is presented with the example PBO (Fig. 1)

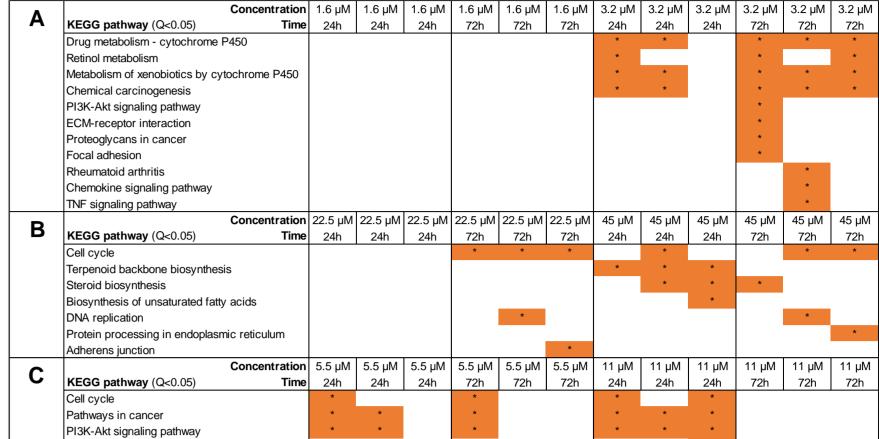
Identification of Relevant Pathways

- The transcriptomics data described was analysed using the program InCroMAP to identify pathways, which are influenced by the treatment of the cells with PBO at different concentrations and time points
- BH FDR correction was applied and fold changes were regarded as significant if the Q-values < 0.05
- Relevant pathways were identified using KEGG pathway enrichment approach (Fig. 2)



PBO (CAS: 51-03-6) Human liver cells

#	Name	Observation 1 [Fo	Observation 2 [Fo	Obse	rvation 3 [Fo	Obs	ervation 4 [Fo	Observation 5 [Fo
	1 DDR1	-0.1484	0.2077	-0.055	52	0.078	31	-0.1476
	2 RFC2	0.2067	0.0576	0.081	2	0.207	74	-0.2214
	3 HSPA6	0.0	0.3789	-0.349	91	0.310	54	-0.5243
	4 PAX8	-0.2768	0.3862	6.885	E-3	-0.09	07	-0.1478
	5 GUCA1A	0.0	0.0	0.0		0.0		0.0
	6 UBA7	0.0593	0.2412	-0.076	58	-0.01	15	-0.0656
	7 THRA	0.0353	-0.1627	-0.213	39	-0.50	3	0.1639
	8 PTPN21	-0.3862	0.0	0.0		0.493	81	0.0
	9 CCL5	0.0	0.0			0.0		0.0
	10 CYP2E1	0.0508	0.3687	0.4542	2	0.07	4	-0.5979
	11 EPHB3	0.0	0.0	0.0		0.0		0.0
	12 ESRRA	0.1552	0.41	0.1881		0.362	28	0.1921
	13 CYP2A6	1.1508	1.258	0.678	1	0.893	39	0.891
	14 GAS6	-0.094	-0.0483	-0.161	17	8.100	iE-3	-0.0735
	15 MMP14	0.1582	0.3125	-0.145	53	0.479	94	-0.2498
	16 TRADD	-0.0159	-0.2934	-0.229	95	0.07	6	-0.1865
	17 AL139022.1	-0.0526	0.1672	-0.205	55	0.059	95	-0.1458
	18 PLD1	0.3473	-0.1168	0.702	8	0.13	4	0.222
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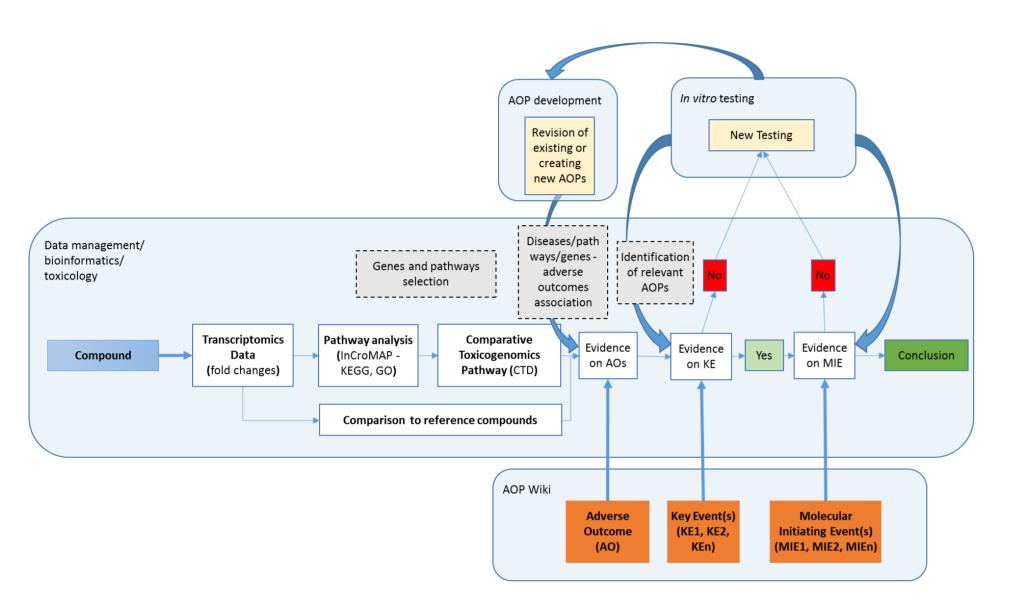


Figure 1. Analysis workflow

Test compound: Piperonyl Butoxide (PBO)

- PBO belongs to a class of chemicals known as the methylenedioxyphenyl compounds and is an insecticide synergist [4]
- PBO was also included in cosmetic products for skin protection [5]
- PBO is classified as a non-genotoxic carcinogen, following in vitro and in vivo investigations [5]
- A PBO metabolite binds to Cytochrome P450 enzymes, thus reducing the ability of the enzymes in breaking down accompanying pesticides
 PBO's effect on Cytochrome P450 enzymes is biphasic it both inhibits and induces enzymatic activity; the inhibition of P450 enzymes occurs rapidly, followed by a slow induction process. The rapid inhibition contributes to PBO's effectiveness as a synergist [7]
 Information and downloadable data on PBO were found in the following databases: diXA, Leadscope, COSMOS, ECHA CHEM, ChEBI, TOXNET, eChemPortal OECD, EPA HHBP, PUBCHEM, CTD, CheLIST, TOXCAST

Figure 2. Data analysis

- The analysis of HepaRG cells confirmed the interaction of PBO with cytochrome P450 but this was not observed in HepG2 and hES-DE-Hep cells
- The pathways identified in HepaRG cells showed concentration- and time-dependent enrichment, as the treatment with the higher concentration showed most significant effects at 72h comparative with 24h
- A similar effect was observed in HepG2 cells but the pathways enriched were different. In the case of of hES-DE-Hep a cell batch-dependent response was seen (Fig. 3)

Correlation between Pathways and

Diseases

- To identify adverse outcomes of PBO, we initially followed a consensus approach using information of all cell lines (Fig. 4)
- The top pathways were selected and analysed further using the services of CTD, which correlate the observed pathways with diseases

FISK-AKI Signaling pairway								
Axon guidance						*		
Prostate cancer		*			*	*		
Colorectal cancer		*				*		
Regulation of actin cytoskeleton	* *	*		*		*		
Adherens junction		*			*	*		
Focal adhesion	* *	*	*	*	*		*	
Prion diseases					*			
Melanoma	*	*		*	*			
ECM-receptor interaction	*	*		*	*		*	
Chronic myeloid leukemia					*			
Glioma		*			*			
Transcriptional misregulation in cancer		*			*			
Proteoglycans in cancer	* *	*		*	*			
Renal cell carcinoma	*	*						
p53 signaling pathway	*	*		*				
Hippo signaling pathway				*				
Viral carcinogenesis	*			*				
HTLV-I infection	*	*		*				
Leukocyte transendothelial migration		*						
Endometrial cancer		*						
DNA replication		*						
Basal cell carcinoma		*						
Hypertrophic cardiomyopathy (HCM)	*	*						
Pancreatic cancer		*						
Dilated cardiomyopathy	*							
Complement and coagulation cascades	*							

Figure 3. Most relevant pathways identified in HepaRG cells (A), HepG2 (B) and hESC DE-Hep cells (C)

		·		
Enriched pathway (Q<0.05)	KEGG ID	HepaRG	HepG2	hESC DE-Hep
ECM-receptor interaction	04512	*		*
Focal adhesion	04510	*		*
PI3K-Akt signaling pathway	04151	*		*
Adherens junction	04520		*	*
Cell cycle	04110		*	*
DNA replication	03030		*	*
Proteoglycans in cancer	05205	*		*

Figure 4. Top pathways enriched at least in two cell lines

Verifying Adverse Effects by Testing for Specific Key Events

 Adverse outcome pathways (AOPs) summarize all available information leading from a molecular initiating event (MIE) to intermediate key events (KE) and finally to the adverse outcome (AO)

Name	Abbreviation	Use		
Data Infrastructure for	diVa	Transcriptomics data on test compound		
Chemical Safety	diXa	PBO [8]		
ArrayExpress Archive of	A	Transcriptomics data on reference		
Functional Genomics Data	ArrayExpress	compounds MTX and VPA [9,10]		
Integrated analysis of Cross-				
platform MicroArray and	InCroMap	Kyoto Encyclopedia of Genes and		
Pathway data		Genomes (KEGG) pathway analysis [11]		
Comparative Toxicogenomics		Pathway-genes-diseases-chemicals		
Database	CTD	associations analysis [12]		
Adverse Outcome Pathway		Verification of adverse effects versus		
Knowledge Base	AOP-KB	specific KE [13]		

Omics Data

- In the case study performed here, an *ab initio* risk assessment is modelled, i.e. it is assumed that no information on the toxicity of the compound is available
- To quickly identify areas of concern, omics data form a good starting point, since they show general adaptations of the cell to the exposure
 Vinken et al. [14] used transcriptomics measurements on three different human liver cell lines, for the development of omics-based *in vitro* carcinogenicity screening assays:

• The applied query resulted in a large number of associated diseases from which the top ranks are given in the table below

Rank	Disease Name	Disease ID	Number of		
			associated genes		
1	Dermatitis, Allergic Contact	MESH:D017449	100		
2	Prostatic Neoplasms	MESH:D011471	49		
3	Breast Neoplasms	MESH:D001943	48		
4	Stomach Neoplasms	MESH:D013274	44		
5	Lung Neoplasms	MESH:D008175	40		
32	Fibrosis	MESH:D005355	15		

- Fibrosis, an adverse effect related to PBO appears on rank 32 of the list of diseases associated with the identified pathways
- Genes of three pathways are associated with this disease. It is also interesting that these genes are mainly related to pathways identified with the HepaRG cells
- A next step included the analysis on separate cell models, showing 21 genes associated with fibrosis in the HepaRG cells (table below), whereas 4 genes were associated with fibrosis in HepG2 cells

Rank	Disease Name	Disease ID	Number of associated genes
4	Drug-Induced Liver Injury	MESH:D056486	50
43	Fibrosis	MESH:D005355	21

 Starting from the adverse outcome (Fig. 5), these relations can now be followed backwards and more specific tests can be performed to verify the occurrence of key events

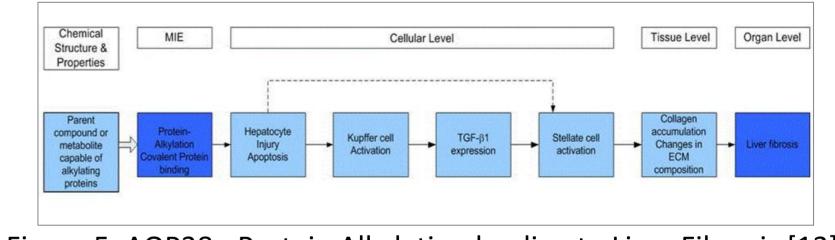


Figure 5. AOP38 - Protein Alkylation leading to Liver Fibrosis [13]

- The transcriptomics data analysis showed that fibrosis could be a potential adverse outcome for exposure to PBO
- The KE preceding the AO is the **collagen accumulation** for which the AOPKB describes three different methods for testing this KE [15]
- The next upstream KE described is **stellate cell activation** for which data does exist (*unpublished data from HeMiBio project*)
- Up-regulation of TGF-β1 expression is the next upstream KE, which can be measured by ELISA [13]. Unfortunately, we could not find any such data for PBO. At the transcription level, in none of the cell systems analyzed here (HepaRG, HepG2 and hES-DE-Hep) is any effect on the TGF-β1 gene expression levels observed. However, the AOP does not list gene expression as a valid test for this KE
- Therefore, we propose as next steps to perform additional tests to confirm or exclude these KEs

Conclusions

- HepaRG (human hepatoma-derived cells),
- HepG2 (hepatocellular carcinoma-derived cell line), and
- hES-DE-Hep (hepatocyte-like cells derived from embryonic stem cells)
- We show here that transcriptomics data is able to identify fibrosis as one potential adverse outcome of treatment with PBO
- HepaRG cells seem to be more appropriate to test for this specific effect
- Using information from AOP-KB, we were then able to verify key events and, in this way, strengthen the evidence for this specific adverse effect
- Additional tests are proposed, especially the TGF-β1 measurements by ELISA to see if the TGF-β1 expression is up-regulated
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